



# The relative importance of fecal and urinary excretion of perfluorooctane sulfonic acid and perfluorooctanoic acid after high exposure – An observational study in Ronneby, Sweden

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## ABSTRACT

**Background:** Many poly- and perfluoroalkyl substances (PFAS) are persistent and have long half-lives in the human body. However, there are limited data on the different routes of elimination. Most pharmacokinetic models assume that the urinary route dominates.

**Objectives:** Our aim was to investigate the relative importance of fecal and urinary elimination for linear perfluorooctane sulfonic acid (L-PFOS), branched PFOS and perfluorooctanoic acid (PFOA), and to estimate volumes of distributions (Vds).

**Methods:** Drinking water highly contaminated with PFAS from firefighting foam was distributed to many households in Ronneby, Sweden, from the 1980s to December 2013. In 2016, PFAS levels were measured in matched serum, feces and urine samples from 147 subjects. Daily urinary and fecal PFAS elimination was estimated through urinary creatinine elimination and dry fecal mass, respectively. Longitudinal serum PFAS elimination rates were used together with fecal and urinary elimination rates to estimate Vds.

**Results:** In 2016, the median serum concentrations were 100 ng/mL for L-PFOS and 10 ng/mL for PFOA. L-PFOS was eliminated primarily through feces, with a median urinary elimination of 91 ng/day and median fecal elimination of 364 ng/day. The branched PFOS had, similarly, a primarily fecal elimination. In contrast, PFOA had a slightly higher urinary elimination, with median urinary elimination of 26 ng/day and fecal elimination of 15 ng/day. Median Vds were estimated at 93 mL/kg for PFOS and 74 mL/kg for PFOA.

**Conclusion:** Fecal elimination was shown to be an important route for PFOS and PFOA elimination. Pharmacokinetic models need to take fecal elimination into consideration.

## 1. Introduction

Per- and polyfluoroalkyl substances (PFAS) are synthetic chemicals used in aqueous firefighting foams, clothing, various consumer products and industrial applications. PFAS are often called “forever chemicals” because of their high persistence in nature and, for some substances, in humans. Apparent half-lives of perfluorinated PFAS in humans range from months to several years (Li et al., 2018, 2022; Xu et al., 2020). The half-lives depend on molecular structure, such as chain length and functional groups: for example, compounds with shorter chain lengths generally have a shorter half-life (Xu et al., 2020). The most epidemiologically and toxicologically studied PFAS substances are perfluorooctanoic acid (PFOA) and perfluorooctane sulfonic acid (PFOS),

partly due to their wide-spread use and their long half-lives in humans.

Urinary elimination has been considered as the primary excretion route in models of half-lives in humans and animals using physiologically based pharmacokinetic (PBPK) models, and is the only route included in some models (Andersen et al., 2006; Loccisano et al., 2011). Although Worley et al. (2017) included fecal elimination as a separate route, they regarded it as insignificant and noted that it was only included for the sake of completeness. However, rat models have shown substantial biliary (Vanden Heuvel, Kuslikis, Van Rafelghem and Peterson, 1991) and fecal elimination (Cui et al., 2010; Vanden Heuvel et al., 1991). In one rodent study, the urinary and fecal elimination of PFOA and PFOS were comparable, albeit with a slightly higher urinary elimination (Cui et al., 2010).

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Strong support for significant fecal elimination in humans was provided by three intervention studies exploring treatment with the bile-sequestering medication cholestyramine, which is thought to block intestinal PFAS reuptake (Genuis et al., 2010; Genuis et al., 2013; Möller et al., 2024). The first study, a case-report, found a substantial decline of serum PFOA, perfluorohexane sulfonic acid (PFHxS) and PFOS levels in one individual after 12 and 20 weeks of cholestyramine intervention (Genuis et al., 2010). The second study, a case-series, found increased fecal elimination of the same compounds in eight individuals after at least 5 days of cholestyramine intervention (Genuis et al., 2013). Lastly, a recent Danish study in 45 subjects confirmed that PFAS serum levels could be substantially lowered with cholestyramine, with 63 % lowering of PFOS serum levels in a twelve-weeks cholestyramine intervention compared to 3 % of twelve weeks without intervention (Möller et al., 2024). The other PFAS investigated, PFOA, perfluorononanoic acid (PFNA), perfluorodecanoic acid (PFDA) and PFHxS, were also lowered, but to lesser degrees.

A pharmacokinetic parameter with limited PFAS data is the volume of distribution (Vd), which can be used to estimate other key pharmacokinetic parameters such as clearance or body burden from serum or drinking water concentrations (Fan and de Lannoy, 2014). Vds are theoretical volumes that assess the extent of compound distribution after absorption, i.e. if the compound is confined to plasma or if it freely distributes to other tissues, and is determined by the compound's chemical properties (Fan and de Lannoy, 2014). For PFAS, different strategies have been used to estimate Vds. Buser et al. extrapolated Vds from experimental studies in monkeys, yielding approximations of 200 mL/kg body weight for PFOA and PFOS (Buser et al., 2021). Thompson et al. used pharmacokinetic modelling with assumptions regarding daily intake of contaminated drinking water, yielding 230 mL/kg body weight for PFOA and 170 mL/kg for PFOS (Thompson et al., 2010). A recent study based on studies which reported both drinking water concentrations and serum concentrations, used Bayesian modelling to estimate Vds of 430 mL/kg for PFOA and 336 mL/kg for PFOS (Chiu et al., 2022). In contrast, considerably lower Vds were found by Abraham et al., (2024), where oral administration of  $^{13}\text{C}$ -labeled PFAS in a single volunteer resulted in Vds of 121 mL/kg for  $^{13}\text{C}$ -PFOA and 152 mL/kg for  $^{13}\text{C}$ -PFOS (Abraham et al., 2024).

To our knowledge, previous studies have not clarified the relative importance of the two major elimination routes of PFAS in humans, urinary and fecal elimination. Likewise, published Vds estimates either required several assumptions, or were only measured in one individual. Therefore, the present study aimed to investigate the roles of urinary and fecal elimination of PFOA and PFOS and to explore the Vds for these compounds.

## 2. Methods

### 2.1. Study design

This observational study is a cross-sectional comparison of PFAS levels in serum, fecal and urine spot samples collected in 2016 from study participants in the follow-up of the Ronneby Biomarker Cohort, 3 years after end of exposure to contaminated drinking water (Xu et al., 2021). In a subset of the study population, we used longitudinal data from repeated serum samplings to estimate elimination rates and Vds. All participants provided informed consent. The study was approved by the Swedish Ethical Review Authority (Lund, 2014/267; Lund, 2015/902 and Lund, 2016/474).

### 2.2. Study setting and study participants

PFAS-containing firefighting foam had been used for fire-extinguishing exercises at the local military air base in Ronneby, southeastern Sweden, since the 1980s. In 2013, water concentrations surpassing 10,000 ng/L of a sum of 8 PFAS (mainly PFHxS and PFOS)

were found in the drinking water of one of the two waterworks in Ronneby (Xu et al., 2021), far above the recent Swedish drinking water maximum limits of 4 ng/L for the sum of four PFAS (PFOA, PFNA, PFHxS and PFOS) (Livsmedelsverket, 2022). The contaminated waterworks was closed in December 2013, and water was instead provided from the uncontaminated waterworks to all inhabitants in Ronneby. Thus, only background exposure remained from 2014 and onwards, which is negligible in comparison to the previous, very high, daily drinking water PFAS exposure.

Blood sampling for exposure assessment was offered to the inhabitants beginning 6 months after the end of exposure from the contaminated water source. A total of 3507 participants provided serum samples (Xu et al., 2021). From this group, 114 volunteers were further recruited to participate in a longitudinal half-life panel study, with serum sampled repeatedly over time, enabling elimination rate and half-life estimations (Li et al., 2018). In addition, all participants were offered reexamination of their PFAS levels in 2016 (Li et al., 2018). In total, 283 individuals provided serum in 2016 and some also provided urine and fecal samples.

This study focuses on 147 individuals aged 16 to 85 who provided serum, urine, and fecal samples (103 from the reexamination group and 44 from the longitudinal half-life panel) (Fig. 1).

PFOA = perfluorooctanoic acid, LOD = limit of detection, Vd = volume of distribution.

For PFOA, 4 individuals were excluded from Vd calculations and 57 from the serum, fecal and urine comparisons due to LOD at 10.8 ng/g dry weight for fecal concentration. Four blood donors were also excluded from Vd calculations.

### 2.3. Biosampling

Blood samples were drawn in two 5 mL Becton Dickinson vacutainer

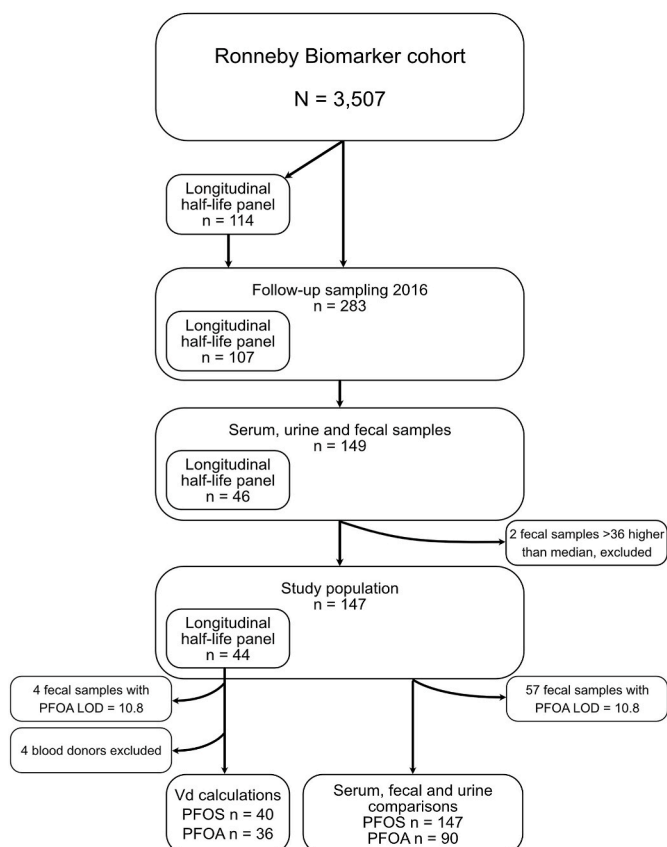


Fig. 1. Flow chart on inclusion of participants.

tubes without gel (BD, Belliver Industrial Estate, Plymouth, UK) and left at room temperature for 30 min to coagulate before centrifugation at 1500 g. Aliquots of serum were transferred to 2 mL cryotubes (Sarstedt, Nümbrecht, Germany) and stored at  $-80^{\circ}\text{C}$  at the Division of Occupational and Environmental Medicine in Lund for further analysis.

Morning urine was collected at home in 13-mL polypropylene screw cap tubes (Sarstedt, Nümbrecht, Germany). A clean paper cup and a tube were provided. The urine samples were frozen at  $-20^{\circ}\text{C}$  and transported frozen to the Division of Occupational and Environmental Medicine, Lund University for analysis and continued storage at  $-20^{\circ}\text{C}$ .

Feces were collected at home in plastic tubes with built-in spatulas for collection of a small amount of feces. The tube was frozen at  $-20^{\circ}\text{C}$  and then transported frozen to the Man-Technology-Environment Research Center, Örebro University.

## 2.4. PFAS measurements

The PFAS measured in the different media (serum, urine, and feces) are given in [Supplementary Table 1](#).

### 2.4.1. Serum and urine

Quantitative analysis of PFAS in serum and urine samples, described in detail previously ([Xu et al., 2020](#)), was performed using liquid chromatography–triple quadrupole linear ion trap mass spectrometry (LC/MS/MS; QTRAP 6500+; AB Sciex, Framingham, MA, USA). Briefly, 25  $\mu\text{L}$  of serum or 50  $\mu\text{L}$  of urine was mixed with isotopically labeled internal extraction standards (MPFAC-C-ES, Wellington Laboratories, Guelph, Canada) and then precipitated using methanol and acetonitrile by vigorous shaking for 30 min and centrifuged prior to analysis. Aliquots of 4  $\mu\text{L}$  of the serum supernatant or 20  $\mu\text{L}$  of urine supernatant were injected onto the analytical column Acquity UPLC® BEH C18 (1.7  $\mu\text{m}$ , 2.1 mm i.d.  $\times$  100 mm, Waters Corporation, Milford, USA). All analyzed sample batches included a standard curve, four chemical blanks and three quality control (QC) samples ([Xu et al., 2020](#)).

Nineteen PFAS compounds were analyzed in the longitudinal half-life cohort ([Li et al., 2022](#)) and fourteen compounds in the re-examination group ([Table S1](#)). In urine, only 8 compounds were analyzed, as the other 11 were below the limit of detection (LOD). These 8 measured were PFOA, perfluoropentane sulfonic acid (PFPeS), PFHxS, perfluoroheptane sulfonic acid (PFHpS), linear PFOS (L-PFOS), perfluoro-1-methylheptane sulfonic acid (1m-PFOS), perfluoro-2/6-methylheptane sulfonic acid (2/6m-PFOS) and perfluoro-3/4/5-methylheptane sulfonic acid (3/4/5m-PFOS). Because 2m-PFOS and 6m-PFOS could not be analytically separated, the sum of the levels is reported as 2/6m-PFOS. Likewise, 3/4/5m-PFOS values represent the sum of 3m-PFOS, 4m-PFOS and 5m-PFOS levels. Chromatogram for the measured PFOS isomers is presented in [Supplementary Fig. S1](#).

The LODs were determined as the concentrations corresponding to the average plus three times the standard deviation of the concentrations in chemical blank samples. Between-run precision was expressed as the coefficient of variation in the QC samples. Urine creatinine concentrations were determined using an enzymatic method ([Mazzachi et al., 2000](#)) and were used to adjust for urine dilution by dividing the urinary PFAS concentration by the urinary creatinine concentration. The Division of Occupational and Environmental Medicine laboratory in Lund participates in the interlaboratory comparison program G-EQUAS (University of Erlangen-Nürnberg) and is approved by the European Human Biomonitoring Initiative (HBM4EU) project for analyses of PFAS in serum.

Previously, we have investigated the influence of fasting versus non-fasting sampling, morning versus afternoon sampling, and a preceding heavy meal, or vigorous physical activity, without finding any significant differences in serum concentrations (results not published).

### 2.4.2. Feces

Fecal samples were freeze-dried and kept at  $-20^{\circ}\text{C}$ . A small portion (0.2 g) of each sample was mixed with isotopically labeled extraction standards for PFOA, PFNA, PFHxS and L-PFOS (from Wellington Laboratories, Guelph, Canada) and was then treated with alkaline methanol (2 mM sodium hydroxide in methanol) before being extracted three times with 2 mL methanol. Hydrochloric acid was added to the combined extracts (6 mL) to neutralize the previously added sodium hydroxide. Extracts were then diluted with ultra-pure water and purified in two steps, based on the method described in [Persson et al., \(2013\); Persson et al. \(2013\)](#). The first step was a weak anion-exchange solid-phase extraction (WAX, Waters Corporation, Milford, USA) that captured anionic and neutral PFAS, followed by a second step with dispersive carbon (Supelclean ENVI-Carb, Supelco) that typically removes planar aromatic or hexagonal ring-shaped molecules. Extracts were filtered through 0.2  $\mu\text{m}$  hydrophilic polypropylene filters (GHP, from PALL Corporation, NY, USA) and evaporated to a final volume of 200  $\mu\text{L}$ . A new set of isotopically labeled injection standards (PFOA, PFNA, PFHxS, PFOS; Wellington Laboratories, Guelph, Canada) and 300  $\mu\text{L}$  2 mM ammonium acetate in water were added before injection into the UPLC MS/MS system described below.

Instrumental analyses were performed on an UPLC MS/MS system (ACQUITY coupled to Xevo TQ-S, Waters Corporation, Milford, USA) in electrospray ionization mode (ESI) by injecting 10  $\mu\text{L}$  of sample onto a UPLC BEH C18 (2.1  $\times$  100 mm, 1.7  $\mu\text{m}$ ) analytical column. Procedural blanks (chemical blanks) and one QC sample were included in each extraction batch, and the LOD was calculated as the mean concentration in blank samples plus three times the standard deviation.

Samples were analyzed in eleven batches and method precision was evaluated by multiple measurements of a homogenized pool from several study samples (QC sample). Precision, expressed as within-day method reproducibility of the in-house quality control sample, was 4–8 % for PFOA, PFHxS, and PFOS, and slightly higher variation of 31 % for PFNA due to concentration close to the detection limit. Between-day reproducibility for the method over a period of several months ( $n = 11$ ) was 19 % for PFOS, 25 % for PFHxS, 67 % for PFOA and 100 % for PFNA. Accuracy could only be assessed from recoveries of spiked samples and was >75 % for PFOA, PFHxS, PFOS and >50 % for PFNA.

PFOA, PFNA, PFHxS, L-PFOS, 2/6m-PFOS, and 3/4/5m-PFOS were analyzed in feces. Chromatogram for the measured PFOS isomers is presented in [Supplementary Fig. S2](#). Dimethyl-PFOS and 1m-PFOS were occasionally detected but were not further investigated because the QC variabilities were too high (95 % and 190 %, respectively).

### 2.4.3. Valid serum/urine/fecal triplets

The PFAS in measurable concentrations in all media (serum, urine and feces) were PFOA, PFHxS, L-PFOS, 2/6m-PFOS and 3/4/5m-PFOS ([Table S1](#)). However, the calculated Vd for PFHxS indicated an underestimation of fecal concentration, which could not be explained by quality control and verification analyses. Therefore, PFHxS was omitted from further analysis. In summary, this study includes PFOA, L-PFOS, 2/6m-PFOS and 3/4/5m-PFOS.

Two individuals had extremely high fecal PFAS concentrations (without corresponding elevation in serum or urine concentrations), with L-PFOS values of 736 and 465 ng/g dry weight, respectively. These concentrations correspond to 57 and 36 times the L-PFOS median value. Thus, they were considered as extreme outliers and excluded from all descriptions and analyses.

The LODs and the number of samples < LOD are presented in [Supplementary Table 2](#). PFAS values below LOD were included in all analyses as LOD/2. For L-PFOS, there were different LODs in the batches LODs – one at 0.27 and one at 1.0. In the case of 1.0, we instead used 0.27/2. An unknown source of contamination of PFOA resulted in elevated LOD at 10.8 ng/g for 57 individuals; the source of the contamination could not be traced. These data on fecal PFOA concentrations were therefore excluded, but the 57 individuals still provided

data for linear and branched PFOS.

In summary, matched sets of serum, urine and fecal concentrations were available for 147 individuals for L-PFOS, 2/6m-PFOS and 3/4/5m-PFOS, and for 90 individuals for PFOA (Fig. 1). However, urine results for PFPeS, PFHxS and PFHpS, as well as fecal results for PFNA, were included in the supplementary.

## 2.5. Covariate information

Data on background covariates such as smoking behavior, pregnancy history, and residential and workplace addresses were obtained from questionnaires. Body mass index (BMI, kg/m<sup>2</sup>) was calculated from height and weight. In the longitudinal half-life panel, weight and height were self-reported, but scales were available during sampling. Height and weight were measured by study personnel in the resampling group. Since the weight and height were two years outdated for the longitudinal half-life panel, current height and weight for three adolescent individuals aged 16–17 at sampling in 2016 were estimated through Swedish growth trajectory curves using their old, self-reported height and weight from 2014. This was done to avoid erroneous BMI estimations for these growing individuals.

Serum creatinine, using an enzymatic method (Mazzachi et al., 2000), and cystatin C, measured on a Cobas 8000 analyzer (Roche Diagnostics, Basel, Switzerland), were measured in the same serum samples as PFAS. The estimated glomerular filtration rate (eGFR) was calculated using the CKD-epi 2021 equation based on creatinine and cystatin C (Inker et al., 2021).

Levels of calprotectin, a clinical marker of gut inflammation (Walsham and Sherwood, 2016), and zonulin, a marker of gut permeability (Fasano, 2012), were measured in feces. Fecal water content was calculated as the percent fecal weight loss due to freeze-drying (wet weight minus dry weight, divided by the wet weight).

Age (in years), BMI, and percent fecal water content were treated as continuous variables. Calprotectin levels were divided into above vs. below the clinical cut-off at 50 mg/kg (Konikoff and Denson, 2006); zonulin levels were divided into above vs. below their median levels; eGFR levels were divided into above vs. below 60 mL/min/1.73 m<sup>2</sup> ("KDIGO, 2024 Clinical Practice Guideline for the Evaluation and Management of Chronic Kidney Disease," 2024).

## 2.6. Estimation of total daily elimination and clearance

### 2.6.1. Estimation of daily urinary PFAS elimination

The daily urinary PFAS elimination for an individual was estimated using the following equation:

$$E(u)_{PFAS} = C(u)_{PFAS} \cdot E_{cr} \quad \text{Eq 1}$$

where  $E(u)_{PFAS}$  was the estimated daily urinary PFAS elimination in ng/day,  $C(u)_{PFAS}$  was the measured creatinine-adjusted urine concentration of PFAS in ng/g creatinine and  $E_{cr}$  was the estimated daily creatinine elimination in g/day.

$E_{cr}$  was estimated using the following equation by Forni Ognà et al. (2015) (Forni Ognà et al., 2015):

$$E_{cr_{women}} = \frac{(218.45 - 2.33 \cdot BMI + 0.66 \cdot age - 0.17 \cdot age^2)}{8.84 \cdot 1000} \cdot weight \quad \text{Eq 2a}$$

$$E_{cr_{men}} = \frac{(266.16 - 2.33 \cdot BMI + 0.66 \cdot age - 0.17 \cdot age^2)}{8.84 \cdot 1000} \cdot weight \quad \text{Eq 2b}$$

### 2.6.2. Estimation of daily fecal PFAS elimination

The amount of daily fecal PFAS elimination was estimated by using the following equation:

$$E(f)_{PFAS} = C(f)_{PFAS} \cdot \frac{29 \cdot body\ weight}{median\ body\ weight} \quad \text{Eq 3}$$

where  $E(f)_{PFAS}$  was the daily fecal elimination in ng/day and  $C(f)_{PFAS}$  was the measured fecal concentration in ng/g dry weight. Published research indicates an average for daily total feces amount of 29 g dry weight, and that this amount varies depending on total food intake, body weight and the type of diet (Rose et al., 2015). We have no information on food intake or types of diet, but we adjusted for body weight assuming that the average of 29 g applies to the median body weight of 76 kg in our study population.

### 2.6.3. Estimation of renal and fecal clearance

Route specific clearance can be estimated by dividing elimination per unit of time by the serum concentration (Fan and de Lannoy, 2014). Therefore, the renal and fecal clearance of PFAS were estimated by the following equations:

$$CL(u)_{PFAS} = E(u)_{PFAS} / C(s)_{PFAS} \quad \text{Eq 4a}$$

$$CL(f)_{PFAS} = E(f)_{PFAS} / C(s)_{PFAS} \quad \text{Eq 4b}$$

where  $CL(u)$  was renal and  $CL(f)$  was fecal clearance (mL/day),  $E_{PFAS}$  was the eliminated amount per day (ng/day) through urine (u) or feces (f), and  $C(s)_{PFAS}$  was the serum PFAS concentrations.

Clearance per kg body weight (mL/day/kg) was also calculated and presented.

## 2.7. Estimation of volume of distribution

Conventionally,  $V_d$  was estimated through the following equation (Fan and de Lannoy, 2014):

$$V[d](t) = A(t) / C(t) \quad \text{Eq 5}$$

where  $A(t)$  was the amount in the body (i.e. body burden, from now on abbreviated as BB) and  $C(t)$  was concentration in serum (or plasma).

Conventionally, one dose ( $A$ ) is given intravenously or orally, and serum concentration ( $C$ ) is measured after the distribution phase. However, our study population was continuously exposed to PFAS through drinking water over a long period, with coincident elimination through urine and feces. Therefore, individual total PFAS amounts/body burdens were impossible to measure. Instead, we estimated the change of body burden over a day through total daily elimination:

$$\Delta BB = F_{el} + U_{el} + B_{el} \quad \text{Eq 6}$$

where  $\Delta BB$  was the change of amount (ng) of PFAS in the body over a day,  $F_{el}$  was the daily total amount of PFAS eliminated through feces,  $U_{el}$  through urine and  $B_{el}$  through blood losses.

We assumed no ongoing PFAS exposure, except for background exposure, which we consider negligible in comparison to the remaining high serum PFAS levels from earlier drinking water exposure.

Likewise, change of serum concentration (ng/mL) over a day was estimated through the following:

$$C_t = C_0 \cdot e^{-k \cdot t} \quad \text{Eq 7}$$

$$\frac{dC_t}{dt} = C_0 \cdot -k \cdot e^{-k \cdot t} \quad \text{Eq 8}$$

where  $k$  was the serum elimination constant per day, estimated from several serum concentrations.

By setting  $dt$  in equation (8) as one day and  $C_0$  to the same timepoint as the samples were provided (i.e. in 2016), equations (6) and (8) were combined into equation (5), to estimate  $V_d$ :

$$V_d = \Delta BB / \Delta C = - (F_{el} + U_{el} + B_{el}) / (C_0 \cdot -k \cdot e^{-k}) \quad \text{Eq 9}$$

### 2.7.1. Estimations of blood loss

Based on previous studies, two sources of blood loss needed to be



considered – menstruation (Wong et al., 2014) and blood donations (Gasiorowski et al., 2022; Genuis et al., 2014). For menstruation, women <51 years old were classified as menstruating, except for three individuals currently using hormonal contraception in the form of progestogen-only pills (as oral contraception has been correlated with higher PFAS levels, possibly through impaired menstruation (Rush et al., 2018)). The remaining eight women were assumed to have a monthly blood loss of 30 mL, based the median blood loss described by Hallberg et al. (Hallberg et al., 1966), corresponding to 1 mL per day of blood and, as serum volume is assumed to be half the blood volume, 0.5 mL of serum per day.

Four individuals reported being blood donors. If they were regular blood donors and donated 450 mL twice yearly (four times is the maximum allowed times in Sweden), that would correspond to an approximate net average loss of 1.25 mL per day. If blood was donated during the period of repeated measurements for estimating half-life, then the elimination constant (k) would have been affected. If blood was donated just before the sampling for the serum and feces, then that specific serum PFAS measurement would have been affected. As there was no information on the frequency and timing of blood donations, these four individuals were excluded from the main analysis.

Therefore, the individual PFAS losses due to menstruation were estimated as

$$B_{el} = \frac{d \text{ [mL/day]} \cdot C \text{ [ng/mL]}}{2}; \quad \text{Eq 10}$$

where  $d = 1 \text{ mL/day}$  if menstruating and  $C$  was the individual serum concentration (ng/mL) of PFAS. In sensitivity analysis where blood donation was adjusted instead of excluded for,  $d$  included  $1.25 \text{ mL/day}$  for blood donors.

### 2.7.2. Elimination rate (k)

Multiple serum measurements are needed to calculate  $k$ . These data were only available for the longitudinal half-life panel ( $n = 44$ ), thus,  $V_d$  calculations were only calculated in these individuals, excluding the 4 who reported blood donating. The  $k$  values and the method behind calculating them have been reported previously (Li et al., 2018, 2022). In brief,  $k$  was estimated by linear mixed models based on ten repeated serum measures, sampled after the end of high drinking water exposure, and adjusted for ongoing exposure by subtracting background levels.

### 2.7.3. Sensitivity analyses

The following sensitivity analyses in relation to the estimates of blood loss were carried out.

- 1) Every subject included ( $n = 44$ ), but no adjustments for blood loss
- 2) Every subject included ( $n = 44$ ), but with adjustments for blood loss
- 3) Both blood donors and menstruating women excluded ( $n = 32$ )
- 4) For the menstruating women ( $n = 9$ ) and for the blood donors ( $n = 4$ ), values of  $V_d$  were estimated with and without the correction for estimated blood loss and compared to the non-blood-loss subjects ( $n = 32$ )
- 5) mean and median values of urinary & fecal elimination, serum concentration and  $k$  ( $n = 40$ ) were used to calculate  $V_d$ s, instead of individual values
- 6) instead of calculating individual daily creatinine production through the equation by Forni Ognà et al. (2015) (which was used to estimate daily urinary PFAS elimination), we assumed daily creatinine production to be the set values of  $1.26 \text{ g creatinine/day}$  for women and  $1.94 \text{ g creatinine/day}$  for men, extrapolated from a study of 60 healthy adults that provided 24h urine (Sallsten and Barregard, 2021).

## 2.8. Statistical methods

Arithmetic means, medians and distributions were presented for fecal and urinary daily elimination, expressed as measured concentrations, amount eliminated per day (ng/day, “elimination rate”), fecal and renal clearance (mL/day), and clearance per body weight (mL/day/kg). The relative importance of the fecal elimination rate was investigated in every individual using a ratio between fecal elimination per day and urinary elimination per day, as well as a percent of the sum of both elimination rates.

The amounts of PFAS eliminated daily in relation to serum concentration were visualized with scatter plots and Pearson correlations. Pearson correlations were also used to compare fecal/urinary ratios between PFAS compounds.

Univariate linear regression models were used to explore potential predictors of fecal and renal PFAS clearance and the fecal/urinary ratio. Because the distributions were skewed, the dependent variable for each model was ln-transformed fecal or urinary PFAS elimination/kg body weight/serum PFAS concentration/day; or ln-transformed fecal/urinary PFAS elimination ratio. For fecal clearance, the following covariates were considered in the model: age, BMI, sex, fecal zonulin, fecal calprotectin and fecal water content. For renal clearance, age, BMI, sex and eGFR were considered. For the ratio, all above independent variables were investigated, as well as fecal and renal clearance. It should be noted that eGFR equation includes age and sex, and that the estimation of daily creatinine production includes age, BMI and sex.

All data analyses were performed in SAS 9.4 (SAS Institute Inc.). Fig. 3 was created in R (RStudio version 2024.4.1.748) using the package ggplot2 version 3.4.2.

## 3. Results

The background characteristics of the study population are presented in Table 1. The 57 individuals excluded from the PFOA elimination

**Table 1**

Description of the study participants with serum, urinary and fecal PFAS measurements.

Parameter	Whole group	PFOA
Sample size	147	90
Female sex n (%)	96 (65)	60 (66)
Age (years) Median (P5, P95)	51 (23, 70)	52 (21, 79)
Weight (kg) Median (P5, P95)	77 (58, 103)	75 (54, 100)
BMI (kg/m <sup>2</sup> ) Median (P5, P95)	26 (21, 35)	26 (21, 35)
Estimated creatinine elimination <sup>a</sup> (g/day) Median (P5, P95)	1.3 (0.9, 2.1)	1.2 (0.9, 2.1)
eGFR <sup>b</sup> (mL/min/1.73 m <sup>2</sup> ) Median (P5, P95)	90 (60, 120)	88 (48, 119)
Fecal zonulin (ng/mL) Median (P5, P95)	113 (37, 317)	113 (33, 317)
Fecal calprotectin (mg/kg) Median (P5, P95)	51 (8, 237)	43 (3, 260)
Fecal water content (%) Median (P5, P95)	73 (61, 84)	72 (61, 84)

There are two different PFAS columns because results for the different PFAS have different amounts of data: the samples of all 147 participants were used for L-PFOS, 2/6m-PFOS and 3/4/5m-PFOS; for PFOA, 57 out of the 147 samples were excluded due to a fecal LOD at  $10.8 \text{ ng/g dry weight}$ .

eGFR = estimated glomerular filtration rate, LOD = limit of detection, PFAS = per- and polyfluoroalkyl substances, PFOA = perfluorooctanoic acid, L-PFOS = linear perfluorooctane sulfonic acid, 2/6m-PFOS = branched, sum of perfluoro-2/6-methylheptane sulfonic acid, 3/4/5m-PFOS = branched, sum of perfluoro-3/4/5-methylheptane sulfonic acid, P5 = fifth percentile, P95 = ninety-fifth percentile.

<sup>a</sup> Estimated creatinine elimination was calculated by the formula by Forni Ognà et al., (2015).

<sup>b</sup> Estimated glomerular filtration rate (eGFR) was calculated by the CKD-epi 2021 formula based on creatinine and cystatin c.

calculations were slightly younger (median age of 47 years vs 52 years) than the 90 individuals included, but otherwise similar.

The median serum concentration was highest for L-PFOS (100 ng/mL), followed by 3/4/5m-PFOS (46 ng/mL), 2/6m-PFOS (28 ng/mL) and PFOA (10 ng/mL) (Table 2). The distributions of serum concentrations were positively skewed, with some individuals having extremely high serum concentrations. The urinary concentrations were about 1000 times lower than the serum concentrations (Table 2).

The estimated daily urinary and fecal elimination rates were highest for L-PFOS at median 91 and 364 ng/day, respectively, followed by 3/4/5m-PFOS at median 39 and 150 ng/day, respectively (Table 3). PFOA had lower median urinary elimination rates at 26 ng/day, and even lower fecal elimination at 15 ng/day. However, since the PFOS isomers were at much higher serum concentrations than PFOA (Table 2), renal and fecal clearance were used to more accurately compare PFOS and PFOA elimination patterns. 2/6m-PFOS had the highest median fecal clearance at 3.7 mL/day, followed by L-PFOS, 3/4/5m-PFOS and PFOA (Table 3). Conversely, PFOA had the highest renal clearance at 3.6 mL/day, followed by the PFOS isomers (Table 3). In other words, the PFOS isomers had higher fecal clearance while PFOA had higher renal clearance. The correlations between serum levels and elimination rates were positive for all PFAS, with clear linear associations (Fig. 2) and Pearson correlation values of 0.35–0.83 (Table S3).

The 57 individuals whose PFOA fecal results were excluded had marginally lower concentrations across all studied compounds in serum (e.g., PFOA median 8 ng/mL vs 10 ng/mL), urine (e.g., PFOA median 0.018 ng/mL vs 0.022 ng/mL) and feces (e.g., L-PFOS 9.6 ng/g vs 14.0 ng/g) compared to the whole study population (n = 147). However, their renal and fecal clearance were similar to the whole study population (Table S4). In summary, the relations between urinary, fecal and serum levels in the excluded subgroup were similar to findings in the whole study population.

The urine concentrations, urine elimination rates and renal clearances of PFPeS, PFHpS and PFHxS, as well as the corresponding fecal parameters for PFNA are presented in Table S5, but comparisons between the routes were not possible because the compounds were not estimated in the other medium (i.e. the first three were not quantified in feces and PFNA were not quantified in urine). The urinary concentrations and corresponding renal clearance of 1m-PFOS are not presented, as 91 % of urine samples were below LOD (Table S2).

By comparing the ratio between fecal and urinary elimination, as well as the percentage of elimination through feces, we found in our study population that the PFOS isomers were predominantly eliminated through feces and PFOA eliminated through urine (Table 3). This was illustrated by the ratio between fecal and urinary elimination, with a median ratio of 5.67 for 2/6m-PFOS and 0.56 for PFOA (Table 3). Additionally, the median fecal percentage of total elimination was 36 % for PFOA, compared to 85 % for 2/6m-PFOS (Table 3).

However, we found large interindividual variation in elimination profiles (Fig. 3). For the PFOS isomers, most individuals showed more fecal than urinary elimination, but a few showed the opposite pattern. Conversely for PFOA, most individuals showed more urinary than fecal excretion, but there were exceptions (Fig. 3). Individuals with a certain elimination profile (e.g. predominantly fecal elimination) for one compound were more likely to have a similar profile for the other compounds, as Pearson correlations showed moderate to high correlations between fecal/urinary ratios in different PFAS compounds (r between 0.40 and 0.74) (Table S6).

Even though there was interindividual variation in elimination profiles, the univariate linear regression models showed few associations between covariates and fecal clearance, renal clearance and the fecal/renal ratio (Table S7). The univariate linear regression models showed a negative association between age and daily fecal clearance, with between −0.3 % and −1.13 % lower fecal elimination per year lived. Consequently, the ratio between fecal and renal clearance was also negatively associated with age. For the fecal/urinary ratio, both

**Table 2**  
Concentrations of PFAS in serum, urine, and feces in 147 study participants.

PFAS	Serum (ng/mL)						Urine (ng/mL)						Feces (ng/g dry weight)					
	Mean	Median	Min	Max	P10	P90	Mean	Median	Min	Max	P10	P90	Mean	Median	Min	Max	P10	P90
	12	10	0.68	46	3	24	0.024	0.021	<0.01	0.098	0.005	0.045	0.6 <sup>a</sup>	0.5 <sup>a</sup>	<0.32 <sup>a</sup>	2.1 <sup>a</sup>	<0.32 <sup>a</sup>	1.1 <sup>a</sup>
PFOA	120	100	0.38	422	33	234	0.086	0.062	<0.01	0.35	0.015	0.19	15.3	12.9	<0.27	65.1	3.4	29
L-PFOS	36	28	0.33	122	7	72	0.017	<0.02	<0.02	0.072	<0.02	0.038	5.3	4.2	<0.27	28.1	<0.27	11.1
2/6m-PFOS	60	46	0.11	212	13	123	0.040	0.030	<0.02	0.19	<0.02	0.091	6.6	5	<0.27	38.5	0.80	13.4
3/4/5m-PFOS																		

PFAS = per- and polyfluoroalkyl substances, PFOA = perfluorooctanoic acid, L-PFOS = linear perfluorooctane sulfonic acid, 2/6m-PFOS = branched, sum of perfluoro-2/6-methylheptane sulfonic acid, 3/4/5m-PFOS = branched, sum of perfluoro-3/4/5-methylheptane sulfonic acid, P10 = tenth percentile, P90 = ninetieth percentile.  
<sup>a</sup> For fecal PFOA, 57 individuals were excluded due to LOD at 10.8 ng/g dry weight.

Table 3

Estimated urinary and fecal elimination rates, clearance and comparisons between the urinary and fecal elimination rates, by division and percentage fecal elimination in 147 study participants.

	PFOA		L-PFOS		2/6m-PFOS		3/4/5m-PFOS	
	Mean	Median (P10, P90)	Mean	Median (P10, P90)	Mean	Median (P10, P90)	Mean	Median (P10, P90)
Urinary elimination rate (ng/day)	36	26 (8, 78)	154	91 (12, 404)	28	15 (7, 67)	70	39 (9, 165)
Renal clearance (mL/day)	3.6	2.7 (1.2, 5.6)	1.5	0.9 (0.2, 3.6)	1.3	0.7 (0.2, 3.2)	1.7	0.9 (0.3, 3.5)
per body weight (mL/day/kg)	0.047	0.034 (0.015, 0.078)	0.020	0.011 (0.003, 0.052)	0.017	0.009 (0.003, 0.040)	0.022	0.012 (0.004, 0.053)
Fecal elimination rate (ng/day)	17 <sup>a</sup>	15 (5, 30) <sup>a</sup>	451	364 (95, 886)	157	126 (5, 316)	192	150 (25, 404)
Fecal Clearance (mL/day)	1.6 <sup>a</sup>	1.2 (0.6, 3.0) <sup>a</sup>	4.2	3.6 (1.9, 6.9)	4.8	3.7 (1.1, 9.6)	4.0	2.8 (0.9, 8)
per body weight (mL/day/kg)	0.022 <sup>a</sup>	0.017 (0.008, 0.041) <sup>a</sup>	0.053	0.048 (0.025, 0.084)	0.060	0.053 (0.014, 0.118)	0.050	0.039 (0.013, 0.094)
Ratio of elimination rates (fecal rate/urinary rate)	0.7 <sup>a</sup>	0.6 (0.2, 1.7) <sup>a</sup>	8.8	4.1 (0.9, 19.2)	9.5	5.7 (0.7, 19.5)	6.0	3.3 (0.5, 12.6)
Percental fecal elimination (%)	36 <sup>a</sup>	36 (14, 63) <sup>a</sup>	76	80 (48, 95)	77	85 (42, 95)	70	77 (34, 93)

PFAS = per- and polyfluoroalkyl substances, PFOA = perfluorooctanoic acid, L-PFOS = linear perfluorooctane sulfonic acid, 2/6m-PFOS = branched, sum of perfluoro-2/6-methylheptane sulfonic acid, 3/4/5m-PFOS = branched, sum of perfluoro-3/4/5-methylheptane sulfonic acid, P10 = tenth percentile, P90 = ninetieth percentile.  
<sup>a</sup> These calculations are based on 90 individuals, as 57 fecal samples were excluded due to LOD at 10.8 ng/g dry weight.

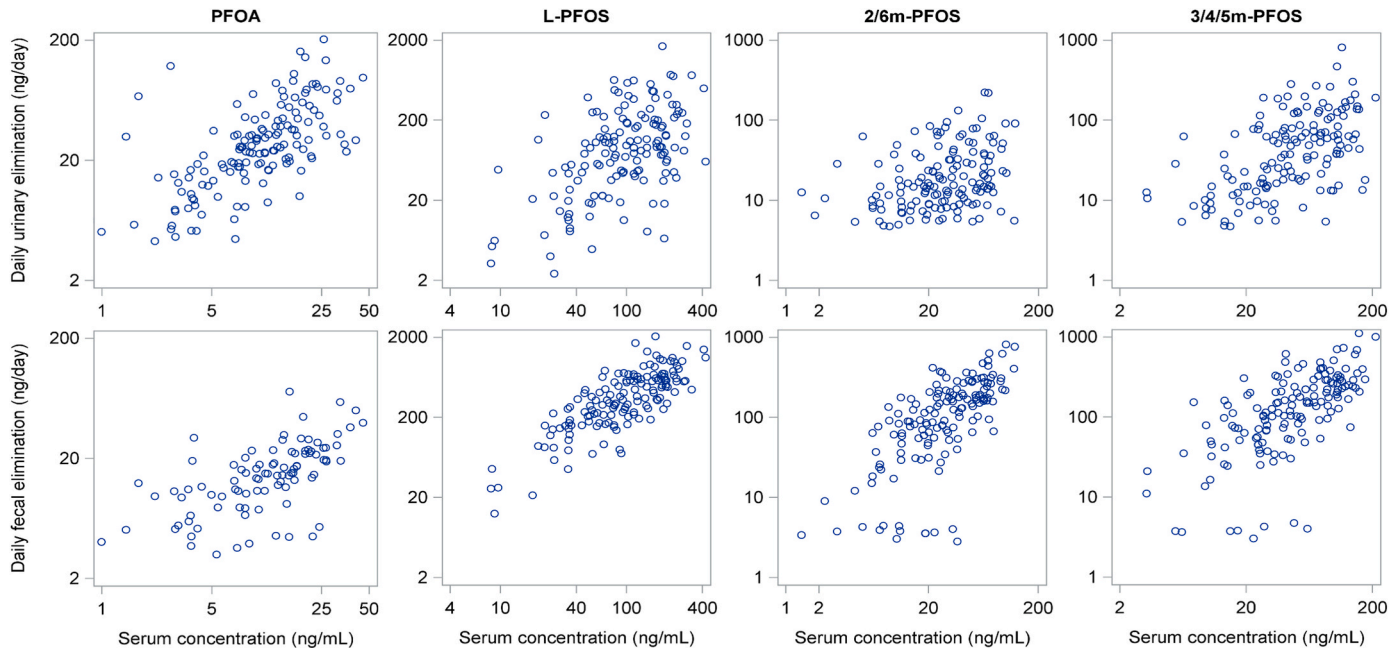


Fig. 2. Scatter plots comparing fecal and urinary elimination and serum concentrations for PFOA, L-PFOS, 2/6m-PFOS and 3/4/5m-PFOS.

fecal and renal clearance were strongly associated, but in different directions (i.e. that fecal clearance was positively associated with fecal/urinary ratio, while renal clearance was negatively associated with the ratio) (Table S7). There were no other clear associations for the other covariates investigated (sex, BMI, fecal zonulin, fecal calprotectin, fecal water content and eGFR).

The median Vd for PFOA was 74 (P10, P90: 48, 176) mL/kg, 93 (P10, P90 52, 192) for L-PFOS, 85 (P10, P90: 31, 184) mL/kg for 2/6m-PFOS, and 91 (P10, P90: 44, 230) mL/kg for 2/6m-PFOS (Table 4). Sensitivity analyses with no blood loss assumed, adjustments for blood donation, exclusion of fertile females and blood donors, and by using median or mean parameter values yielded similar Vds (Table S8). The largest impact of sensitivity analyses came from applying Sallsten and Barregard (2021) median daily creatinine values, which yielded 5–13 mL/kg higher Vds (Table S8).

Fecal elimination was calculated from concentration in dry feces and assumed daily dry weight, adjusted for weight. Urinary elimination was calculated from urinary concentrations, adjusted for urine creatinine concentrations, and daily creatinine production. Note the logarithmic

scales on both axes. Urine and fecal data from one individual with serum L-PFOS levels of 0.38 ng/mL are not shown. For fecal PFOA, 57 individuals were excluded due to LOD at 10.8 ng/g dry weight.

PFOA = perfluorooctanoic acid, L-PFOS = linear perfluorooctane sulfonic acid, 2/6m-PFOS = branched, sum of perfluoro-2/6 methylheptane sulfonic acid, 3/4/5m-PFOS = branched, sum of perfluoro-3/4/5-methylheptane sulfonic acid.

Violin and Sion plot showing the distribution of the fecal percentage of the total elimination. Fecal elimination was calculated from the concentration in dry feces and the assumed daily dry weight, adjusted for weight. Urinary elimination was calculated from urinary concentrations, adjusted for urine creatinine concentrations, and daily creatinine production. Note that 57 out of the 147 PFOA samples were excluded due to a LOD at 10.8 ng/mL dry weight.

PFOA = perfluorooctanoic acid, L-PFOS = linear perfluorooctane sulfonic acid, 2/6m-PFOS = branched, sum of perfluoro-2/6-methylheptane sulfonic acid, 3/4/5m-PFOS = branched, sum of perfluoro-3/4/5-methylheptane sulfonic acid.

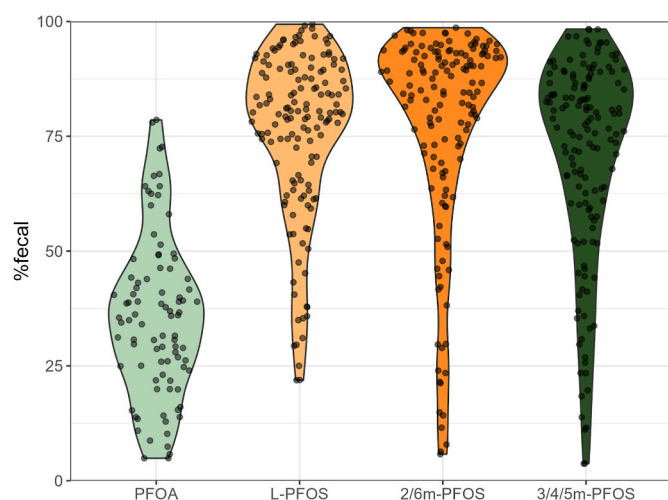


Fig. 3. Relative fecal to total (urinary plus fecal) elimination for PFOS isomers and PFOA in the 147 study subjects.

Table 4

Serum elimination constants (per year) and estimated volume of distributions in 40 individuals with repeated measurements.

PFAS	N	Serum elimination constant (k) [per year]		Volume of distribution (Vd) [mL/kg body weight]	
		Mean	Median (P10, P90)	Mean	Median (P10, P90)
PFOA	36	0.27	0.23 (0.17, 0.41)	96	74 (48, 176)
L-PFOS	40	0.23	0.22 (0.15, 0.32)	116	93 (52, 192)
2/6m-PFOS	40	0.23	0.21 (0.17, 0.31)	96	85 (31, 184)
3/4/5m-PFOS	40	0.18	0.17 (0.12, 0.25)	119	91 (44, 230)

PFAS = per- and polyfluoroalkyl substances, PFOA = perfluorooctanoic acid, L-PFOS = linear perfluorooctane sulfonic acid, 2/6m-PFOS = branched, sum of perfluoro-2/6-methylheptane sulfonic acid, 3/4/5m-PFOS = branched, sum of perfluoro-3/4/5-methylheptane sulfonic acid, P10 = tenth percentile, P90 = ninetieth percentile.

For PFOA, 4 individuals were excluded due to LOD at 10.8 ng/g dry weight for fecal concentration. When calculating Vd, the serum elimination constant per day (i.e. the k above divided by 365) was used.

#### 4. Discussion

To our knowledge, this is the first study to present PFOA and PFOS concentrations in serum, urine and feces in a large sample group with PFAS serum levels ranging from low to very high levels. Even though previous small studies in humans presented fecal concentrations (e.g., Genuis et al., 2013; Genuis et al., 2013)) and urine concentrations (e.g., Li et al., 2022; Li et al., 2022)) in relation to serum concentrations, no previous study has investigated the differences and the relationships between the two different elimination routes for PFOA and PFOS. We found that both PFOA and PFOS were eliminated through both urine and feces. For the PFOS isomers, the fecal elimination was even higher than the urinary elimination. This is also the first study where Vds were estimated using data on individual change of body burden and serum concentrations.

We showed that PFOS and PFOA have different main elimination routes: the PFOS isomers (L-PFOS, 2/6m-PFOS and 3/4/5m-PFOS) were predominantly eliminated via feces, whereas PFOA was predominantly eliminated via urine. There are several possible explanations for this observation. It could be that the affinity for renal, hepatic and intestinal transporters such as organic anion transporters (OATs) differs among PFAS, as recently shown in proximal tubule cells *in vitro* (Ryu et al., 2024). Also, interactions with the gut microbiome could affect PFAS elimination differently, suggested by a recent study of PFAS

bioaccumulation in bacteria strains *in vitro* and PFNA fecal elimination *in vivo* in mice with and without human gut bacteria (Lindell et al., 2024). Interestingly, two studies in 4 Angus steers by Lupton et al. (2012 & 2014) replicated this different elimination profiles between PFOS and PFOA – 11.1 % and 0.5 % of the given PFOS dose (max mean serum concentration 52.6 µg/mL) were eliminated through feces and urine, respectively, over 28 days (Lupton et al., 2014); while 4.6 % and 100.7 % of the given PFOA dose (max mean plasma concentration 4.9 µg/mL) were eliminated through feces and urine, respectively, over 8 days (Lupton et al., 2012). However, such dramatic differences between PFOS and PFOA were not seen in our study.

This is also the first human study to show the variation between individuals in fecal and urinary elimination of PFAS, with some individuals having a predominant fecal elimination route and others having a predominant urinary route. These patterns were correlated between PFAS compounds, suggesting common elimination mechanisms. There could be several different mechanisms, such as differences in genetic expression of PFAS-binding transporters such as OATs, kidney function, or differences in the gut microbiome and gut inflammation. However, we did not find any evidence for markers of kidney function (eGFR), gut inflammation (calprotectin) or gut permeability (zonulin) being associated with the elimination patterns. Interestingly, the negative association between fecal elimination and age found in this study could be the explanation behind the longer half-lives with advancing age we previously observed in the Ronneby half-life panel study (Li et al., 2022).

While no other study has measured fecal PFAS concentrations and from this estimated fecal clearance in humans, two studies using 24-h urine measurements yielded similar mean renal clearances of 0.033 (mL/day/kg) in women (K. Harada et al., 2005), 0.027 in men (K. Harada et al., 2005) and 0.044 in both men and women (Yukiko Fujii et al., 2015) for PFOA and 0.019 for women (K. Harada et al., 2005) and 0.012 for men (K. Harada et al., 2005) for PFOS, compared to our mean 0.047 for PFOA and 0.020 for L-PFOS. However, other studies based on spot urine samples estimated higher mean values of 0.065–0.77 mL/day/kg for PFOA, but similar mean values 0.015–0.045 mL/day/kg for L-PFOS (Zhang et al., 2013; Zhou et al., 2014). Fujii et al. also estimated a mean fecal PFOA clearance of 0.052 mL/day/kg (Yukiko Fujii et al., 2015) from human bile PFOA concentrations, based on a half-life of 3.8 years and Vd of 200 mL/kg. They also estimated a mean fecal PFOA clearance of 0.021 mL/day/kg (Y. Fujii and Harada, 2025) based on the absorption ratio from the mouse gut. These estimations of fecal PFOA clearance are similar to our finding of mean 0.022 mL/day/kg.

The enterohepatic circulation of PFAS is likely extensive. It has been investigated in animal models (Cao et al., 2022; Lupton et al., 2014), but less is known in humans. A study measuring PFAS levels in both bile and serum from four patients with biliary disorders reported median concentrations of 1.0 ng/mL for PFOA and 27.9 ng/mL for PFOS in bile, and 3.8 ng/mL for PFOA and 23.2 ng/mL for PFOS in serum (K. H. Harada et al., 2007). Based on the assumptions of a constant PFAS bile concentration and a bile flow of 350 mL/day, Harada et al. estimated the biliary clearance to be 1.06 mL/day/kg for PFOA and 2.98 mL/day/kg for PFOS (K. H. Harada et al., 2007). Dividing our fecal clearance of 0.017 mL/day/kg for PFOA and 0.046 mL/day/kg for PFOS (mean of the median fecal clearance of L-PFOS, 2/6m-PFOS and 3/4/5m-PFOS) (Table 3) with their respective biliary clearance yield ratios of 1.6 % for PFOA and 1.5 % for PFOS. However, Harada et al. assumed a conservative bile flow of 350 mL/day, while another study estimated it to be 604 mL/day (Boyer and Bloomer, 1974). If a bile flow of 604 mL is instead assumed, the ratios of biliary/fecal clearance would be 0.9 % for both PFOA and PFOS. This suggests that only a small percentage of the PFOA and PFOS (and possibly other PFAS) that is excreted in bile ends up in feces, i.e., there is an enterohepatic recirculation of approximately 99 %.

If the enterohepatic recirculation percentages are similar for PFOA and PFOS, then the difference in fecal elimination between PFOA and



PFOS is more likely a result of hepatic differences (i.e. that PFOS is more secreted into bile than PFOA). This is also supported by the almost three times higher biliary clearance presented by Harada et al. (K. H. Harada et al., 2007). Furthermore, since the enterohepatic recirculation may be as extensive as 99 %, there is a large potential for increasing the elimination rate by blocking bile reuptake pharmacologically. The cholestyramine studies cited earlier (Genuis et al., 2010, 2013; Møller et al., 2024) support this notion.

This is also the first study that estimates Vd with total elimination amounts corresponding to change of body burden, yielding median Vd values of 74 mL/kg for PFOA and 93 mL/kg for L-PFOS. These values are more than 2- to 3-fold less than the Vds estimated through serum to drinking water ratios (Chiu et al., 2022; Thompson et al., 2010). It is possible that our fecal and/or urinary elimination estimations, based on spot samples instead of 24-h measurements, could underestimate the true daily elimination and thus the numerator in the Vd calculation.

One sensitivity analysis where daily creatinine production was extrapolated from Sallsten and Barregard (2021) (Sallsten and Barregard, 2021), yielded 5–13 mL/kg higher Vds (Table S8). However, the effect is too small to account for the large difference in Vds compared to other studies, and it is unclear if using set creatinine levels for women and men are better than estimating individual values using the equation from Forni Ogna et al. (2015) (Forni Ogna et al., 2015).

Conversely, the calculations using drinking water (e.g. Chiu et al., 2022; Thompson et al., 2010) could be overestimating Vd values, since they are necessarily based on several assumptions. These include that the populations have reached a steady state where excretion and intake are balanced, using average estimates of the amount of tap water consumed, and not having data on individual excretion rate constants. None of these limitations are present in this study. The only other study with individual data on a single subject, derived Vds using oral doses and multiple measurements in serum and ended up with estimations closer to ours, at 121 mL/kg for PFOA and 152 mL/kg for PFOS (Abraham et al., 2024).

There are limitations in this study. A limitation was the need to exclude the 57 PFOA measurements with LOD >10.8 ng/g dry weight, which reduced the precision in our measurements of the rate of fecal elimination. Fortunately, it seems that this did not introduce bias, as the 57 excluded individuals were similar to the other 90 regarding age, sex, and serum-adjusted PFAS elimination rates (Table S4).

Our experience of measuring PFAS in feces has been challenging. There are few laboratories performing fecal PFAS analyses, compared to serum and urine analyses, and the method needs to be developed further to increase accuracy and robustness.

Another limitation was that only single spot samples of urine and feces were taken each day, and daily elimination rates were estimated indirectly through creatinine and fecal dry weight. These estimations risk introducing errors, which would have been reduced by conducting 24-h sampling or repeated sampling. But it is not clear in which direction any resulting bias would operate. Regardless, these estimation methods need to be refined and validated.

In conclusion, the fecal elimination route are important and for PFOS, higher than the urinary elimination route. Current values for Vd, used in risk assessments and pharmacokinetic modeling, may be overestimated. Assuming no fecal elimination would result in overestimations of half-lives and, consequently, overly cautious risk assessment of drinking water exposure. Similarly, the use of overestimated Vds would result in overestimations in body burdens. Revision of pharmacokinetic models is thus needed to ensure that improved estimates of PFAS fecal elimination and Vds are properly taken into consideration.

#### CRedit authorship contribution statement

**Axel G. Andersson:** Writing – review & editing, Writing – original draft, Visualization, Software, Project administration, Formal analysis,

Data curation, Conceptualization. **Tony Fletcher:** Writing – review & editing, Resources, Funding acquisition, Conceptualization. **Yiyi Xu:** Writing – review & editing. **Anna Kärrman:** Writing – review & editing, Resources, Methodology, Investigation. **Daniela Pineda:** Writing – review & editing, Investigation. **Carina A. Nilsson:** Writing – review & editing, Investigation. **Christian H. Lindh:** Writing – review & editing, Resources, Methodology, Investigation, Funding acquisition. **Kristina Jakobsson:** Writing – review & editing, Supervision, Project administration, Funding acquisition, Conceptualization. **Ying Li:** Writing – review & editing, Supervision, Formal analysis, Conceptualization.

#### Declaration of competing interest

The authors declare the following financial interests/personal relationships which may be considered as potential competing interests: Kristina Jakobsson reports financial support was provided by Swedish Research Council Formas. If there are other authors, they declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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#### Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.envres.2025.122487>.

#### Data availability

Data will be made available on request.

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