

Effect of dietary consumption as a modifier on the association between *FTO* gene variants and excess body weight in children from an admixed population in Brazil: the Social Changes, Asthma and Allergy in Latin America (SCAALA) cohort study

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Abstract

Previous studies have shown associations of variants of the *FTO* gene with body weight, but none of these have involved Latin American populations with a high level of miscegenation, as is seen in the north-eastern Brazilian population. This study evaluated the association between SNP in the *FTO* gene and excess weight in Salvador, Bahia, Brazil. In addition, the effect of diet as a modifier on this association was also investigated. This cross-sectional study included 1191 participants aged 4–11 years, who were genotyped for 400 variants of the *FTO* gene. Direct anthropometric measures were made and dietary data were obtained by 24-h food recall. Multivariate logistic regression analyses were used to assess the associations of interest. Overall, 11.2% of the individuals included in the study were overweight/obese. Interactions were identified between the percentage energy intake from proteins and obesity risk linked to the rs62048379 SNP ($P_{\text{interaction}} = 0.01$) and also between fat intake (PUFA:SFA ratio) and obesity risk linked to the rs62048379 SNP ($P_{\text{interaction}} = 0.01$). The T allele for the variant rs62048379 was positively associated with overweight/obesity in individuals whose percentage energy intake from protein was above the median (OR 2.00; 95% CI 1.05, 3.82). The rs62048379 SNP was also associated with overweight/obesity in individuals whose PUFA:SFA ratio was below the median (OR 1.63; 95% CI 1.05, 2.55). The association between *FTO* gene variants and excess body weight can be modulated by dietary characteristics, particularly by fatty acid distribution and dietary protein intake in children.

Key words: Dietary intake: *FTO* variants: Adiposity: Children

Genetic factors and several postnatal factors such as environmental, social, economic, cultural, psychosocial and behavioural elements are associated with an increased risk for overweight and obesity, and constitute a set of factors that are interrelated and mutually compounding, making overweight/obesity a complex and multifaceted trait⁽¹⁾. Nevertheless, the factors that best explain the growing number of overweight individuals are related to changes in

lifestyle, such as the increased consumption of high-energy processed food, particularly those foods with a high content of SFA and simple carbohydrates, and by increased sedentariness⁽²⁾. Currently, there is a growing interest in understanding the genetics of complex traits such as overweight/obesity, which are characterised by a multiplicity of interactions between genetic determinants and conditions associated with lifestyle and the environment.

Abbreviation: FTO, fat mass and obesity.

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With the exception of the Y chromosome, studies have shown that all other human chromosomes contain loci associated with the risk for excess weight^(3,4). In various studies using different protocols and different methodologies, more than 430 genes associated with overweight and obesity have been identified⁽⁵⁾. Of all of the genes associated with overweight and obesity, the fat mass and obesity-associated (*FTO*) gene merits particular mention because it explains the largest amount of genetic variance in obesity traits over the lifespan⁽⁴⁾. The *FTO* gene has been associated with energy homeostasis, participating directly in the control of body fat reserves⁽⁶⁾. There is evidence supporting the role of some common genetic variants of the *FTO* gene in increased food intake and the choice of more energy-dense foods⁽⁷⁻⁹⁾. Currently, studies have shown the effects of components consumed in the diet on gene expression⁽⁸⁾. Some dietary components, particularly SFA, are hypothesised to interact with some of the variants of the *FTO* gene and modify the effect of the association between these genetic variants and obesity in such a way as to compound the obesogenic effect of the *FTO* gene^(7,10). However, the mechanisms by which these associations are established remain to be fully clarified.

Previous studies have shown associations of variants of the *FTO* gene with body weight⁽¹¹⁻¹⁸⁾ but none of these has involved Latin American populations with a high level of miscegenation, as is seen in the north-eastern Brazilian population. Therefore, the objective of the present study was to evaluate the association between SNP in the *FTO* gene and excess weight in a population of children living in the city of Salvador, Brazil. In addition, the effect of diet as a modifier of this association was assessed in these Brazilian children. The hypothesis was that SNP in the *FTO* gene are associated with overweight/obesity and that any associations could be modified by dietary intake to modulate the effect of these polymorphisms on excess weight.

Methods

Ethical aspects

The parents or legal guardians of each participating child signed an informed consent form in which the study procedures were described in detail. The study protocol was approved by the internal ethics committee of the Collective Health Institute of the Federal University of Bahia and by the National Research Ethics Council (CONEP) under references 003-05/CEP-ISC and 15.895/2011, respectively.

Study design and population

This is a cross-sectional, population-based study nested within the cohort study Social Changes, Asthma and Allergy in Latin America (SCAALA), which was conducted to evaluate risk factors for asthma in Salvador. The methodology used in that study has been described previously⁽¹⁹⁾. The city of Salvador has a population of over 2.6 million inhabitants, of whom 80% consider themselves black or of mixed race⁽²⁰⁾. The sample consisted of 1445 children who were randomly selected from 20 000 households encompassing a variety of different

socio-economic levels and environmental conditions. Of these children, genetic data were available for 1309 children. Following quality control, two children were excluded from the study because data on their obesity/overweight status were missing, and sixty-one were excluded because data on kinship were lacking, leaving 1246 eligible children. After a review of the questionnaires (24-h diet recall), we excluded fifty-five cases because of inconsistent data, and the final sample was made up of 1191 children.

Data collection

Anthropometric data. The participants were weighed on a portable electronic microscale (Filizola[®], model E-150/3P, São Paulo, Brazil), and their heights were measured using a portable stadiometer (Leicester Height Measure[®]; Seca). BMI was used for the diagnosis of anthropometric status, adopting the percentiles for age and sex proposed by the WHO⁽²¹⁾. Anthropometric status was classified as follows: underweight, <3rd percentile; normal weight, ≥3rd percentile and <85th percentile; overweight, ≥85th percentile and <97th percentile; and obese, ≥97th percentile. The categories of overweight and obese were grouped together; therefore, children classified as having excess weight were those ≥ the 85th percentile.

Genotyping

DNA extraction was performed according to the manufacturer's instructions using the Genra Puregene Blood Kit. The Qubit fluorometer (Invitrogen) was used for DNA quantification⁽²²⁾. All children were genotyped for 458 variants of the *FTO* gene using the Illumina genotyping platform HumanOmni2.5-8 BeadChip. The Illumina Human Omni 2.5 MV-3 chip delivers comprehensive coverage of both common and rare SNP content from the 1000 Genomes Project (1KGP; minor-allele frequency (MAF) >2.5%), designed to be maximally informative for diverse world populations. We selected all SNP in the *FTO* gene (Assembly GRCh37.p13, localised between 53737875 and 54148379 positions) covered by this platform. Genotyping data have been deposited in the European Genome-phenome Archive (EGA, <http://www.ebi.ac.uk/ega/>), which is hosted by the European Bioinformatics Institute (EBI), under the accession number EGAS00001001245. Imputation was performed only for the variant rs9939609 using the IMPUTE2 package⁽²³⁾ on the public panel from 1000 Genomes Project Phase I data 'version 3' (ALL.integrated_phase1_SHAPEIT_16-06-14.nomono.integrated_phase1_v3.20101123.snps_indels_svsgenotypes.nomono.haplotypes.gz), which contained 1092 individuals of various ethnicities.

Quality control was performed before conducting the tests for association. All procedures were automated and performed using the PLINK program, version 1.9⁽²⁴⁾. To evaluate family structure, kinship coefficients were calculated for every possible pair. A total of sixty-one individuals were removed from the sample because of the relationship determined by kinship coefficients for each possible pair of individuals. This method is implemented in the REAP software (Relatedness Estimation in Admixed Populations)⁽²⁵⁾. Quality control was conducted in stages to exclude SNP with a genotyping call rate <0.98;

deviation from Hardy–Weinberg equilibrium was determined using controls only, with a P -value <0.05 and MAF $<1\%$ ⁽²⁶⁾.

Population structure

Principal component analysis (PCA) was conducted to identify different population groups based on the ethnic history/origin of each individual, and correct the population structure⁽²⁷⁾. Details on the use of PCA to evaluate population stratification can be found in the paper published by Costa *et al.* in 2015⁽²⁸⁾.

Dietary intake

The 24-h diet-recall method (R24h) was used to determine dietary intake. Parents reported their children's dietary intake. However, the information given by the children at the time of the interview complemented the information given by their parents. Food consumed in school or at day-care centres was also recorded. The food consumption recorded was converted into energy and macronutrients using DietPro software, version 4.0, 2006⁽²⁹⁾. Foods that did not form part of the database of this software were added, using information contained in the Brazilian Food Composition Table (TACO)⁽³⁰⁾, Assessment Table for Food Consumption from Home-Cooking Measurements⁽³¹⁾, the ENDEF (*Estudo Nacional de Despesas Familiares*) food composition table⁽³²⁾ and packaged-food labels. Macronutrient intake was expressed as the percentage of total energy intake.

Statistical analysis

The characteristics of the population were identified by conducting a descriptive analysis. χ^2 Tests were used for categorical variables. The Mann–Whitney U test was used, where appropriate, in order to compare dietary intake variables for 'overweight' and 'not overweight' groups because of their highly skewed distribution. Logistic regression analysis was used to evaluate the association between genetic variants and excess weight. Each SNP of the *FTO* gene was analysed separately. Because of the low numbers of AA homozygotes, the genotype was analysed using the dominant-allele model of genetic heritability. The models were adjusted for sex, age, population structure (determined by the first three principal components) and energy intake. These potential confounding variables were selected on the basis of data published in other studies⁽³³⁾. Genomics and proteomics analyses regularly involve the simultaneous testing of hundreds of hypotheses, either on numerical or on categorical data. To correct for the occurrence of false positives, validation tests based on multiple testing correction, such as the Bonferroni and the Benjamini and Hochberg false discovery rate as well as re-sampling techniques (i.e. a permutation-based test) are frequently used. In this paper, we used a permutation test as it is less stringent and has become a widely accepted and recommended approach in studies that involved multiple statistical tests for genetic markers⁽³⁴⁾.

Therefore, the empirical P -values were obtained after 50 000 phenotype permutations to limit the occurrence of type-I errors

(false-positive results). After the permutation tests, P -values <0.05 were considered statistically significant. Statistical power was estimated with the GAS Power Calculator (<http://csg.sph.umich.edu//abecasis/CaTS/>)⁽³⁵⁾. Linkage disequilibrium values between the evaluated SNP were determined by r^2 in addition to D' analysis using the Haploview program, version 4⁽³⁶⁾. Interactions between the *FTO* genotype and dietary intake, and their effect on overweight/obesity were tested by including the respective interaction terms in the models. All statistical tests were two-tailed, and the significance level was defined as 5%. All of the statistical analyses were performed using PLINK software, version 1.9⁽²⁴⁾.

Results

The eligible study population consisted of 1445 children aged 4–11 years. Of these, 1191 children were included in the analysis, and 11.2% of these children were overweight/obese. There was a slightly higher percentage of boys than girls (54 *v.* 46%) and of children aged 6–7 years (41.8%). With respect to skin colour, the vast majority of children were reported as being brown or black (91.50%). The median energy intake was 7045.85 (1928.78–38940.99) kJ. The median intake of carbohydrates, proteins and fat as a percentage of energy content was 61.68 (range: 27.08–87.38), 12.58 (range: 4.28–10.93) and 26.08 (range: 5.28–51.28)%, respectively. The median PUFA:SFA intake was 0.93 (range: 0.02–3.89). There were significant differences in the percentage of energy from carbohydrates ($P=0.046$) and fat ($P=0.037$) between the non-overweight/non-obese and overweight/obese groups (Table 1).

A genetic analysis, using a logistic regression model adjusted for sex, age, population structure and energy intake, was conducted to evaluate the association between the *FTO* gene variants and excess weight. As shown in Table 2, ten SNP in introns 1, 7 and 8 of the *FTO* gene were associated ($P<0.05$) with excess weight in the population evaluated (online Supplementary Fig. S1). The top SNP associated with excess weight were rs115530394 and rs75066479 for the dominant genetic model. The results of all 400 SNP are presented (online Supplementary Table S1). In addition, a regional association plot was produced using LocusZoom⁽³⁷⁾ for better understanding of the genomic context (online Supplementary Fig. S2).

A polymorphism associated with a particular trait is not necessarily a causal mutation; rather, it may be in linkage disequilibrium with a functional variant. For this reason, an initial investigation was conducted to determine whether SNP associated with excess weight in the Brazilian population capture a single signal or whether they are independently associated with this condition. Of all *FTO* polymorphisms significantly associated with excess weight, only two pairs were in strong linkage disequilibrium. One pair is located in intron 7 (rs115530394/rs114019148) and the other in intron 8 (rs75066479/rs115662052). In addition, we observed low linkage disequilibrium between the variants rs62048379 and rs9939609 (r^2 0.004 and $D'=0.17$; r^2 0.004 and $D'=0.17$). We included rs9939609 in the analysis and indicated that it was imputed (Fig. 1).

Table 1. Characteristics of the population. Salvador, Bahia, Brazil, 2004–2005

Variables	Anthropometric status						<i>P</i> *	
	Total		Not overweight		Overweight			
	<i>n</i>	%	<i>n</i>	%	<i>n</i>	%		
Sex								
Male	552	46.3	495	46.8	57	42.9	0.390	
Female	639	53.7	563	53.2	76	57.1		
Age (years)								
4–5	315	26.4	284	26.8	31	23.3	0.295	
6–7	498	41.8	446	42.2	52	39.1		
8–11	378	31.7	328	31.0	50	37.6		
Skin colour								
White	101	8.5	86	8.1	15	11.3	0.219	
Black/Brown	1090	91.5	972	91.9	118	88.7		
Dietary intake	<i>n</i>	Median	Minimum–maximum	Median	Minimum–maximum	Median	Minimum–maximum	<i>P</i> †
Energy content (kJ)	1191	7848.11	1928.78–80780.99	6980.29	1928.78–20134.03	7386.55	2582.36–80780.99	0.081
Carbohydrates (% energy)	1191	61.68	27.08–87.38	61.77	27.08–87.38	60.65	42.53–77.19	0.046
Proteins (% energy)	1191	12.70	4.28–82.69	12.53	4.28–82.69	13.19	4.86–27.63	0.061
Fat (% energy)	1191	26.08	5.28–51.28	25.93	5.28–51.28	27.26	10.24–51.17	0.037
Dietary PUFA:SFA	1191	0.93	0.02–3.89	0.92	0.02–3.89	0.94	0.11–3.66	0.833

* χ^2 Test.† Mann–Whitney *U* test.**Table 2.** Logistic regression between *FTO* variants and excess weight. Salvador, Bahia, Brazil, 2004–2005

SNP	BMI-increasing allele	MAF	Dominant model		
			OR	<i>P</i> *	Power
rs115530394	G	0.02	2.42	0.003	0.83
rs75066479	G	0.01	2.47	0.008	0.58
rs2003583	A	0.26	1.64	0.010	0.93
rs115662052	G	0.01	2.52	0.012	0.60
rs114019148	A	0.02	2.10	0.014	0.67
rs62048379	A	0.08	1.66	0.024	0.82
rs62034079	T	0.05	1.75	0.044	0.73
rs79149291	A	0.01	2.20	0.046	0.46
rs1123817	A	0.04	1.74	0.044	0.68
rs16952663	A	0.05	1.70	0.049	0.71

FTO, fat mass and obesity; MAF, minor-allele frequency.* *P* = permutation test (50 000 permutations). Dominant model.

Interaction between *FTO* variants and dietary intake on overweight/obesity

An analysis was conducted of the effect of dietary intake on the association between *FTO* gene variants and excess weight with the objective of evaluating the modifying effect of diet on this association. Interactions between the *FTO* genotype and dietary intake, and the effect on overweight/obesity were tested by including the respective interaction terms in the models. For these analyses, the percentage contribution of energy obtained from carbohydrates, proteins, total fat and MUFA was dichotomised according to whether this percentage was above or below the median, as was the ratio between PUFA:SFA.

Table 3 shows the results of the multivariate logistic regression analysis conducted to evaluate the association between *FTO* gene variants and overweight/obesity according to dietary intake. An interaction was found between protein intake and the *FTO* gene variant rs62048379 ($P_{\text{interaction}}=0.01$) in relation to

overweight/obesity. The risk allele for the *FTO* gene (A allele) was positively associated with overweight/obesity in individuals whose percentage energy intake from protein was above the median (OR 2.00; 95% CI 1.05, 3.82). An interaction was also found between fat intake (the ratio between PUFA:SFA) and the *FTO* gene variant rs62048379 ($P_{\text{interaction}}=0.01$) in relation to overweight and obesity. The risk allele for the *FTO* gene (A allele) for the variant rs62048379 was positively associated with overweight/obesity in individuals whose PUFA:SFA ratio was below the median (OR 1.63; 95% CI 1.05, 2.55). These data are complemented by the data in the online Supplementary Table S2. Analyses of the remaining *FTO* gene variants associated with overweight/obesity as a function of dietary intake are shown in the online Supplementary Table S3.

Discussion

The present study investigated the possible associations between variants of the *FTO* gene and overweight/obesity in children in Salvador, Bahia, Brazil. The Brazilian population is considered to be highly admixed. According to Table 1, 91.5% of the SCAALA population declares itself to be black or brown. Actually, Lima-Costa *et al.* show that 50% of the genetic composition in the Salvador-SCAALA cohort is of African origin⁽³⁸⁾. Following the appropriate adjustments, SNP that were significantly associated with overweight/obesity were rs115530394, rs75066479, rs115662052, rs114019148, rs62048379, rs62034079, rs79149291, rs1123817, rs16952663 and rs2003583. Variants of the *FTO* gene influencing obesity have been studied in children in Europe⁽¹¹⁾, China⁽¹²⁾ and Japan⁽¹³⁾, as well as in African American⁽³⁹⁾ and Brazilian children (including the rs9939609 SNP)^(14,15). However, other studies conducted with children of the same age group in Mexico⁽¹⁶⁾, Greece⁽¹⁷⁾ and China⁽¹⁸⁾ failed to identify any significant association. The association between overweight/obesity and different variants of the *FTO* gene in different populations may

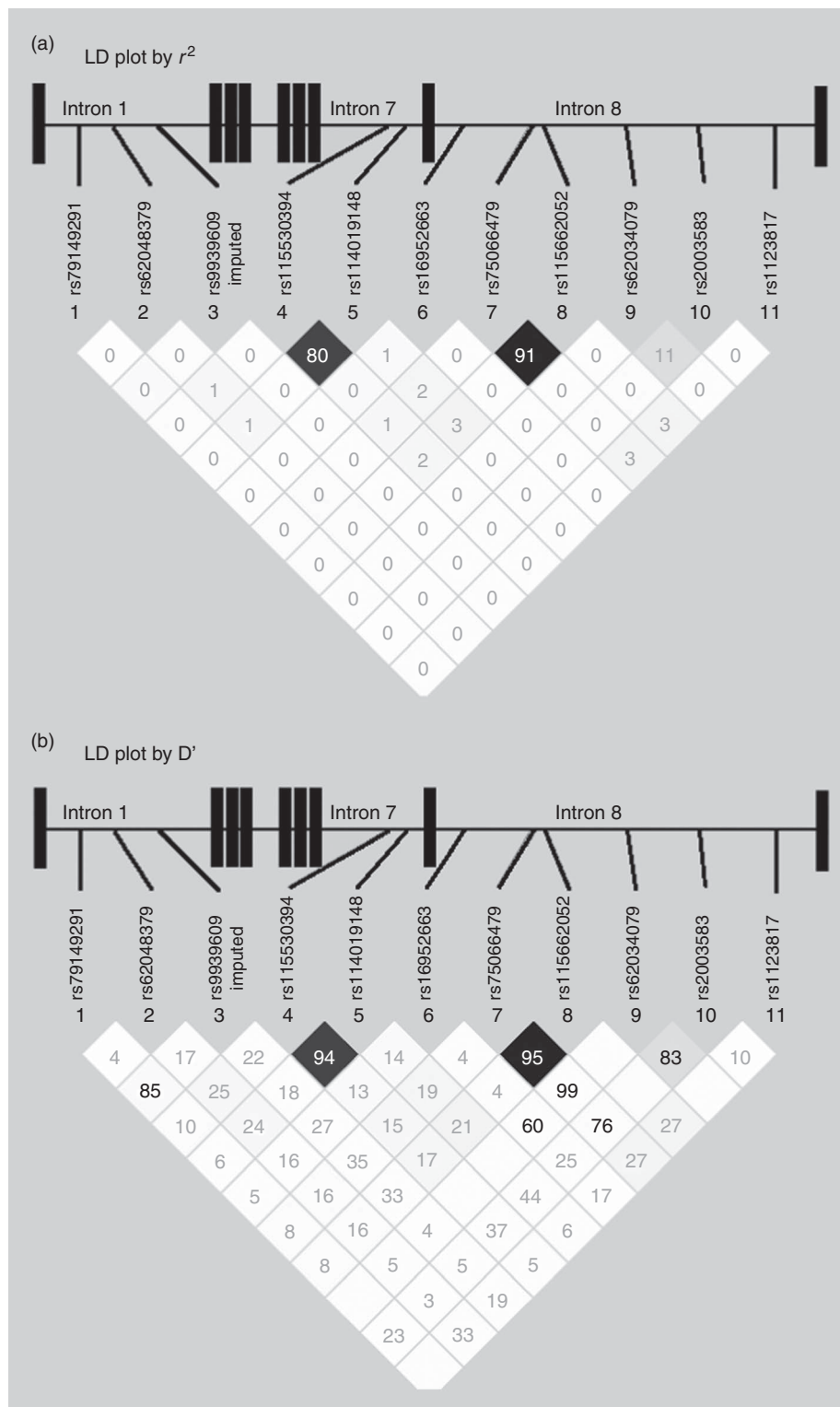


Fig. 1. Linkage disequilibrium (LD) of the SNP associated with excess weight. Salvador, Bahia, Brazil, 2004–2005.

depend on ethnicity or differences in gene–environment interactions⁽⁴⁰⁾.

Previous studies identified associations between obesity and polymorphisms in introns 1^(41,33) and 8⁽⁴²⁾ of the *FTO* gene. Those previous findings, together with the present results,

suggest the occurrence of variants with a regulatory potential in different regions of the *FTO* gene. In the present analysis, variants in introns 1, 7 and 8 of the *FTO* gene were identified as being associated with overweight/obesity. Here, most SNP associated with the trait in question are located in intron 8. This

Table 3. Association between polymorphisms (rs62048379) and phenotypes according to dietary intake. Salvador, Bahia, Brazil, 2004–2005 (Odds ratios and 95% confidence intervals)

	Phenotypes	OR	95% CI	$P_{\text{interaction}}$	
Protein (% energy)	≥Median	Excess weight	2.00	1.05, 3.82	0.01
	<Median	Excess weight	1.32	0.71, 2.45	
Dietary PUFA:SFA	≥Median	Excess weight	0.87	0.43, 1.72	0.01
	<Median	Excess weight	1.64	1.05, 2.55	

indicates that, in addition to intron 1, intron 8 may constitute a second site of *FTO* variants related to overweight/obesity. In a study of African American individuals, an association was found between seven SNP from the *FTO* gene (rs708262, rs11076017, rs16952725, rs9932411, rs7191513, rs2689269, rs16952987 and rs8057044) and overweight/obesity. Some of these SNP are also situated in intron 8⁽⁴²⁾. These results reinforce the suggestion that there are similarities between genetic factors that predispose individuals of African American descent to obesity and predisposing genetic factors in the Brazilian population, particularly in the admixed population of Bahia⁽⁴⁵⁾.

The possible associations between *FTO* gene variants and overweight/obesity were evaluated in accordance with the composition of the diet. The results of studies investigating the interaction between dietary factors and *FTO* gene variants on overweight/obesity have generated conflicting results regarding potential interactions^(44,45,10). In the present study, an interaction was identified between the percentage of energy intake from proteins and obesity risk linked to the rs62048379 SNP ($P_{\text{interaction}}=0.01$). An interaction was also found between fat intake (PUFA:SFA ratio) and obesity risk linked to the rs62048379 SNP ($P_{\text{interaction}}=0.01$). The OR that reflect the magnitude of the association between *FTO* gene variants and overweight/obesity are clearly heterogeneous in this study. Therefore, in individuals whose percentage energy intake from proteins was above the median, the presence of the risk allele (A) for the rs62048379 variant increased the prevalence of excess weight by 2.0 times compared with when the allele was absent. These findings are consistent with the results of a meta-analysis conducted by Qi *et al.*⁽⁸⁾. Those authors showed that a lower dietary protein intake attenuated the association between the *FTO* genotype and adiposity in children and adolescents. Other studies, however, reported contradictory results^(46,47). Studies suggest that *FTO* may influence body composition through cellular sensing of amino acids^(48,49). Given the increasing evidence supporting the role of *FTO* in protein metabolism and body composition, future investigations on this topic may help clarify the mechanisms underlying the observed interaction between the *FTO* variant and protein intake and its effect on overweight/obesity.

The presence of the allele (A) for the rs62048379 variant increased the prevalence of excess weight by 1.63 times in those individuals whose PUFA:SFA ratio was below the median compared with when this allele is absent. A novel finding in this study was that low dietary PUFA:SFA intake ratio accentuated obesity risk in the A allele carriers but not in the TT homozygote carriers in

this population, suggesting that genetic predisposition to obesity may be modulated by dietary SFA intake. The findings of the present study reinforce the results of other studies in which the authors emphasise that diets based on unsaturated fats^(45,50) may attenuate the obesogenic effect of *FTO* on the expression of the obesity phenotype. Studies have indicated a possible ‘anti-obesity’ effect attributable to PUFA. This may be due to their greater oxidative rates when compared with SFA⁽⁵¹⁾. Several investigators have shown that fatty acid oxidation increases directly with a concomitant increase in PUFA:SFA intake^(10,52). The mechanisms allowing dietary SFA to interact with *FTO* are unknown and require further investigation.

The large amount of genome-wide association studies leave no doubt that *FTO* genotypes in humans are linked to obesity. Recently, researchers have begun to elucidate the underlying pathophysiology behind the mechanism by which this gene may act to increase the risk for obesity^(53,54). In fact, very new data obtained using integrated analyses of long-distance regulation, chromatin–chromatin interactions, topologically associated domains, chromatin modifications, gene expression and transgenic animal models have shown that intronic variants within *FTO* may interact with the promoter region of *IRX3/5*, affecting its expression^(53,54). Genetic variants in *FTO* and *IRX3* were already described to be in high linkage disequilibrium⁽⁵⁵⁾. *IRX3* was originally found to be related to the early stage of neural development⁽⁵⁶⁾ and has already been described to be up-regulated in the hypothalamus, which plays a role in food intake or appetite regulation⁽⁵⁴⁾. In this way, *IRX3* may act as a functional long-range target of obesity-associated variants within the *FTO* gene and might drive weight gain and the development of overweight and obesity in carriers of common SNP in the *FTO* gene⁽⁵⁴⁾. Further studies are needed to better explore this point.

Our study was based on an analysis of cross-sectional data, which limits the ability to investigate causality. Another limitation of the present study is related to limited statistical power due to the relatively small cohort size. Thus, we chose the dominant heritability model instead of the additive or recessive model in order to improve the power for statistical analysis. Even so, studies with larger sample sizes are needed to support the findings of the present study. In addition, we were unable to examine other adiposity proxies but were limited to the consideration of BMI, which cannot distinguish body composition and does not provide any indication of body fat distribution. The R24h used in the present study to investigate dietary intake may also represent another limitation. Although it is a rapid, relatively inexpensive and easily applied method, the success of its use depends on the respondent’s memory and requires a well-trained investigator to obtain accurate estimations of the portions consumed. However, this method supplies reliable estimates of the mean dietary intake of a population, even when applied only once, as long as the designated methodology is followed and analytical resources are appropriate⁽⁵⁷⁾. Most of the children included in our analysis are predominantly of African descent, and it is unknown whether our results can be generalised to other ethnic groups. In contrast, the strengths of this study include the genotyping of various SNP along the *FTO* gene (400 SNP). An additional strength

is the use of a permutation test rather than other methods to test for associations and control for family-wise error rate (FWER), such as Bonferroni and false discovery ratio (FDR). The Bonferroni correction assumes complete independence between markers, but markers in proximity to each other or in low linkage disequilibrium are not completely independent, which makes this approach overly stringent here. The FDR avoids the problem of dependence of tests, but it offers weak control for FWER. It permits a number of false discoveries; therefore, uncertainty remains about the accuracy of each significant result. Permutation analyses provide much stronger control for FWER than the FDR⁽⁵⁸⁾. As in all approaches, permutation has its weaknesses. Power decreases when there is a very large number of tests. It is most accurate for simple analyses, unless the sample size is large.

In summary, the majority of SNP associated with obesity in this study are located in intron 8, rather than in intron 1, suggesting that there may be many genetic variants that have not yet been reported in the literature that affect obesity in humans. Furthermore, the association between *FTO* gene variants and excess body weight can be modified by dietary characteristics, particularly by fatty acid distribution and lower dietary protein intake in children, offering new insights into the interrelationships between *FTO* genetic variants, dietary intake and obesity.

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The authors declare that there are no conflicts of interest.

Supplementary material

For supplementary material/s referred to in this article, please visit <https://doi.org/10.1017/S0007114517001386>

References

- Perez LM, Garcia K & Herrera R (2013) Psychological, behavioral and familial factors in obese Cuban children and adolescents. *MEDICC Rev* **15**, 24–28.
- Kourlaba G, Panagiotakos DB, Mihos K, *et al.* (2009) Dietary patterns in relation to socio-economic and lifestyle characteristics among Greek adolescents: a multivariate analysis. *Public Health Nutr* **12**, 1366–1372.
- Berndt SI, Gustafsson S, Magi R, *et al.* (2013) Genome-wide meta-analysis identifies 11 new loci for anthropometric traits and provides insights into genetic architecture. *Nat Genet* **45**, 501–512.
- Livingstone KM, Celis-Morales C, Papandonatos GD, *et al.* (2016) *FTO* genotype and weight loss: systematic review and meta-analysis of 9563 individual participant data from eight randomised controlled trials. *BMJ* **354**, i4707.
- Snyder EE, Walts B, Perusse L, *et al.* (2004) The human obesity gene map: the 2003 update. *Obes Res* **12**, 369–439.
- Gerken T, Girard CA, Tung YC, *et al.* (2007) The obesity-associated *FTO* gene encodes a 2-oxoglutarate-dependent nucleic acid demethylase. *Science* **318**, 1469–1472.
- Moleres A, Ochoa MC, Rendo-Urteaga T, *et al.* (2012) Dietary fatty acid distribution modifies obesity risk linked to the rs9939609 polymorphism of the fat mass and obesity-associated gene in a Spanish case-control study of children. *Br J Nutr* **107**, 533–538.
- Qi Q, Downer MK, Kilpelainen TO, *et al.* (2015) Dietary intake, *FTO* genetic variants and adiposity: a combined analysis of over 16,000 children and adolescents. *Diabetes* **64**, 2467–2476.
- Karra E, O'Daly OG, Choudhury AI, *et al.* (2013) A link between *FTO*, ghrelin, and impaired brain food-cue responsiveness. *J Clin Invest* **123**, 3539–3551.
- Phillips CM, Kesse-Guyot E, McManus R, *et al.* (2012) High dietary saturated fat intake accentuates obesity risk associated with the fat mass and obesity-associated gene in adults. *J Nutr* **142**, 824–831.
- Albuquerque D, Nobrega C & Manco L (2013) Association of *FTO* polymorphisms with obesity and obesity-related outcomes in Portuguese children. *PLOS ONE* **8**, e54370.
- Fang H, Li Y, Du S, *et al.* (2010) Variant rs9939609 in the *FTO* gene is associated with body mass index among Chinese children. *BMC Med Genet* **11**, 136.
- Okuda M, Hinoda Y, Okayama N, *et al.* (2011) Association between the *FTO* gene and overweight in Japanese children and adolescents. *Pediatr Diabetes* **12**, 494–500.
- da Silva CF, Zandona MR, Vitolo MR, *et al.* (2013) Association between a frequent variant of the *FTO* gene and anthropometric phenotypes in Brazilian children. *BMC Med Genet* **14**, 34.
- Lourenco BH, Qi L, Willett WC, *et al.* (2014) *FTO* genotype, vitamin D status, and weight gain during childhood. *Diabetes* **63**, 808–814.
- Mejia-Benitez A, Klunder-Klunder M, Yengo L, *et al.* (2013) Analysis of the contribution of *FTO*, *NPC1*, *ENPP1*, *NEGR1*, *GNPDA2* and *MC4R* genes to obesity in Mexican children. *BMC Med Genet* **14**, 21.
- Scott RA, Bailey ME, Moran CN, *et al.* (2010) *FTO* genotype and adiposity in children: physical activity levels influence the effect of the risk genotype in adolescent males. *Eur J Hum Genet* **18**, 1339–1343.
- Zhang M, Zhao X, Cheng H, *et al.* (2014) Age- and sex-dependent association between *FTO* rs9939609 and obesity-related traits in Chinese children and adolescents. *PLOS ONE* **9**, e97545.
- Barreto ML, Cunha SS, Alcantara-Neves N, *et al.* (2006) Risk factors and immunological pathways for asthma and other allergic diseases in children: background and methodology of a longitudinal study in a large urban center in Northeastern Brazil (Salvador-SCAALA study). *BMC Pulm Med* **6**, 15.
- Instituto Brasileiro de Geografia e Estatística (2016) Sistema SIDRA. <http://www.sidra.ibge.gov.br/bda/tabela/listabl.asp?z=cd&o=4&i=P&c=3145> (accessed June 2016).
- de Onis M, Onyango AW, Borghi E, *et al.* (2007) Development of a WHO growth reference for school-aged children and adolescents. *Bull World Health Organ* **85**, 660–667.
- Costa GN, Dudbridge F, Fiaccone RL, *et al.* (2015) A genome-wide association study of asthma symptoms in Latin American children. *BMC Genet* **6**, 141.
- Howie BN, Donnelly P & Marchini J (2009) A flexible and accurate genotype imputation method for the next generation of genome-wide association studies. *PLoS Genet* **5**, e1000529.



24. Purcell S, Neale B, Todd-Brown K, *et al.* (2007) PLINK: a tool set for whole-genome association and population-based linkage analyses. *Am J Hum Genet* **81**, 559–575.
25. Thornton T, Tang H, Hoffmann TJ, *et al.* (2012) Estimating kinship in admixed populations. *Am J Hum Genet* **91**, 122–138.
26. Laurie CC, Doheny KF, Mirel DB, *et al.* (2010) Quality control and quality assurance in genotypic data for genome-wide association studies. *Genet Epidemiol* **34**, 591–602.
27. Price AL, Patterson NJ, Plenge RM, *et al.* (2006) Principal components analysis corrects for stratification in genome-wide association studies. *Nat Genet* **38**, 904–909.
28. Costa GN, Dudbridge F, Fiaccone RL, *et al.* (2015) A genome-wide association study of asthma symptoms in Latin American children. *BMC Genet* **16**, 141.
29. DietPro (2006) DietPro. Sistema de análise nutricional. Versão 4.0.
30. Núcleo de Estudos e pesquisas em Alimentação (2006) Tabela de composição de alimentos, versão 2. Campinas: Universidade Estadual de Campinas.
31. Pinheiro ABV, Lacerda EMA, Benzecry EH, *et al.* (2005) *Tabela para avaliação de consumo alimentar em medidas caseiras*, 5a ed. Rio de Janeiro: Atheneu.
32. Estudos Nacional da Despesa Familiar (ENDEF) (1996) *Tabelas de composição de alimentos*, 4a ed. Rio de Janeiro: Fundação Instituto Brasileiro de Geografia e Estatística.
33. Qi Q, Downer MK, Kilpelainen TO, *et al.* (2015) Dietary intake, FTO genetic variants, and adiposity: a combined analysis of over 16,000 children and adolescents. *Diabetes* **64**, 2467–2476.
34. Belmonte M & Yurgelun-Todd D (2001) Permutation testing made practical for functional magnetic resonance image analysis. *IEEE Trans Med Imaging* **20**, 243–248.
35. Skol AD, Scott LJ, Abecasis GR, *et al.* (2006) Joint analysis is more efficient than replication-based analysis for two-stage genome-wide association studies. *Nat Genet* **38**, 209–213.
36. Barrett JC, Fry B, Maller J, *et al.* (2005) Haploview: analysis and visualization of LD and haplotype maps. *Bioinformatics* **21**, 263–265.
37. Pruim RJ, Welch RP, Sanna S, *et al.* (2010) LocusZoom: regional visualization of genome-wide association scan results. *Bioinformatics* **26**, 2336–2337.
38. Lima-Costa MF, Rodrigues LC, Barreto ML, *et al.* (2015) Genomic ancestry and ethnoracial self-classification based on 5,871 community-dwelling Brazilians (The Epigen Initiative). *Sci Rep* **5**, 9812.
39. Bollepalli S, Dolan LM, Deka R, *et al.* (2010) Association of FTO gene variants with adiposity in African-American adolescents. *Obesity (Silver Spring)* **18**, 1959–1963.
40. Tan LJ, Zhu H, He H, *et al.* (2014) Replication of 6 obesity genes in a meta-analysis of genome-wide association studies from diverse ancestries. *PLOS ONE* **9**, e96149.
41. Foraita R, Gunther F, Gwozdz W, *et al.* (2015) Does the FTO gene interact with the socioeconomic status on the obesity development among young European children? Results from the IDEFICS study. *Int J Obes* **39**, 1–6.
42. Adeyemo A, Chen G, Zhou J, *et al.* (2010) FTO genetic variation and association with obesity in West Africans and African Americans. *Diabetes* **59**, 1549–1554.
43. Kehdy FS, Gouveia MH, Machado M, *et al.* (2015) Origin and dynamics of admixture in Brazilians and its effect on the pattern of deleterious mutations. *Proc Natl Acad Sci U S A* **112**, 8696–8701.
44. Sonestedt E, Roos C, Gullberg B, *et al.* (2009) Fat and carbohydrate intake modify the association between genetic variation in the FTO genotype and obesity. *Am J Clin Nutr* **90**, 1418–1425.
45. Corella D, Arnett DK, Tucker KL, *et al.* (2011) A high intake of saturated fatty acids strengthens the association between the fat mass and obesity-associated gene and BMI. *J Nutr* **141**, 2219–2225.
46. Huang T, Qi Q, Li Y, *et al.* (2014) FTO genotype, dietary protein, and change in appetite: the Preventing Overweight Using Novel Dietary Strategies trial. *Am J Clin Nutr* **99**, 1126–1130.
47. Zhang X, Qi Q, Zhang C, *et al.* (2012) FTO genotype and 2-year change in body composition and fat distribution in response to weight-loss diets: the POUNDS LOST Trial. *Diabetes* **61**, 3005–3011.
48. Gulati P, Cheung MK, Antrobus R, *et al.* (2013) Role for the obesity-related FTO gene in the cellular sensing of amino acids. *Proc Natl Acad Sci U S A* **110**, 2557–2562.
49. Cheung MK, Gulati P, O’Rahilly S, *et al.* (2013) FTO expression is regulated by availability of essential amino acids. *Int J Obes (Lond)* **37**, 744–747.
50. Molerés A, Rendo-Urteaga T, Zulet MA, *et al.* (2012) Obesity susceptibility loci on body mass index and weight loss in Spanish adolescents after a lifestyle intervention. *J Pediatr* **161**, 466–470.e462.
51. Krishnan S & Cooper JA (2014) Effect of dietary fatty acid composition on substrate utilization and body weight maintenance in humans. *Eur J Nutr* **53**, 691–710.
52. Westerterp KR, Smeets A, Lejeune MP, *et al.* (2008) Dietary fat oxidation as a function of body fat. *Am J Clin Nutr* **87**, 132–135.
53. Claussnitzer M, Dankel SN, Kim KH, *et al.* (2015) FTO obesity variant circuitry and adipocyte browning in humans. *N Engl J Med* **373**, 895–907.
54. Smemo S, Tena JJ, Kim KH, *et al.* (2014) Obesity-associated variants within FTO form long-range functional connections with IRX3. *Nature* **507**, 371–375.
55. Srivastava A, Mittal B, Prakash J, *et al.* (2016) Association of FTO and IRX3 genetic variants to obesity risk in north India. *Ann Hum Biol* **43**, 451–456.
56. Bellefroid EJ, Kobbe A, Gruss P, *et al.* (1998) Xiro3 encodes a Xenopus homolog of the Drosophila Iroquois genes and functions in neural specification. *EMBO J* **17**, 191–203.
57. Willett W & Stampfer MJ (1986) Total energy intake: implications for epidemiologic analyses. *Am J Epidemiol* **124**, 17–27.
58. Groppe DM, Urbach TP & Kutas M (2011) Mass univariate analysis of event-related brain potentials/fields II: simulation studies. *Psychophysiology* **48**, 1726–1737.