

1 **Abstract**

2 *Background:* Early clinical trials of a *Clostridium difficile* toxoid vaccine show efficacy in
3 preventing *C. difficile* infection (CDI). The optimal patient group to target for vaccination
4 programmes remains unexplored. This study performed a model-based evaluation of the
5 effectiveness of different CDI vaccination strategies, within the context of existing infection
6 prevention and control strategies such as antimicrobial stewardship.

7 *Methods:* An individual-based transmission model of CDI in a high-risk hospital setting was
8 developed. The model incorporated data on patient movements between the hospital, and
9 catchment populations from the community and long-term care facilities (LTCF), using
10 English national and local level data for model-parameterisation. We evaluated vaccination
11 of: 1) discharged patients who had an CDI-occurrence in the ward; 2) LTCF-residents; 3)
12 Planned elective surgical admissions and 4) All three strategies combined.

13 *Results:* Without vaccination, 10.9 [Interquartile range: 10.0 – 11.8] patients per 1000 ward
14 admissions developed CDI, of which 31% were ward-acquired. Immunising all three patient
15 groups resulted in a 43% [42 – 44], reduction of ward-onset CDI on average. Among the
16 strategies restricting vaccination to one target group, vaccinating elective surgical patients
17 proved most effective (35% [34 – 36] reduction), but least efficient, requiring 146 [133 –
18 162] courses to prevent one ICU-onset case. Immunising LTCF residents was most efficient,
19 requiring just 13 [11 – 16] courses to prevent one case, but considering this only comprised a
20 small group of our hospital population, it only reduced ICU-onset CDI by 9% [8 – 11].
21 Vaccination proved most efficient when ward-based transmission rates and antimicrobial
22 consumption were high.

23 *Conclusions:* Strategy success depends on the interaction between hospital and catchment
24 populations, and importantly, consideration of importations of CDI from outside the hospital

25 which we found to substantially impact hospital dynamics. Vaccination may be most
26 desirable in settings or patient groups where levels of broad-spectrum antimicrobial use are high
27 and difficult to reduce.

28

29 **Introduction**

30 *Clostridium difficile* infection (CDI) is a source of considerable morbidity and
31 mortality and places a substantial burden on healthcare systems[1]. Though traditionally
32 viewed as a healthcare-associated bacteria, intensivied surveillance reveal increasing reports
33 of CDI cases without recent hospitalisation [2]. Antimicrobial stewardship, mandatory
34 surveillance, and enhanced infection prevention and control measures (IPC) to prevent *C.*
35 *difficile* transmission have been implemented in hospitals and the community with success in
36 some countries[3]. Nonetheless there remains the need to prevent CDI in settings and
37 vulnerable patient groups where strict antimicrobial stewardship or IPC is not possible or
38 desirable.

39 Three vaccines targeting the main virulence factors of *C. difficile* (TcdA and TcdB)
40 are currently under development and have showed promising results in phase I and II clinical
41 trials [4–9], with the first Phase III trials now underway[10]. These vaccines induce an IgG
42 antibody response against TcdA and TcdB and therefore aim to prevent the development of
43 symptomatic disease in exposed individuals[e.g. 2,6,7]. A successful vaccine that prevented
44 primary or recurrent onset of CDI would reduce morbidity and mortality directly in the
45 vaccinated individual. It could also have a population-level effect by reducing the spread of
46 infectious spores from infected individuals into the environment, and thus preventing onward
47 transmission of the bacteria. Current evidence, based on highly discriminatory genetic typing-
48 methods[12–14] as well as statistical modelling[15], suggests patients with symptomatic CDI
49 are not the only sources of infection and have pointed to the possible role of asymptomatic
50 carriers. Therefore, any examination of the overall impact of vaccination needs to account for
51 *C. difficile* transmission, including the potential role of asymptomatic carriers [14,16].

52 Previous studies have shown direct healthcare costs due to excess length of hospital
53 stay to be the main driver of infection costs[17,18]. Hospital admissions from LTCF have
54 been associated with increased risk of hospital-onset CDI[19] and residing in LTCF was
55 identified as an independent risk-factor for developing CDI[20]. Hence, this group of
56 individuals are a potential target population for vaccination, as are patients with planned
57 elective surgery who share many of the underlying risk factors (frailty, hospital admission
58 and antimicrobial usage) in common with the LTCF cohort [10]. Moreover, about 20% of
59 CDI patients experience recurrent CDI[21], either due to re-infection or relapse[22], and
60 primarily as a result of continued exposure to factors disturbing the gut flora post
61 identification of CDI[23]. Therefore, patients with a history of CDI are a potential third target
62 group for vaccination.

63 Mathematical modelling is a well-established tool that can be used to extrapolate
64 vaccination trial results to the population-level[24]. In the case of *C. difficile*, these methods
65 would allow the exploration of the impact of vaccination, taking into account the different
66 modes of acquisition[25], as well as the indirect effect of prevention of onward transmission.
67 Therefore, a dynamic transmission model[26–37] was developed to investigate the
68 effectiveness of four vaccination strategies, in terms of preventing CDI in a hospital-ward
69 setting with patients at greater risk of acquiring the infection, such as the Intensive Care Unit
70 (ICU)[38]. Although ICUs can vary markedly in their case mix, their critically ill status often
71 causes a state of immunosuppression[39], and requires high levels of antimicrobial
72 prescribing[40]. The model was designed to capture potential population-effects, as well as
73 uncertainties related to the epidemiology of CDI, and is among the first to incorporate
74 heterogeneous community populations.

75

76 **Methods**

77 *Model framework*

78 A discrete-time, individual-based dynamic transmission model[41] was developed,
79 simulating the transmission and control of CDI in a 30-bed single ICU, serving a community
80 of 100,000 individuals, over a five-year time period. Individual patient movements between
81 the hospital, the surrounding community and LTCF were modelled (Figure 1).
82 Transmission-events were explicitly simulated in the ICU, whereas patients could be
83 admitted and re-admitted from the general community or LTCFs[42], each holding patients
84 with different characteristics (Table 1). If a patient developed symptom-onset post-ICU
85 discharge, the model captured this, however, onward-transmission in the community and
86 LTCFs from these cases was not considered. Likewise, the time spent elsewhere in hospital
87 (and thus the transmission elsewhere in hospital) prior to ICU admission and post-ICU
88 discharge is not captured in the model. However, the importation rates of colonised and
89 infected individuals (a_{i_ltof} , a_{c_ltof} , a_{i_com} , a_{i_com}) were informed by ICU admission data (Table
90 1, Table S5), therefore we implicitly incorporated acquisition during the time spent elsewhere
91 in hospital, as well as readmission of still colonised individuals from outside the hospital.

92 *Transmission process*

93 Patients with normal gut flora were assumed to be protected against *C. difficile* colonisation
94 (compartment P, Figure 1). Although colonisation in healthy individuals with a normal gut
95 flora has been reported, this is likely to be transient, with persistent colonisation among this
96 group found to be rare [43–45]. Moreover, such healthy individuals are at a much lower risk
97 of progressing to symptomatic disease. Consumption of ‘high risk’ antimicrobials (defined as
98 broad-spectrum penicillins, cephalosporins, clindamycin, and quinolones) was assumed to
99 result in susceptibility to colonisation (compartment S) because of their deleterious effect on

100 the microbiota [46]. Each day, susceptible patients (S) could become colonised with *C.*
101 *difficile* through transmission, with the daily risk of colonisation (λ_i) increasing linearly with
102 the number of transmitting CDI patients in the ICU ward (Table 1). The per day probability
103 of colonisation, given at least one CDI or colonised patient on the ward, described the
104 likelihood of transmission through direct contact between susceptible and infectious patients,
105 and indirect contact between susceptible patients, contaminated staff and the environment. As
106 vaccination is unlikely to affect the level of *C. difficile* carriage in the gastrointestinal tract,
107 vaccinated and non-vaccinated colonised individuals were considered to contribute equally to
108 the bacterium's daily probability of colonisation. It was assumed that contacts (with patients,
109 staff or the environment) occurred randomly and were homogeneously distributed among
110 patients.

111 A proportion of patients can mount a natural immune response against *C. difficile*
112 toxins, and are protected from infection[47]. Therefore, a distinction was made between
113 patients that remained asymptomatic (compartment C) and those that suffered from CDI
114 (compartment I) following an incubation period. After successful treatment, patients lost
115 their infection status but remained colonised with *C. difficile*. Colonisation status was lost
116 after an average period of four weeks[48]. To simulate relapse whilst still colonised, the
117 model allowed recovered patients to have another episode of CDI following successive
118 antimicrobial use, but without transmission from another patient. Post-discharge, colonised
119 patients recovered from *C. difficile* colonisation at a constant rate $1/c$, where c is the average
120 duration of colonisation [48,49] in days (Table 1). For individuals with onset post-discharge,
121 this was $1/(s+c)$, with s representing the duration of symptomatic disease [12,50]. Finally,
122 post-vaccination, patients were assumed protected from CDI, but not from
123 colonisation[47,51]. For further model assumptions on bed occupancy, admission and

124 discharge dynamics and transmission dynamics in the community-settings see supplementary
125 material.

126 *Interventions*

127 Compared to no vaccination, we simulated four strategies: 1) patients who have
128 experienced an episode of CDI in the ward, at the time of discharge from the hospital, as they
129 are at risk of experiencing recurrent infection; 2) LTCF residents in the catchment area of the
130 hospital irrespective of whether they are to have planned elective surgery; 3) patients with
131 planned elective surgery in the hospital catchment area and 4) all the above listed patient
132 groups. The strategies involving LTCF residents and elective patients both concerned
133 community-based strategies. All LTCF residents were assumed vaccinated and protected at
134 the start of the simulation and for a period of two years after which a booster vaccine course
135 was provided. Elective patients were vaccinated and protected pre-admission, assuming the
136 time of their appointment being made allowed for enough time to receive vaccination and
137 mount a successful immune response before hospital admission. Finally, ICU-patients that
138 experienced CDI that hospital episode, were vaccinated at the time of hospital discharge, and
139 assumed to be protected from that time onwards for a period of two years to represent waning
140 immunity.

141 *Model parameterisation and validation*

142

143 Table 1 summarises the model parameter values. These values were derived from
144 extensive analysis of national and regional healthcare data and peer-reviewed research
145 articles otherwise. The transmission potential from symptomatic carriers (β_1) and
146 asymptomatic carriers (β_2) in English ICUs is largely unknown. Therefore we estimated
147 these parameters by fitting the model output to CDI acquisition rates as reported in English

148 national ICU audit data [52]. Furthermore, we populated the model with national Hospital
149 Episode statistics data on patient movements[53,54]. Model parameterisation is discussed in
150 further detail in the supplementary material. We validated the model by comparing our model
151 outputs to a list of targets based on external data sources, depicted in supplementary table S7.

152 *Scenario and sensitivity analysis*

153 Due to a lack of knowledge regarding vaccine efficacy and the role of asymptomatic
154 carriers in the transmission of *C.difficile* [14,16], scenario analysis was performed. This
155 incorporated three levels of vaccine efficacy (1, 0.7 and 0.5) and three assumptions for
156 asymptomatic transmission (where asymptomatic carriers transmitted at half the rate of
157 symptomatic carriers (2:1), i.e. the “base case”; no asymptomatic transmission (1:0); and
158 asymptomatic carriers transmitted as efficiently as symptomatic carriers (1:1), Table 2).
159 Analysis of national data (described in more detail in previous publications [52,55]), showed
160 that CDI acquisition rates and ward-based antimicrobial use varied nationally (Table S3).
161 Therefore, two different levels of transmission (baseline and high) and three levels of
162 antimicrobial use (baseline, low and high) were assumed. Here, the baseline scenarios
163 represented the average acquisition and antimicrobial use rates in English hospitals as
164 estimated from national data (Table S2)[40]. Combinations of the above listed possibilities
165 were simulated as listed in Table 2.

166 To account for parameter uncertainty (Table 1), probabilistic sensitivity analysis was
167 performed using Latin hypercube sampling [56] as follows. One thousand random samples
168 were drawn covering the whole range of possible values for each parameter equally and
169 combined at random to create 1000 different parameter sets. As the model was stochastic, a
170 different result could be expected for a given parameter set. Hence the medians of 100

171 simulation runs per parameter set were combined to obtain the overall median and
172 interquartile range (IQR) of the model output encompassing parameter uncertainty.

173 *Model output*

174 The absolute reduction in number of cases per 1000 admissions for each strategy
175 compared to a strategy without vaccination (strategy effectiveness) was evaluated, as well as
176 the number of courses required to avert one case in the ICU (strategy efficiency).

177 **Results**

178 *Simulation Results: Base Case Scenario*

179 No vaccination

180 In the base-case scenario, without vaccination (strategy 0), the median number of
181 ICU-onset cases per 1000 admissions was 10.9 [IQR: 10.0 – 11.8] (Figure 2A). A majority of
182 these cases (69%) were imported from outside the ICU. In total, 14.1 [13.2 – 15.0] ward-
183 acquired (symptomatic and asymptomatic) cases were observed per 1000 admissions (Figure
184 2B). Seventy-nine per cent of acquisitions resulted in symptomatic infection (Figure 2B), but
185 over half developed symptoms post ward discharge (57%), and thus remained asymptomatic
186 whilst in the ICU.

187 Vaccine programme effectiveness & efficiency

188 Vaccinating all target populations (strategy 4) resulted in a 43% [IQR: 42 – 44]
189 reduction in ICU-onset cases over five years, equal to 4.7 [4.3 – 5.1] CDI cases per 1000
190 admissions (Table 3). Reviewing the strategies restricting vaccination to one target group,
191 vaccinating all patients awaiting elective surgery (strategy 3) was the most effective. For all
192 four strategies, vaccination prevented more imported cases than cases acquiring CDI within

193 the ICU (Table 3). This was particularly true for strategy 1 (vaccinating patients with a
194 history of CDI) and 2 (LTCF residents). Strategy 2 proved the most efficient, i.e. required the
195 lowest number of courses to avert one case of ICU-onset CDI in the base case scenario (13
196 [11 – 16]), despite the low effectiveness of this strategy. In contrast, strategy 3 proved highly
197 inefficient, requiring 146 [133 – 162] to prevent on case in the ICU (Table 3).

198 Population-effect of the vaccine

199 We assumed that vaccination did not provide direct protection against *C. difficile*
200 colonisation. Therefore, vaccination is likely to result in an increase in asymptomatic cases.
201 Indeed, the number of asymptomatic acquisitions did increase post-vaccination for strategy 2,
202 3 and 4 (Figure 2B). However, as the drop in symptomatic infections was higher, the total
203 number of acquisitions for these three strategies decreased, indicating a population-effect was
204 present (Figure 2B). Of note, in the ICU alone, this meant that, post-vaccination, a reduction
205 was observed in both symptomatic and asymptomatic acquisitions (Figure 2A) as without
206 vaccination, a large fraction of patients would have developed symptoms post-discharge.

207 *Simulation Results: Scenario analysis*

208 Cross transmission and antimicrobial use

209 The ordering of the most effective and efficient strategies remained unchanged under
210 all the simulated scenarios of transmission and antimicrobial use (Figure 3). All strategies
211 proved most effective and efficient under scenarios of high transmission and high
212 antimicrobial usage (scenario 6, Table S7). In particular, vaccination of elective patients
213 (strategy 3) and therefore vaccination of all target groups (strategy 4) became more efficient,
214 as they were most successful in preventing onward transmission (Figure 3).

215 Impact of asymptomatic carriers

216 The comparative effectiveness and efficiency of each strategy also remained
217 unchanged under different asymptomatic transmission assumptions. Post-vaccination,
218 reduction in ward-based acquisition was greatest when asymptomatic carriers were not
219 transmitting, and marginal when equal transmission between symptomatic and asymptomatic
220 carriers was assumed (Figure S2). As a result, in the scenario without asymptomatic
221 transmission, the marked decrease in ICU-acquisitions resulted in a reduction in
222 asymptomatic carriers in- and also outside the ICU. Under the equal asymptomatic
223 transmission assumption, vaccination resulted in a slight increase of asymptomatic carriers
224 both in- as well as outside the ICU for the most effective strategies 3 and 4 (Figure S3).

225 Impact of vaccine efficacy

226 With vaccine efficacy reduced to 70% (scenario 7), vaccinating all target groups still
227 averted 32% [31 – 33] of the ICU-onset CDI cases (Table S7). This was 24% [23 -25] when
228 efficacy was as low as 50% (scenario 8). However, while the number of vaccine courses
229 required for strategy 2 remained low even under our lowest vaccine efficacy scenario (23 [18
230 – 32]), strategy 3 and 4 became very inefficient, with 281 [251 – 313] and 229 [206 – 255]
231 courses required to prevent one case of CDI in the ICU (Table S7).

232 **Discussion**

233 This study is the first dynamic-transmission model of different vaccination strategies
234 against CDI in a high-risk hospital setting. We observed that immunising all three patient
235 groups (LTCF residents, elective patients and patients with a history of ICU-onset CDI) could
236 prevent up to 43% of CDI-onset cases in our simulated 30-bed ward. With ~17 CDI cases
237 observed annually, representing current average incidence rates in English ICUs[52], this
238 represented the prevention of ~7 ICU-onset cases per year. Of the three individual target

239 groups, vaccinating all patients awaiting elective surgery yielded the largest net reduction in
240 ICU-onset cases.

241 We did not conduct a formal cost-effectiveness evaluation, since costs are likely to be highly
242 specific to a particular setting. However, we did consider efficiency – measured as the
243 number of vaccination courses needed to prevent one case of CDI. In our study, the balance
244 between ward-importations and ward-acquisitions of CDI drove the projected efficiency of
245 vaccination. Previous statistical and molecular studies have questioned the importance of in-
246 hospital transmission from symptomatic patients in the development of CDI in endemic
247 settings[12,13,15], and hint at other acquisition sources. The fitted model suggested that the
248 majority (~70%) of ICU-onset cases were imported from outside the ICU. These importations
249 were primarily asymptomatic admitted patients who developed CDI following antimicrobial
250 treatment. As a result, the identification and targeting of patients groups at heightened risk of
251 colonisation, became increasingly important when hospital-based CDI-onset was not
252 primarily driven by hospital-based acquisitions. Vaccinating LTCF residents would be an
253 example of such target populations [19,20]. We found that vaccinating LTCF residents
254 proved highly efficient in terms of courses per case prevented (13 [11 – 16]), primarily as
255 asymptomatic colonisation is frequent among the elderly residents of LTCF[57], and the high
256 rates of antimicrobial prescribing in this group[58,59] compared to the rest of the population
257 [60,61]. For the least efficient strategy, i.e. vaccinating elective surgery patients, this was 146
258 [133 – 162]. To compare, for the human papilloma virus (HPV) vaccine, it is suggested that
259 about 129 [62] up to 324 [63] young women will need to be vaccinated to prevent one case of
260 cervical cancer, whereas for influenza in individuals >65 years old this is estimated to be 43
261 (95% CI: 16–192) and as high as 3,333 (1,429–12,500) respectively [64]. Importantly, the
262 low total number of admissions from LTCF means that this strategy only resulted in a small
263 reduction in the overall reduction of cases in the ICU. Therefore, the proportion of

264 admissions from the LTCF is an important consideration in the generalisability of our
265 findings.

266 Our scenario analysis revealed that with lower levels of CDI-associated antimicrobial
267 prescribing (e.g. clindamycin and cephalosporins), the efficiency of vaccination was greatly
268 reduced, even under scenarios of high transmission rates, and the converse for high
269 antimicrobial use was also true. Therefore, vaccination may be most efficient (and perhaps
270 cost-effective) in settings where levels of broad-spectrum antimicrobial use are high and
271 difficult to reduce.

272 Another important finding we observed however is that, when asymptomatic carriers
273 contributed to transmission, the number of colonisations outside the ICU increased following
274 vaccination. These asymptomatic cases are more likely to remain undetected than
275 symptomatic cases. When transmission from such individuals is present, this may lead to
276 unintended consequences for the transmission of *C. difficile* and CDI incidence outside the
277 ICU. When asymptomatic carriers were non-transmissible, this increase in colonisations was
278 not present. Recently, a study by Durham and colleagues (2016) estimated that asymptomatic
279 carriers transmitted at a 15 times reduced rate compared to CDI cases in a hospital-wide
280 setting, as well as in the community[37]. Following our scenario analysis, this would suggest
281 that the indirect effects of the vaccine might actually be higher than identified in our study
282 under baseline assumptions of half the rate. Until we are more certain about the role of
283 asymptomatic patients in the transmission of CDI in different settings, it is difficult to define
284 the true effectiveness of vaccination, as well as any infection prevention control strategy.

285 Only one previous modelling study by Lee *et al*, has quantified the impact of CDI
286 vaccination in a comparative manner. They found that a CDI vaccine will most likely be
287 cost-effective in the United States when aimed at preventing recurrent CDI[65]. In our study,

288 vaccinating patients with a history of CDI (in the ICU) had close to no effect on CDI
289 incidence in the ICU, and required ~80 courses to prevent one case. Lee and colleagues
290 assumed that recurrent CDI would always occur in hospital or result in hospitalisation. In our
291 model, active admission of recurrent cases was not incorporated; primarily because such
292 patients are unlikely to be admitted to the ICU. We did incorporate readmission for other
293 reasons however, and in the absence of vaccination, less than one case per 1000 admissions
294 of the patients with a recurrent ICU-onset CDI was seen, either in the same episode or after
295 re-admissions. This was for two reasons: firstly we observed a low number of relapses during
296 the same hospital stay, secondly, the risk of ICU readmission when colonised was low, as the
297 mean colonisation time, as observed by others (e.g. [34]) was approximately similar to the
298 average number of days between ICU-admissions in England. Admittedly, our single-ward
299 model framework did not include re-admission elsewhere in hospital, nor discharge to other
300 hospital wards. Hence these constraints did not allow for a full investigation of this strategy.

301 This study had several other limitations. The calculated number of vaccine courses for
302 strategy 2, 3 and 4 are approximations, and in particularly for strategy 2, was likely to be an
303 underestimate, as we did not account for the high mortality rates among LTCF residents and
304 frequent new admissions to the cohort [66]. Secondly, due to the single-ward framework of
305 our model, it is likely that we have underestimated the impact of vaccination: although we
306 considered importations from and infection-onset post ICU discharge, our model only
307 evaluated CDI-dynamics in the ICU. About 57% of the acquired *C. difficile* in the ICU
308 developed onset after ICU-discharge. This is similar to estimates from active population-
309 based surveillance data from the United States revealing that ~63% of the healthcare-
310 associated CDI cases in 2011 developed symptoms in the community [67]. Onward
311 transmission prevented from these cases with symptom-onset or recurrence outside the ICU
312 was not captured, resulting in a potential underestimation of vaccine effectiveness in terms of

313 preventing hospital- as well as community-onset CDI. Incorporation of discharge and
314 (re)admission dynamics elsewhere in the hospital may have improved the effectiveness of
315 some strategies (notably vaccinating CDI cases) in preventing healthcare-onset CDI.
316 However, data to more realistically inform such a holistic model would have required
317 surveillance data on CDI occurrence in community-settings including the LTCF, which are
318 currently lacking for most countries, including England, as is national-level data on ward
319 movements and ward-specific CDI incidence rates. Therefore, any such model would have
320 been highly theoretical and its results uncertain. Thirdly, our transmission parameters were
321 estimated with all parameters at their baseline value, including antimicrobial use (i.e. the
322 prevalence of usage of traditionally defined high-risk classes [46]). Other classes (e.g.
323 macrolides) have been associated with CDI as well, albeit with much lower risks[46,68].
324 Inclusion of these classes, as well as their heterogeneity in associated CDI-risk, could
325 potentially have resulted in a larger net-number of susceptible individuals at each ICU-day,
326 resulting in lower fitted transmission rates, and subsequently lower population-effects of
327 the vaccine. As this would have affected the strategies involving elective patients and all
328 patient groups in particular, we do not expect this would change our conclusions on the
329 comparative performance of the strategies. Finally, our model did not explicitly consider the
330 time required for seroconversion post-vaccination. All three vaccines under development are
331 considering a 3-dose schedule covering a time period of ~30 days. The latest Phase II clinical
332 trial data found seroconversion rates to peak at 60 days. With average waiting times for
333 elective patients in England of ~70 days, this would support our assumption that elective
334 patients were protected on admission. However, our mean time of ICU readmission was 30
335 days, suggesting that our strategy involving patients with a history of CDI would have been
336 even less effective when accounting for this preliminary findings on the timing of
337 seroconversion. Once more data is available on the optimal dose regimen for the respective

338 vaccines, future modelling research should take both the seroconversion rate and dosing
339 strategies in consideration.

340

341 ***Conclusions***

342 Through careful modelling of the admission and discharge dynamics between
343 healthcare and community settings, this study has provided useful insights as to how and
344 where respective vaccination strategies involving different target groups are most likely to
345 have an impact on CDI incidence rates. Vaccinating LTCF residents and elective patients
346 may aid in preventing CDI in high-risk hospital settings such as the ICU. However, in
347 settings with comparable ICU-acquisition and antimicrobial usage rates to England, this
348 would require a high number of vaccine courses. Therefore, vaccination may be most useful
349 in settings where IPC or reaching low levels of antimicrobial usage proves challenging.

350 **Conflict of interest statement**

351 None.

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References

1. Bauer MP, Notermans DW, van Benthem BHB, Brazier JS, Wilcox MH, Rupnik M, et al. Clostridium difficile infection in Europe: a hospital-based survey. Lancet. Elsevier Ltd; 2011 Jan 1;377(9759):63–73.
2. Chitnis AS, Holzbauer SM, Belflower RM, Winston LG, Bamberg WM, Lyons C, et al. Epidemiology of Community-Associated Clostridium difficile Infection, 2009 Through 2011. JAMA Intern Med. 2013 Jul 22;173(14):1359–67.
3. Public Health England (former Health Protection Agency). Results from the mandatory Clostridium difficile reporting scheme. London; 2013.
4. Bezay N, Ayad A, Dubischar K, Firbas C, Hochreiter R, Kiermayr S, et al. Safety, immunogenicity and dose response of VLA84, a new vaccine candidate against Clostridium difficile, in healthy volunteers. Vaccine. Elsevier Ltd; 2016;34(23):2585–92.
5. de Bruyn G, Saleh J, Workman D, Pollak R, Elinoff V, Fraser NJ, et al. Defining the optimal formulation and schedule of a candidate toxoid vaccine against Clostridium difficile infection: A randomized Phase 2 clinical trial. Vaccine. Elsevier Ltd; 2016;34(19):2170–8.
6. Foglia G, Shah S, Luxemburger C, Freda PJ. Clostridium difficile : Development of a novel candidate vaccine. Vaccine. 2012;30:4307–9.
7. Greenberg RN, Marbury TC, Foglia G, Warny M. Phase I dose finding studies of an adjuvanted Clostridium difficile toxoid vaccine. Vaccine. Elsevier Ltd; 2012 Mar 16;30(13):2245–9.
8. Kotloff KL, Wasserman SS, Genevieve A, Jr WT, Nichols R, Bridwell M, et al. Safety and Immunogenicity of Increasing Doses of a Clostridium difficile Toxoid Vaccine Administered to Healthy Adults Safety and Immunogenicity of Increasing Doses of a Clostridium difficile Toxoid Vaccine Administered to Healthy Adults. Infect Immun. 2001;69(2):988.
9. Sheldon E, Kitchin N, Peng Y, Eiden J, Gruber W, Johnson E, et al. A phase 1, placebo-controlled, randomized study of the safety, tolerability, and immunogenicity

- of a *Clostridium difficile* vaccine administered with or without aluminum hydroxide in healthy adults. *Vaccine*. 2016;34(18):2082–91.
10. Sanofi pasteur. Sanofi Pasteur Investigational Vaccine against *Clostridium difficile* Fact Sheet [Internet]. 2013 [cited 2013 Oct 8]. Available from: <http://www.multivu.com/assets/62652/documents/62652-SP-C-diff-Vaccine-Fact-Sheet-FINAL-8-2-13-original.pdf>
 11. Ward SJ, Douce G, Dougan G, Wren BW. Local and systemic neutralizing antibody responses induced by intranasal immunization with the nontoxic binding domain of toxin A from *Clostridium difficile*. *Infect Immun*. 1999 Oct;67(10):5124–32.
 12. Walker A, Eyre D, Wyllie D, Dingle K, Harding R, O'Connor L, et al. Characterisation of *Clostridium difficile* Hospital Ward-Based Transmission Using Extensive Epidemiological Data and Molecular Typing. *PLoS Med*. 2012 Feb;9(2):e1001172.
 13. Eyre DW, Cule ML, Wilson DJ, Griffiths D, Vaughan A, O'Connor L, et al. Diverse Sources of *C. difficile* Infection Identified on Whole-Genome Sequencing. *N Engl J Med*. 2013 Sep 26;369(13):1195–205.
 14. Curry SR, Muto C a, Schlackman JL, Pasculle a W, Shutt K a, Marsh JW, et al. Use of multilocus variable number of tandem repeats analysis genotyping to determine the role of asymptomatic carriers in *Clostridium difficile* transmission. *Clin Infect Dis*. 2013 Oct;57(8):1094–102.
 15. van Kleef E, Gasparri A, Guy R, Cookson B, Hope R, Jit M, et al. Nosocomial transmission of *C. difficile* in English hospitals from patients with symptomatic infection. *PLoS One*. 2014 Jan;9(6):e99860.
 16. Eyre DW, Griffiths D, Vaughan A, Golubchik T, Acharya M, O'Connor L, et al. Asymptomatic *Clostridium difficile* colonisation and onward transmission. *PLoS One*. 2013 Jan;8(11):e78445.
 17. Dubberke ER, Olsen MA. Burden of *Clostridium difficile* on the healthcare system. *Clin Infect Dis*. United States: Department of Medicine, Washington University School of Medicine, St Louis, Missouri, USA. edubberk@dom.wustl.edu; 2012;55 Suppl 2:S88–92.

18. McGlone SM, Bailey RR, Zimmer SM, Popovich MJ, Tian Y, Ufberg P, et al. The economic burden of *Clostridium difficile*. *Clin Microbiol Infect*. 2012 Mar;18(3):282–9.
19. Ricciardi R, Nelson J, Griffith JL, Concannon TW. Do admissions and discharges to long-term care facilities influence hospital burden of *Clostridium difficile* infection? *J Hosp Infect*. Elsevier Ltd; 2012 Feb;80(2):156–61.
20. Vesteyndottir I, Gudlaugsdottir S, Einarsdottir R, Kalaitzakis E, Sigurdardottir O, Bjornsson ES. Risk factors for *clostridium difficile* toxin-positive diarrhea: A population-based prospective case-control study. *Eur J Clin Microbiol Infect Dis*. 2012;31(10):2601–10.
21. Eyre DW, Walker a S, Wyllie D, Dingle KE, Griffiths D, Finney J, et al. Predictors of first recurrence of *Clostridium difficile* infection: implications for initial management. *Clin Infect Dis*. 2012 Aug;55 Suppl 2(Suppl 2):S77–87.
22. Wilcox M., Fawley WN, Settle CD, Davidson A. Recurrence of symptoms in *Clostridium difficile* infection - relapse or reinfection? *Hosp Infect Soc*. 1998;38:93–100.
23. Garey KW, Sethi S, Yadav Y, DuPont HL. Meta-analysis to assess risk factors for recurrent *Clostridium difficile* infection. *J Hosp Infect*. 2008;70(4):298–304.
24. Kim S-Y, Goldie SJ. Cost-effectiveness analyses of vaccination programmes : a focused review of modelling approaches. *Pharmacoeconomics*. 2008;26(3):191–215.
25. Anderson RM, May RM. *Infectious Diseases of Humans*. Oxford University Press; 1991.
26. Grigoras CA, Zervou FN, Zacharioudakis IM, Siettos CI, Mylonakis E. Isolation of *C. difficile* Carriers Alone and as Part of a Bundle Approach for the Prevention of *Clostridium difficile* Infection (CDI): A Mathematical Model Based on Clinical Study Data. 2016;(Cdi):1–12.
27. Lanzas C, Dubberke ER, Lu Z, Reske KA, Grohn YT. Epidemiological model for *Clostridium difficile* transmission in healthcare settings. *Infect Control Hosp Epidemiol*. 2011;32(6):553–61.

28. Lanzas C, Dubberke ER. Effectiveness of Screening Hospital Admissions to Detect Asymptomatic Carriers of *Clostridium difficile*: A Modeling Evaluation. *Infect Control Hosp Epidemiol*. 2014 Aug;35(8):1043–50.
29. Lofgren ET, Moehring RW, Anderson DJ, Weber DJ, Fefferman NH, Hill C, et al. A Mathematical Model to Evaluate the Routine Use of Fecal Microbiota Transplantation to Prevent Incident and Recurrent *Clostridium difficile* Infection. *Infect Control Hosp Epidemiol*. 2015;35(1):18–27.
30. Nelson RE, Jones M, Leecaster M, Samore MH, Ray W, Huttner A, et al. An Economic Analysis of Strategies to Control *Clostridium Difficile* Transmission and Infection Using an Agent-Based Simulation Model. *PLoS One*. 2016;11(3):e0152248.
31. Rubin M a, Jones M, Leecaster M, Khader K, Ray W, Huttner A, et al. A simulation-based assessment of strategies to control *clostridium difficile* transmission and infection. *PLoS One*. 2013 Jan;8(11):e80671.
32. Starr JM, Campbell A. Mathematical modeling of *Clostridium difficile* infection. *Clin Microbiol Infect*. 2001;7(8):432–7.
33. Starr JM, Rogers TR, Impallomeni M. Hospital-acquired *Clostridium difficile* diarrhoea and herd immunity. *Lancet*. 1997;349(9049):426–8.
34. Starr JM, Campbell A, Renshaw E, Poxton IR, Gibson GJ. Spatio-temporal stochastic modelling of *Clostridium difficile*. *J Hosp Infect*. 2009;71(1):49–56.
35. Yakob L, Riley T V, Paterson DL, Marquess J, Clements AC. Assessing control bundles for *Clostridium difficile*: a review and mathematical model. *Emerg Microbes Infect*. 2014 Jun 18;3(6):e43.
36. Yakob L, Riley T V, Paterson DL, Clements AC. *Clostridium difficile* exposure as an insidious source of infection in healthcare settings: an epidemiological model. *BMC Infect Dis*. *BMC Infectious Diseases*; 2013;13(1):376.
37. Durham DP, Olsen MA, Dubberke ER, Galvani AP, Townsend JP. Quantifying transmission of *Clostridium difficile* within and outside healthcare settings. *Emerg Infect Dis*. 2016;22(4):608–16.
38. Karanika S, Paudel S, Zervou FN, Grigoras C, Zacharioudakis IM, Mylonakis E.

- Prevalence and Clinical Outcomes of Clostridium difficile Infection in the Intensive Care Unit: A Systematic Review and Meta-Analysis. *Open forum Infect Dis.* 2015;3(1):ofv186.
39. Markwart R, Condotta S a., Requardt RP, Borken F, Schubert K, Weigel C, et al. Immunosuppression after Sepsis: Systemic Inflammation and Sepsis Induce a Loss of Naïve T-Cells but No Enduring Cell-Autonomous Defects in T-Cell Function. *PLoS One.* 2014;9(12):e115094.
 40. Public Health England (former Health Protection Agency). English National Point Prevalence Survey on Healthcare-associated Infections and Antimicrobial Use , 2011 - Appendices. 2011.
 41. Jit M, Brisson M. Modelling the Epidemiology of Infectious Diseases for Decision Analysis A Primer. 2011;29(5):371–86.
 42. Fuller C, Robotham J, Savage J, Deeny S, Hopkins S, Cookson B, et al. The National One Week Prevalence Audit of MRSA Screening. Dept. of Health Report. 2013;(March).
 43. Johnson S, Homami SR, Bcttin KM, Quick JN, Clabots CR, Peterson LR, et al. Treatment of Asymptomatic Clostridium difficile Carriers (Fecal Excretors) with Vancomycin or Metronidazole. *Ann Med.* 1992;117(4):297–302.
 44. Galdys AL, Nelson JS, Shutt K a, Schlackman JL, Pakstis DL, Pasculle a W, et al. Prevalence and Duration of Asymptomatic Clostridium difficile Carriage Among Healthy Subjects in Pittsburgh, Pennsylvania. *J Clin Microbiol.* 2014 Apr 23;(April).
 45. Ozaki E, Kato H, Kita H, Karasawa T, Maegawa T, Koino Y, et al. Clostridium difficile colonization in healthy adults: transient colonization and correlation with enterococcal colonization. *J Med Microbiol.* 2004 Feb 1;53(2):167–72.
 46. National Institute for Health and Care Excellence. Clostridium difficile infection: risk with broad-spectrum antibiotics. *NICE Guidel.* 2015;(March):1–30.
 47. Kyne L, Warny M, Qamar A, Ciaran P. Asymptomatic carriage of clostridium difficile and serum levels of IgG antibody against toxin A. *N Engl J Med.* 2000;10:390–7.
 48. Abujamel T, Cadnum JL, Jury L a, Sunkesula VCK, Kundrapu S, Jump RL, et al.

- Defining the vulnerable period for re-establishment of *Clostridium difficile* colonization after treatment of *C. difficile* infection with oral vancomycin or metronidazole. *PLoS One*. 2013 Jan;8(10):e76269.
49. Clabots CR, Johnson S, Olson MM, Peterson LR, Gerding DN. Acquisition of *Clostridium difficile* by hospitalized patients: evidence for colonized new admissions as a source of infection. *J Infect Dis*. 1992 Sep;166(3):561–7.
 50. Teasley DG, Olson MM, Gebhard RL, Gerding DN, Peterson LR, Schwartz MJ, et al. Prospective randomised trial of metronidazole versus vancomycin for clostridium-difficile-associated diarrhoea and colitis. *Lancet*. 1983;322(8358):1043–6.
 51. Siddiqui F, O'Connor JR, Nagaro K, Cheknis A, Sambol SP, Vedantam G, et al. Vaccination with parenteral toxoid B protects hamsters against lethal challenge with toxin A-negative, toxin B-positive clostridium difficile but does not prevent colonization. *J Infect Dis*. 2012;205(1):128–33.
 52. ICNARC. Key statistics from the Case Mix Programme. London, England; 2014.
 53. Health & Social Care Information Centre. National Statistics Hospital Episode Statistics, Admitted Patient Care, England - 2013-14 [NS] [Internet]. HSCIC. 2015. Available from: <http://www.hscic.gov.uk/catalogue/PUB16719>
 54. Health & Social Care Information Centre. Adult Critical Care Data in England - April 2013 to March 2014 [Internet]. HSCIC. 2015. Available from: <http://www.hscic.gov.uk/searchcatalogue?q=title:“Adult+Critical+Care+data+in+England”&size=10&sort=Relevance>
 55. Public Health England (former Health Protection Agency). English National Point Prevalence Survey on Healthcare-associated Infections and Antimicrobial Use, 2011 - preliminary data. London; 2012.
 56. McKay M, Beckman R, Conover W. A Comparison of Three Methods for Selecting Values of Input Variables in the Analysis of Output from a Computer Code. *Technometrics*. 1979;21(2):239–45.
 57. Ziakas PD, Zacharioudakis IM, Zervou FN, Grigoras C, Pliakos EE, Mylonakis E. Asymptomatic Carriers of Toxigenic *C. difficile* in Long-Term Care Facilities: A

- Meta-Analysis of Prevalence and Risk Factors. *PLoS One*. 2015;10(2):e0117195.
58. European Centre for Disease Prevention and Control. Point prevalence survey of healthcare-associated infections and antimicrobial use in European long-term care facilities - April - May 2013. Stockholm: European Centre for Disease Prevention and Control; 2013.
 59. European Centre for Disease Prevention and Control. Point prevalence survey of healthcare-associated infections and antimicrobial use in European long-term care facilities - May - September 2010. Stockholm; 2010.
 60. European Centre for Diseases Control. Surveillance of antimicrobial consumption in Europe 2010. Sweden: European Centre for Disease Prevention and Control; 2013.
 61. European Centre for Disease Prevention and Control. Surveillance of antimicrobial consumption in Europe 2011. Sweden: European Centre for Disease Prevention and Control; 2014.
 62. Sawaya GF, Smith-mccune K, Ph D. HPV Vaccination — More Answers , More Questions. *N Engl J Med*. 2007;356:1991–4.
 63. Brisson M, Van de Velde N, De Wals P, Boily M-C. Estimating the number needed to vaccinate to prevent diseases and death related to human papillomavirus infection. *Can Med Assoc J*. 2007;177(5):464–8.
 64. Kelly H, Attia J, Andrews R, Heller RF. The number needed to vaccinate (NNV) and population extensions of the NNV: Comparison of influenza and pneumococcal vaccine programmes for people aged 65 years and over. *Vaccine*. 2004;22(17-18):2192–8.
 65. Lee BY, Popovich MJ, Tian Y, Bailey RR, Ufberg PJ, Wiringa AE, et al. The potential value of *Clostridium difficile* vaccine: an economic computer simulation model. *Vaccine*. Elsevier Ltd; 2010 Jul 19;28(32):5245–53.
 66. Forder A. A brief history of infection control - past and present. *South African Med J*. 2007;97(11):1161–4.
 67. Lessa F, Yi Mu M, Bamberg W, Beldavs Z, Dumyati G, Dunn J, et al. Burden of *Clostridium difficile* Infection in the United States. *N Engl J Med*. 2015;372:825–34.

68. Aldeyab MA, Harbarth S, Vernaz N, Kearney MP, Scott MG, Funston C, et al. Quasiexperimental study of the effects of antibiotic use, gastric acid-suppressive agents, and infection control practices on the incidence of *Clostridium difficile*-associated diarrhea in hospitalized patients. *Antimicrob Agents Chemother.* 2009;53(5):2082–8.

Figure legends

Figure 1: Model framework

P = Patients protected from colonisation, hence infection; **S** = Patients susceptible to colonisation, hence infection; **C_{imm}** = Patients colonised with *C. difficile* that are protected from disease due to natural immunity or vaccination; **C_{n_imm}** = Patients colonised with *C. difficile* not protected from disease whilst failing to mount natural immunity or immunity following vaccination; **I** = Patients with CDI; **LTCF** = Long-term care facility

Figure 2: Absolute number of cases averted per 1000 admissions shown for all four vaccination strategies; A) ICU-acquisitions and importations with onset in the ICU only; B) ICU-acquisitions with onset in- & outside the ICU.

Model outcomes at baseline for **strategy 0** (no vaccination); and number of cases averted under **strategy 1** (CDI history); **strategy 2** (LTCF residents); **strategy 3** (elective patients) and **strategy 4** (all combined). The middle line in the box represents the median difference of 1000 model parameter sets between strategy 0 and each vaccination strategy, and upper and lower areas of the box indicate the seventy-fifth and twenty-fifth percentiles.

Figure 3: Absolute number of imported and acquired cases averted per 1000 admissions in the ICU for the vaccination strategies under scenarios 1 to 6.

Left panels: baseline transmission scenarios with A) Baseline; C) Low; E) High antimicrobial use. Right panels: high transmission transmission scenarios with B) Baseline; D) Low; F) High antimicrobial use. Black points: median absolute number ICU-acquired cases averted (x-axis) and imported cases averted (y-axis) of the 1000 parameter sets under the different vaccination strategies. Transparent ellipses plot the 95% coverage intervals.

Table legends

Table 1: Model parameters and assumptions

* Included in probabilistic sensitivity analysis; # Included in scenario analysis. PPS = Point prevalence survey data (reference provided refers to which point prevalence data); H= Individual hospital data; HES = Hospital Episode Statistics; A = Assumption, CQC = Care Quality Commission data

Table 2: Simulated scenarios

2:1 = asymptomatic carriers transmitted at half the rate of symptomatic carriers, the “base case”; 1:0 = asymptomatic carriers did not spread *C. difficile*; 1:1 = asymptomatic carriers transmitted as efficiently as symptomatic carriers.

Table 3: Number of ICU-onset cases averted per 1000 admissions and courses required to avert 1 case of ICU-onset CDI for strategies in the base case scenario

Note NA: under some of the LHS parameter values, the CDI history strategy had no impact