

CD4⁺CD25⁺ Regulatory Cells Contribute to the Regulation of Colonic Th2 Granulomatous Pathology Caused by Schistosome Infection

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Abstract

Eggs of the helminth *Schistosoma mansoni* accumulate in the colon following infection and generate Th2-biased inflammatory granulomas which become down-modulated in size as the infection proceeds to chronicity. However, although CD4⁺CD25⁺FoxP3⁺ regulatory T cells (T_{regs}) are known to suppress Th1-mediated colitis, it is not clear whether they control Th2-associated pathologies of the large intestine which characterise several helminth infections. Here we used a novel 3D-multiphoton confocal microscopy approach to visualise and quantify changes in the size and composition of colonic granulomas at the acute and chronic phases of *S. mansoni* infection. We observed decreased granuloma size, as well as reductions in the abundance of DsRed⁺ T cells and collagen deposition at 14 weeks (chronic) compared to 8 weeks (acute) post-infection. Th2 cytokine production (*i.e.* IL-4, IL-5) in the colonic tissue and draining mesenteric lymph node (mLN) decreased during the chronic phase of infection, whilst levels of TGF-β1 increased, co-incident with reduced mLN proliferative responses, granuloma size and fibrosis. The proportion of CD4⁺CD25⁺FoxP3⁺T_{regs}: CD4⁺ cells in the mLN increased during chronic disease, while within colonic granulomas there was an approximate 4-fold increase. The proportion of CD4⁺CD25⁺FoxP3⁺T_{regs} in the mLN that were CD103⁺ and CCR5⁺ also increased indicating an enhanced potential to home to intestinal sites. CD4⁺CD25⁺ cells suppressed antigen-specific Th2 mLN cell proliferation *in vitro*, while their removal during chronic disease resulted in significantly larger granulomas, partial reversal of Th2 hypo-responsiveness and an increase in the number of eosinophils in colonic granulomas. Finally, transfer of schistosome infection-expanded CD4⁺CD25⁺T_{regs} down-modulated the development of colonic granulomas, including collagen deposition. Therefore, CD4⁺CD25⁺FoxP3⁺T_{regs} appear to control Th2 colonic granulomas during chronic infection, and are likely to play a role in containing pathology during intestinal schistosomiasis.

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Introduction

Schistosomiasis is an important parasitic helminth disease afflicting more than 200 million people, causing approximately 280 thousand deaths annually, with a further estimated 700 million at risk of infection [1,2]. In the case of *Schistosoma mansoni*, infections are typically chronic (>10 years) and the majority (>90%) give rise to an intestinal form of disease [3] caused by the deposition of parasite eggs in the intestinal mesenteries (mainly of the colon and terminal ileum) and the subsequent development of Th2-associated granulomatous infiltrates rich in macrophages and eosinophils [4]. Such infections lead to diarrhoea, pseudopolypoidosis, microulceration, bleeding and fibrosis [5]. Recent reappraisal of Disability-Associated Life Years (DALYs) attributable to schistosomiasis, where more subtle disease manifestations such as intestinal schistosomiasis have been included, raises the disease burden caused by this infection as much as 40-fold, putting schistosomiasis on a par with malaria as a global public health problem [6]. Variation in granuloma size in the colon between

patients is positively associated with peripheral blood mononuclear cell (PBMC) reactivity to soluble egg antigens (SEA) [7]. Thus, changes in lymphocyte responsiveness appear to be related to the size of granulomas in the intestine and by implication, the severity of pathologies in patients with intestinal disease.

In order to investigate the phenomenon of Th2-associated colonic inflammation and possible mechanisms underlying its regulation, we utilized a murine model of infection with *S. mansoni* which provides a well accepted permissive experimental host. In the murine model, myeloid antigen presenting cells, including dendritic cells [8,9], and basophils [10], are primed to induce potent anti-egg Th2 CD4⁺ lymphocyte responses. Th2 activation appears necessary to protect the host from lethal hepatic and intestinal damage during acute infection [11] and to keep Th1 inflammatory immunopathology in check [12]. However, survival to the chronic stage of infection, representative of human disease, is dependent on modulation of the Th2 granulomatous response in order to subvert IL-4/IL-13-driven morbidity [13]. 'Naturally occurring' (n)T_{regs} bearing the IL-2 receptor α chain molecule

Author Summary

Schistosomiasis is an important parasitic helminth disease afflicting more than 200 million people worldwide. Infections are typically chronic and in the case of *Schistosoma mansoni* and *S. japonicum* the majority give rise to an intestinal form of disease caused by the deposition of parasite eggs in the colon and terminal ileum. The eggs cause Th2-associated inflammatory immune granulomas to form, which as the disease develops, are down-regulated by cells of the immune system. However, the mechanisms which underpin the down-regulation of granulomas in the large intestine are not known. In order to investigate the phenomenon of Th2-associated colonic inflammation, we utilized a murine model of infection with *S. mansoni* and compared immune responses at the acute and chronic phases of infection. We show that a type of regulatory T helper lymphocyte (CD4⁺CD25⁺FoxP3⁺T_{reg}) contributes to regulation of colonic inflammation. These cells modulate anti-egg Th2 responses within the mesenteric lymph nodes and granulomatous pro-fibrotic Th2 responses within the colon. Our study highlights the importance of CD4⁺CD25⁺FoxP3⁺T_{regs} as a source of regulatory pressure on granuloma formation in the colon and by implication humans with chronic intestinal schistosomiasis.

(CD25) and expressing the transcription factor forkhead box P3 (FoxP3) have been demonstrated to play a role in the regulation of Th2 anti-egg hepatic inflammation in an IL-10-independent manner [14,15], although their role in regulating intestinal inflammation induced by egg deposition has not been determined.

Our data presented herein support a role for CD4⁺CD25⁺FoxP3⁺T_{regs} in regulating colonic inflammation by modulating both anti-egg Th2 responses within the mesenteric lymph nodes (mLN) and granulomatous, pro-fibrotic Th2 responses within the colon. Thus, our study implicates CD4⁺CD25⁺FoxP3⁺T_{regs} as a source of regulatory pressure during chronic intestinal schistosomiasis and in the wider context, as suppressors of Th2-driven pathology in the colon.

Materials and Methods

Ethics statement

All experiments were carried out in accordance with UK Animal's Scientific Procedures Act 1986 and with the approval of The University of York Ethics Committee.

Experimental infection and parasitological readout

C57BL/6 (B.6) and hCD2-VaDsRed-B.6 mice were maintained within the University of York under specific pathogen-free conditions. hCD2-DsRed-B6 mice, were a gift of D. Kioussis and A. Patel (National Institute for Medical Research, London) and express fluorescent DsRed T cells (>90% CD3⁺) to facilitate in situ detection of T cells by multiphoton microscopy (see below). Eight to ten-week female mice were infected percutaneously via the abdomen with 25 *S. mansoni* cercariae, and infections allowed to mature for either 8 or 14 weeks representing the acute and chronic phases of infection respectively. Adoptive transfer recipients were infected with 100 cercariae. Egg burdens in the 5 cm of colon proximal to the cecum were enumerated following digestion in 4% KOH. Eggs in faecal material were enumerated following dispersion in PBS, filtration through 100 µm pore mesh, and concentration. Colonic granulomas were isolated as previously

described [16]. Volumes were calculated by measuring the longest and widest points and extrapolating volume using standard formulae for sphere or cylinder, depending on individual granuloma shape.

Histology and confocal microscopy

Colonic tissue were fixed in 4% formaldehyde and embedded in wax. Transverse cross-sections (5 µm) were stained with H&E, or haematoxylin and Van Geison (Department of Veterinary Pathology, University of Liverpool). Digital photomicrographs were analysed using AxioVision software (Zeiss).

For multiphoton imaging, proximal colon segments were mounted within 10 mm depression slides, and granulomas imaged from the serosal surface to egg mid-point using a 510 NLO laser-scanning microscope (LSM, Zeiss) with multi-photon laser (Coherent) tuned to 872 nm. 3D projections of 'half-granulomas' were rendered from z stacks using Volocity 4 software (Improvision). Quantification of Ds-Red⁺ lymphocytes, granuloma and collagen volumes were performed using "ROI" and "RGB" measurement tools within Volocity.

For immunofluorescent staining, frozen tissues were cryosectioned at 8 µm intervals, fixed with 10% methanol, permeabilised with 0.5% saponin (Sigma), and blocked with 5% rabbit serum / 1% FCS. Sections were labelled with anti-CD4 AF488 and anti-FoxP3 AF647 (both eBioscience) and fluorescence captured using the 510 NLO LSM. Settings for acute and chronic fluorescence images are matched both with respect to laser scanning settings at the time of image capture and post-image digital enhancement. Baseline laser scanning settings were undertaken on isotype controls and resultant negative control images contain undetectable fluorescent signal.

Anti-CD25 mAb treatment

Three doses of anti-CD25 mAb (50 µg; clone PC61, a gift from F. Powrie, University of Oxford), or purified rat IgG2a, were delivered intraperitoneally to infected mice at 9, 11, and 13 weeks.

T_{reg} cell purification, adoptive transfer and *in vitro* culture

T_{regs} from the mLN were purified by depletion of non-CD4⁺ cells followed by isolation of CD25⁺ cells using antibodies conjugated to magnetic beads (Miltenyi Biotec). For adoptive transfer, 2.5×10⁶ CD4⁺CD25⁺T_{regs} (>90% purity) were injected via the lateral tail vein. Total mLN cells (2×10⁶/ml), sorted CD4⁺CD25⁻ effector cells (1×10⁶/ml), and CD4⁺CD25⁺T_{regs} (0.5×10⁶/ml) from infected mice cultured in complete RPMI-1640 medium (containing 10% FCS, 50 µg/ml penicillin/streptomycin), in combination with naïve mLN CD4⁻CD25⁻ cells (0.1×10⁶/ml) as a source of APC. Cells were stimulated with plate-bound anti-CD3 mAb (1 µg; Becton Dickinson), or SEA (50 µg/ml) [16]. Cells were cultured for 72 h and supernatants retained for cytokine analysis. Proliferation was measured from 72 to 96 h by ³H-thymidine incorporation and scintillation counting.

Cytokine and collagen quantifications

ELISAs were used to quantify IL-4, IL-5 and IFNγ [17], while IL-10 and IL-13 were measured by Cytoset (Invitrogen) or DuoSet (R&D Systems) kits respectively. A TGFβ-sensitive, mink lung epithelial cell bio-assay (MLEC transfected with firefly luciferase; gift from Daniel Rifkin, NY Medical Center) was used to determine levels of bio-active TGFβ1 [18]. As the bio-assay was not compatible with tissue extracts, a TGFβ1 ELISA (R&D Systems) was employed. In order to determine cytokine levels in the colon, frozen tissues were first homogenised in proprietary

tissue extraction buffer containing detergent and protease inhibitors (Thermo Scientific) and then incubated/rotated overnight at 4°C and the soluble fractions isolated by centrifugation prior analysis by ELISA. Salt-soluble collagen was quantified using colorimetric assay (Sircol, Biocolor).

Quantitative Real Time PCR

Total colonic mRNA was used to generate cDNA using Superscript III DNA polymerase (Invitrogen) and *foxp3* transcript analysed by qRT-PCR (ABI PRISM 7000; Applied Biosystems) using Taqman probes (Sigma-Aldrich). The relative expression of *foxp3* was normalised to values obtained for *cd3*. Primer pairs and probes were; *foxp3* 5'-GCAGTGTGGACCGTAGATGA, 5'-CACAGCCTCAGTCTCATGGT, Probe 5'-ACAAGTGCTC-CAATCCCTGCCCTT and *cd3* 5'-GAGCACCTGCTACTC-CTTG, 5'-ATGTCCCAGCACTGGCTACT, Probe 5'-TGC-TCTTCAGCCTCCTGGTGAACAC.

Flow Cytometry

Cells were blocked with anti-CD16/CD32 (eBioscience) at 0.5 µg / 1×10⁶ cells, then labelled with anti-CD4-Pacific Blue, anti-CD25-APC (PC-61), anti-CD103-PE (all eBioscience), anti-CD25-FITC (7D4), anti-CTLA-4-FITC, or anti-CCR5-biotin (BD Bioscience) for 30 minutes. Biotinylated antibodies were sequentially detected with streptavidin-PE-Cy7 (eBioscience). For intracellular staining of FoxP3, cells were fixed in 1% formalin, re-suspended in permeabilisation buffer (Becton Dickinson) prior to labelling with anti-FoxP3-PE or -AF647 (eBioscience). Cells were analysed using a Cyan flow cytometer with Summit software (Beckman Coulter).

Statistical analyses

Significant differences between two experimental groups were determined by unpaired Student's T test, and between three or more groups by 1-way ANOVA with Tukey post-hoc tests using Prism software (GraphPad). Because colonic egg counts were skewed, analysis was undertaken after Log₁₀ transformation. All data are representative of a minimum of two independent experiments. Significance is indicated ***P<0.001, **P<0.01, *P<0.05. Significance values are shown on the figures with line connectors between the appropriate groups. Where statistical significance was not achieved (P>0.05), figures are intentionally left blank.

Results

Colonic granuloma size and anti-egg Th2 responses decline with chronicity of infection

Egg deposition and anti-egg granulomatous responses in proximal colons were examined over a time-course of infection in B.6 mice. The numbers of eggs increased during infection from 292±121.8 (day 42) to 2339±863.5 (day 98) (Fig. 1A). Mean areas of isolated colonic granulomas declined from the acute to chronic time point visualised by H&E staining (Fig. 1B & C), supporting previous observations [16,19]. Estimates of volumes of granulomas isolated from enzymatically digested colons (Fig. 1D) corroborated histological observation and showed a significant decrease in size between the acute and chronic stage of infection. In addition, the decrease in granuloma size at the chronic stage was accompanied by a decrease in collagen, indicative of fibrosis, as shown by Van-Geison stained sections (Fig. 1E).

Multiphoton imaging of proximal colon derived from infected hCD2-VaDsRed-B.6 mice revealed further quantitative information on temporal granuloma modulation in situ (Fig. 1F, Videos S1

& S2) and analysis of 3D images showed granuloma volumes were significantly decreased at the chronic stage (Fig. 1G). Furthermore, numbers of granuloma-associated DsRed⁺ lymphocytes (Fig 1H; >90% CD3⁺T lymphocytes, data not shown), and granuloma-associated type-1 collagen deposition, revealed as second harmonic imaging (blue), was significantly also reduced (Fig. 1I). Although a significant increase in the recently synthesised (salt-soluble) collagen pool within the colon was apparent by the chronic phase, modulation of the egg-driven fibrotic response was demonstrable when adjusted for the increased numbers of eggs (as a surrogate for numbers of granulomas) in chronic infected colons (Fig. 1J).

At the acute stage, anti-CD3 mAb and SEA-specific proliferation of mLN cells were significantly elevated and biased towards secreting Th2-type cytokines (Fig. 2A). However, by the chronic stage, SEA-induced cell proliferation and production of IL-4, IL-5 and IL-13 were significantly reduced (Fig. 2B). While the secretion of IL-10 in response to SEA was significantly lower during chronic compared to acute infection the production of bio-active TGFβ1 to SEA at the chronic stage was significantly elevated compared to naïve state (Fig. 2B).

It was not possible to obtain sufficient numbers of viable lymphocytes via enzymatic digestion of granulomatous colons due to the fibrotic nature of these intestinal granulomas but levels of IL-4 and IL-5 in whole colonic extracts were elevated at the acute phase (Fig. 2C), suggesting that tissue inflammatory responses in infected colons mirrored the Th2 response in the mLN. Surprisingly, levels of IL-10 significantly decreased in infected colonic tissue (Fig. 2C) but levels of bio-active colonic TGFβ1 were elevated during chronic disease. When adjusted for numbers of deposited eggs, production of colonic IL-4 and IL-5 were significantly diminished at the chronic phase (Fig. 2D). Thus, whilst local cytokine responses to egg deposition has both shared and distinct facets to those of the mLN, measurements indicate that colonic Th2 responses establish during the onset of egg deposition and subsequently diminish as chronicity proceeds.

Increased numbers of CD4⁺FoxP3⁺T_{reg} cells occur in the mLN and colonic granulomas as chronicity proceeds

The proportion of CD4⁺ FoxP3⁺T_{regs} in the mLN as a proportion of total CD4⁺ cells, as determined by flow cytometry of cell suspensions, increased from 13.1±0.2% in naïve mice to 16.0±0.4% during acute infection (P<0.001), and increased further to 20.6±0.2% during chronic infection (P<0.001; Fig. 3A). Absolute numbers of both mLN CD4⁺ effector and CD4⁺FoxP3⁺T_{reg} cells increased during acute disease from naïve levels, and remained significantly elevated during chronic infection (Fig. 3A). The increase was confirmed by enumeration of FoxP3⁺T_{regs} in stained sections of mLN from naïve mice and those with acute and chronic infection (Fig. 3B & C). In contrast, absolute numbers or proportions of CD4⁺FoxP3⁺T_{regs} in the spleens did not expand (12.8±0.6% cf. 13.9±0.8% cf. 14.8±1.3%, Fig. 3A).

A pronounced increase in the proportion of CD4⁺FoxP3⁺T_{regs} within colonic granulomas at the chronic phase of infection, from 2.9±0.6% to 18.8±0.7% was revealed by enumeration of double positive versus single positive cells in anti-CD4 / anti-FoxP3 immunostained cryosections of colonic tissue (Fig. 3D & E). This profound (>10 fold) proportional elevation in FoxP3⁺ cells compared with total number of T lymphocytes in gut tissue was corroborated by qRT-PCR of FoxP3 transcript normalised to CD3 transcript (Fig. 3F). Thus, during enteric *S. mansoni* infection, relative and absolute expansion in the numbers of CD4⁺FoxP3⁺ cells occurs preferentially within gut-associated lymphoid tissue (GALT). Furthermore, relative increases of CD4⁺FoxP3⁺ cells within colonic granulomas are apparent during chronic disease.

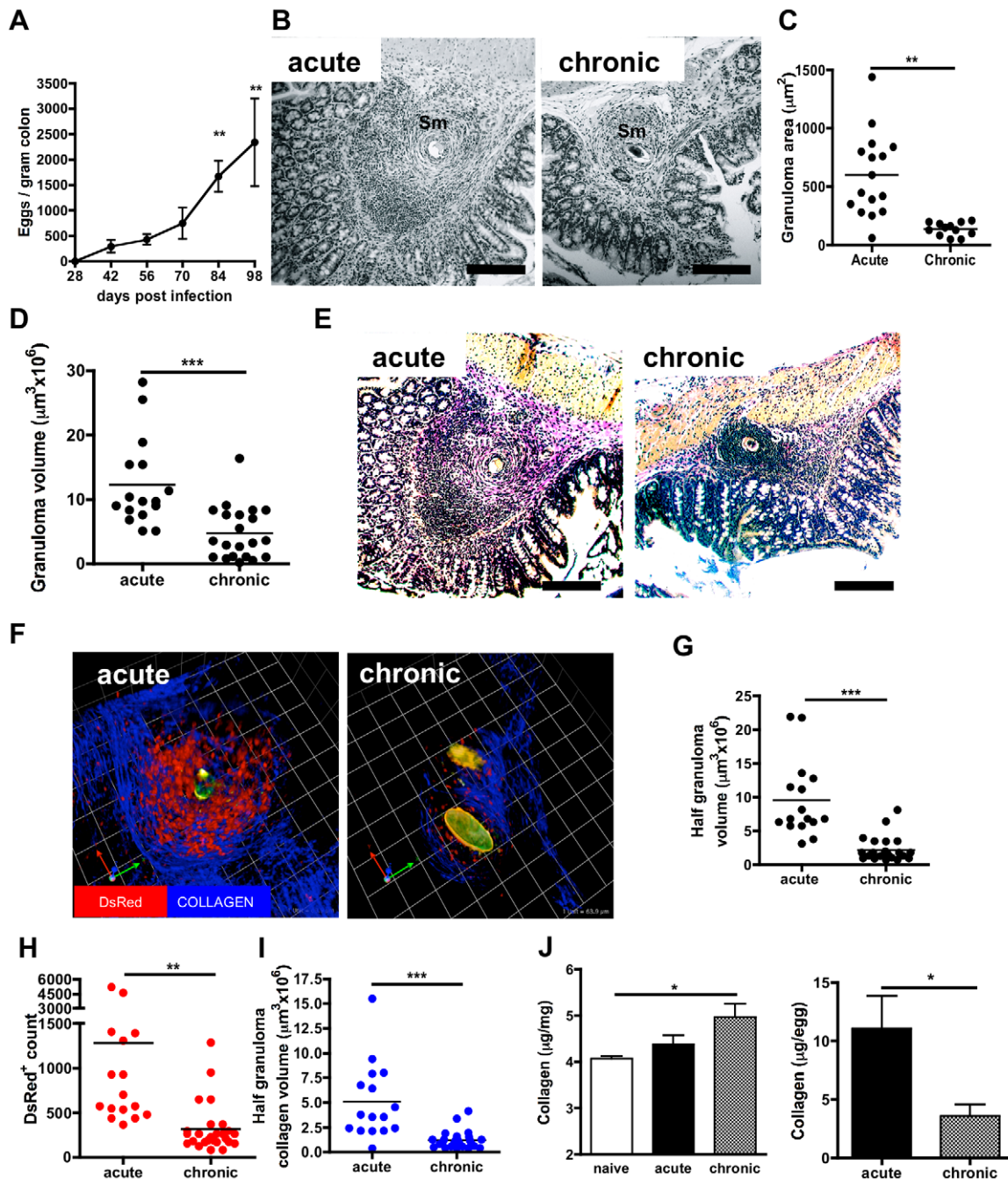


Figure 1. Colonic granuloma size and fibrosis is reduced in the chronic phase of schistosome infection. **A)** Accumulation of eggs /gram of colon tissue (n = 4 mice/time point); mean eggs (\pm SEM). **B)** Representative photomicrographs of the colon at the acute (8 wks) or chronic (14 wks) stage of infection stained with H&E. Scale bars are 200 μ m; egg denoted 'Sm'. **C)** Granuloma area at the acute or chronic stage; bars are mean / animal (n = 4) from three separate histological sections. **D)** Granuloma volumes calculated from isolated granulomas at the acute or chronic stage calculated from three separate granulomas per individual animal. **E)** Cross sections of colon at the acute (8 wks) or chronic (14 wks) stage of infection stained with haematoxylin / Van Geison (collagen fibres = pink). All scale bars are 200 μ m. Parasite egg is denoted 'Sm'. **F)** 3D images of multiphoton confocal stacks of colonic tissue from infected hCD2-VaDsRed-B.6 mice sampled *in situ* during acute or chronic infection. DsRed fluorescent cells are >90% CD3⁺; blue fluorescence is second harmonic generation of type 1 collagen; green/yellow auto-fluorescence are schistosome eggs. Grid squares are 63.9 μ m². Quantification of multiphoton confocal stacks from infected hCD2-VaDsRed-B.6 mice: **G)** granuloma half-volumes, **H)** DsRed⁺ cell counts and **I)** collagen half-volumes. Data calculated from four separate granulomas from individual animals (n = 4). **J)** Salt-soluble collagen from colonic tissues of naive, acute, and chronic mice (n = 4). Data is mean (\pm SEM) collagen conc^a/mg of tissue (left), or adjusted for numbers of eggs/mg (right). doi:10.1371/journal.pntd.0001269.g001

CD103, the α E molecule of the α E β 7 mucosal integrin involved in homing of T cells to intestinal sites [20], increased on mLN CD4⁺FoxP3⁺T_{regs} at the chronic stage compared to naive mice

(31.0 \pm 2.3% *of*: 62.4 \pm 3.1%, P<0.001, Fig. 3G). CCR5 is also involved in T cell homing to intestinal inflammatory sites [21], and significantly, the proportion of CCR5⁺CD4⁺FoxP3⁺T_{regs} in the

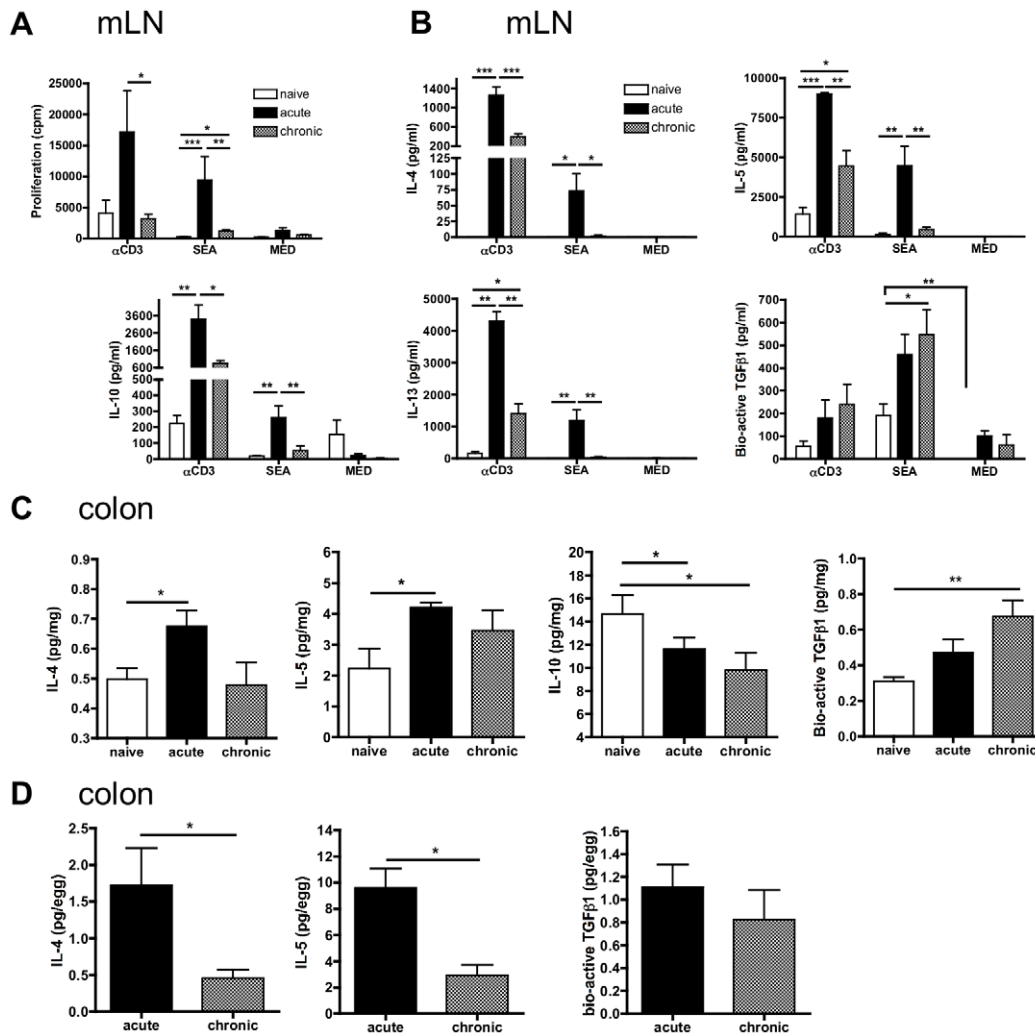


Figure 2. Egg antigen-specific Th2, but not TGF- β 1 responses become down-modulated within the mLN and colon during chronic infection. **A**) Proliferative responses and **B**) cytokine release (pg/ml) by mLN cells from naïve, acute, or chronic mice ($n = 4$ /group) to anti-CD3 mAb, or SEA, or MED. **C**) Cytokine levels within colon tissues (pg/mg tissue) and **D**) adjusted for numbers of eggs / mg of tissue. Data are mean proliferative response / cytokine concentration \pm SEM. doi:10.1371/journal.pntd.0001269.g002

mLN increased markedly ($9.6 \pm 2.3\%$ of $19.3 \pm 2.6\%$; Fig. 3G). Thus, increases in the number of mLN $CD4^+FoxP3^+T_{regs}$ expressing CD103 and CCR5 suggests these cells have enhanced potential to be recruited and retained within the colonic infection site during chronic infection.

The majority of Schistosome-expanded $CD4^+CD25^+$ mLN cells are $FoxP3^+$ and suppress antigen-specific $CD4^+$ proliferative responses *in vitro*

More than 75% of $CD4^+CD25^+$ mLN cells co-expressed $FoxP3$, regardless of the stage of infection confirming that the majority of $CD4^+CD25^+$ cells can be classified as $CD4^+FoxP3^+T_{regs}$ (Fig. 4A). In addition, while $CD4^+CD25^-$ effector cells from mice with an acute infection exhibited a strong proliferative response *in vitro* to SEA, $CD4^+CD25^+T_{regs}$ taken at the chronic stage of infection did not proliferate (Fig. 4B). Moreover, co-culture of these two cell populations in a 2:1 ratio prevented optimum antigen-specific proliferation of $CD4^+CD25^-$ effector T cells (Fig. 4B). Thus, $CD4^+CD25^+$ ($FoxP3^+$) cells within the mLN during chronic schistosome infection displayed a regulatory phenotype *in vitro*.

$CD4^+CD25^+$ T_{regs} isolated from the mLN of naïve mice, compared to mice with an acute or chronic infection, exerted similar degrees of suppression on the acute-stage anti-SEA $CD4^+$ T cell proliferative response (Fig. 4C). Moreover, following depletion of $CD4^+CD25^+$ T_{regs} , or after their re-addition in a 1:2 T_{reg} / effector T cell ratio, we observed that while the T_{regs} conferred a significant degree of suppression on the anti-SEA response, chronic $CD4^+$ $CD25^-$ effector T cells remained hypo-responsive, or anergic, compared with their acute-stage counterparts (Fig. 4D). Taken together, these *in vitro* assays provide evidence that schistosome-expanded $CD4^+CD25^+T_{regs}$ suppress the pre-dominant Th2 anti-SEA response. However, compared with T_{regs} from naïve mice, they are not enhanced in their ability to suppress antigen-specific $CD4^+$ proliferation.

Anti-CD25mAb treatment impairs the regulation of colonic granuloma size

Depletion of $CD25^+$ cells is a common technique to experimentally induce Treg deficiency [14,22,23,24,25,26]. Whilst not all $CD25^+$ cells are $FoxP3^+$ Tregs (75%–80% in our infection

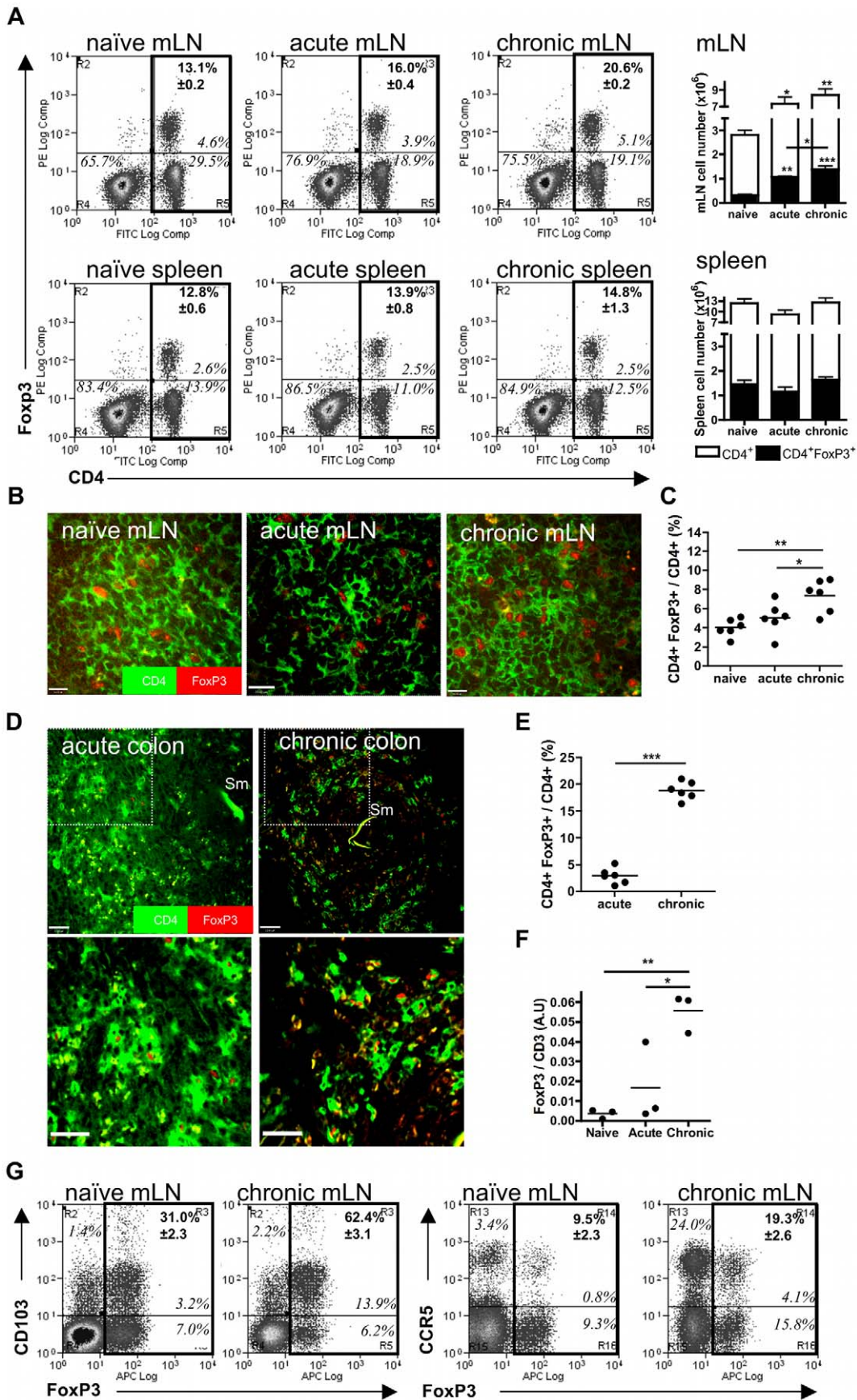


Figure 3. Frequencies of CD4⁺FoxP3⁺T_{regs} increase at the chronic phase in both the mLN and colonic granulomas. **A)** Representative flow cytograms showing the frequencies of labelled CD4⁺ and FoxP3⁺ cells in suspensions of the mLN and spleen. Values in italics are quadrant percentages. Values in bold, upper right-hand quadrant, are mean (\pm SEM) CD4⁺FoxP3⁺ cells as a % of total CD4⁺ cells (relevant quadrants outlined in bold). Histograms show mean (\pm SEM) total CD4⁺ (open) and CD4⁺FoxP3⁺ (closed), mLN or spleen cell or numbers (n=3 mice). Significant differences compared with naïve cell numbers are indicated. Bar indicates significant difference between CD4⁺FoxP3⁺ mLN numbers at the chronic compared with acute stage. **B)** Representative images of CD4⁺ (green) and FoxP3⁺ (red) cells in labelled cryosections of mLN and, **C)** proportion of CD4⁺FoxP3⁺ / CD4⁺ cells / field of view (2 fields of view / animal, n=3) Scale = 14 μ m. **D)** Representative images of CD4⁺ and FoxP3⁺ cells in colonic granulomas, with high power insert also shown. **E)** Frequencies of labelled CD4⁺FoxP3⁺ / CD4⁺ colonic granuloma cells / field of view (2 fields of view / animal, n=3). **F)** qRT-PCR analysis of FoxP3 mRNA in colonic tissue plotted as arbitrary units of FoxP3 normalised to CD3 transcript in colonic tissue. Bars are mean FoxP3 A.U. transcript per group. **G.)** Flow plots of FoxP3 and CD103, and FoxP3 and CCR5 expression, gated on CD4⁺ mLN cells of naïve or chronic mice (n=4). Values in italics are quadrant percentages. Values in bold, upper right-hand quadrant, are mean (\pm SEM) CD4⁺FoxP3⁺CD103⁺ cells, or CD4⁺FoxP3⁺CCR5⁺ cells as a % of total CD4⁺FoxP3⁺ cells (relevant quadrants outlined in bold). doi:10.1371/journal.pntd.0001269.g003

model) and FoxP3⁺Treg populations can be rapidly induced following CD25⁺ depletion during infection [27], a single antibody treatment with anti-CD25 clone PC61 can significantly reduce FoxP3⁺ cells by 70% [28], with reduced FoxP3⁺ cells persisting in the face of ensuing inflammation for two weeks [27]. We therefore treated 9 week schistosome infected mice with a regimen of PC61 once per two weeks for six weeks and assessed CD25⁺ lymphocyte depletion and effects on intestinal granuloma parameters one week (+14 weeks infected) following the last PC61 antibody treatment.

Anti-CD25 mAb treatment of infected mice effectively depleted CD25⁺ cells in the mLN, and reduced the proportion of CTLA-4⁺ lymphocytes (Fig. 5A). Anti-CD25-treated mice also retained significantly larger granulomas in chronic colonic tissues compared to their isotype control cohorts (P<0.01; Fig. 5B). Eosinophils adjacent to schistosome eggs were significantly more numerous in anti-CD25 treated mice, while the numbers of large mononuclear cells remained stable (Fig. 5C & D). This suggests that reductions in CD25⁺ cells, the majority of which are

FoxP3⁺T_{regs}, lifted suppression of eosinophil recruitment, or their retention within colonic granulomas. It was also co-incident with increased antigen-specific mLN cell proliferation and IL-4 production at the chronic phase of infection (Fig. 5E). However, in CD25⁺-depleted mice, colonic granulomas remained, on average, significantly smaller in area compared to those at the acute stage of infection (43,397 \pm 5477 μ m² compared with 107,128 \pm 12062 μ m², P<0.01). Thus, in vivo depletion of CD25⁺ lymphocytes partially, yet significantly, reverses the down-modulation of Th2 granulomatous pathology in the colon during chronic *S. mansoni* infection.

Transfer of infection-expanded CD4⁺CD25⁺T_{regs} modulates acute colonic granuloma development

Purified CD4⁺CD25⁺ mLN cells (2.5 \times 10⁶, >90% purity; Fig. 6A) from mice at the chronic stage of infection were administered to hCD2-VaDsRed-B.6 mice co-incident with the onset of egg deposition. Four weeks later (+9 weeks post-infection),

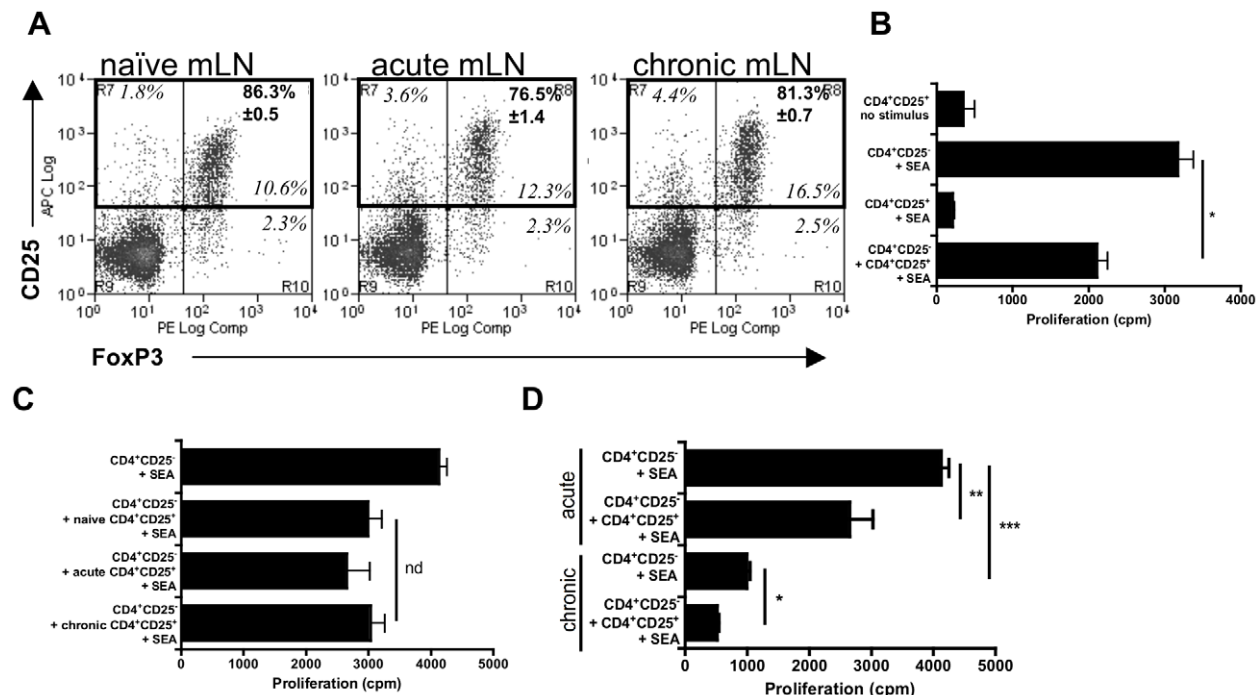


Figure 4. CD4⁺CD25⁺ mLN cells suppress antigen-specific CD4⁺ Th2 responses in vitro. **A)** Flow plots of mLN cell suspensions (n=3 mice) labelled with anti-CD25 and anti-FoxP3, gated on CD4 expression. Values in italics are quadrant percentages. Values in bold, upper right-hand quadrant, are mean (\pm SEM) CD4⁺CD25⁺FoxP3⁺ cells as a % of total CD4⁺CD25⁺ cells (relevant quadrants outlined in bold). **B)** Antigen-specific proliferation of CD4⁺CD25⁺ effector cells from acute mice and CD4⁺CD25⁺T_{regs} from chronic mice, or co-cultured together in a 2:1 ratio. **C)** Co-culture of CD4⁺CD25⁺ effector cells derived from acute mice cultured with CD4⁺CD25⁺T_{regs} derived from naïve, acute or chronic mice. **D)** CD4⁺CD25⁺ effector cells from mice with an acute or chronic infection, either depleted of CD4⁺CD25⁺ cells, or with CD4⁺CD25⁺ cell 'add-back' co-cultures from the same stage of infection at a 2:1 ratio. All histograms are mean (\pm SEM) cpm ³H-thymidine incorporation. doi:10.1371/journal.pntd.0001269.g004

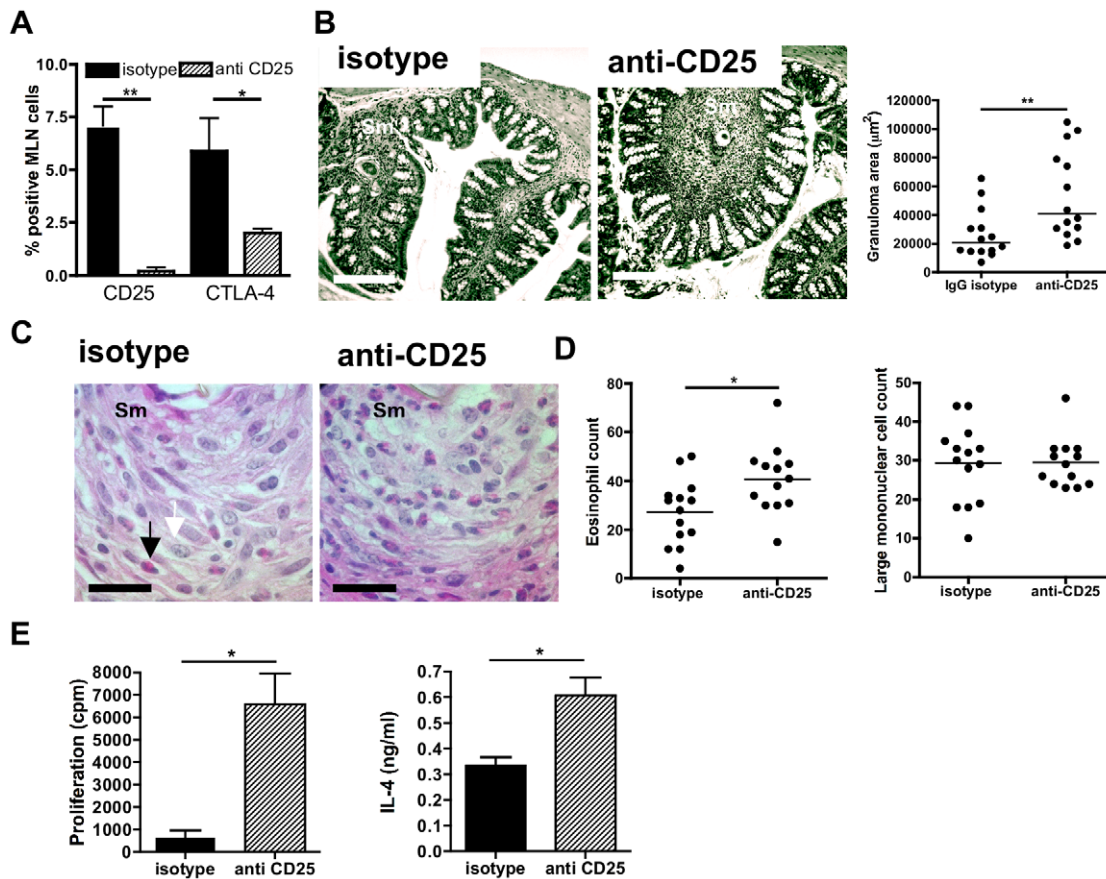


Figure 5. *In vivo* ablation of CD25⁺ cells impairs regulation of colonic granulomas and antigen-specific Th2 responses. **A**) Percentage of CD25⁺ or CTLA-4⁺ mLN lymphocytes from mice with chronic infection after treatment with anti-CD25 mAb, or isotype control. Antibodies given at 2 week intervals from wk 9 to wk 13, tissues sampled at wk 14. Data are mean % positive (\pm SEM). **B**) Photomicrographs of colonic tissue and quantification of granuloma areas after antibody treatment. Scale bar = 200 μ m; egg denoted 'Sm'. Data is granuloma area (μ m²) at wk 14 calculated from 3-4 separate histological sections per individual animal (n=4). **C**) Colonic granulomas (x100) stained with H&E showing eosinophils (closed arrow) and large mononuclear cells (open arrow) in isotype mAb treated (left) and anti-CD25 treated (right) infected mice. Scale bar = 200 μ m; egg denoted 'Sm'. **D**) Numbers of eosinophils (left) and large mononuclear cells enumerated from the sections above; Bars are mean cell counts / group (n=4) with 3-4 fields of view / mouse. **E**) Antigen-specific mLN responses in anti-CD25 mAb treated mice; data are means of cpm ³H-thymidine incorporation and IL-4 secretion pg/ml (n=3). doi:10.1371/journal.pntd.0001269.g005

the number of eggs in tissues or excreted from CD25⁺ cell recipients were not significantly different compared with controls (Table S1). This showed that immune cell transfers did not significantly affect adult worm development, fecundity, or egg transmission. Recipients of CD4⁺CD25⁺ cells had significantly smaller granuloma area and collagen deposits but Ds-Red⁺ T cell numbers within the colonic granulomas were not significantly altered (Fig. 6B & C; Videos S3 & S4). Recipients also had decreased levels of recently synthesised collagen and IL-4 in colonic extracts (Fig. 6D) although simultaneous down-regulation of Th2 mLN responses were not observable in recipient mice (data not shown). Lower numbers of transferred cells (1×10^6) did not significantly alter acute-stage enteric granuloma formation (data not shown). Thus, CD4⁺CD25⁺T_{regs} cells, which expand within the mLN of chronically-infected mice, exert a suppressive effect on the development of acute-phase Th2 inflammation.

Discussion

Our data provides both *in vitro* and *in vivo* evidence that intestinal-associated CD4⁺CD25⁺FoxP3⁺T_{regs} expand during chronic, schistosome-induced colitic inflammation. They mediate

significant levels of antigen-specific Th2 suppression *in vivo* including reduced IL-4 production, eosinophil recruitment, collagen production, and an overall reduction in the size of egg-induced granulomas in the large intestine. Thus, our data demonstrates experimentally, that CD4⁺CD25⁺T_{regs} are capable of modulating Th2 inflammation and fibrosis associated with intestinal disorders.

Whilst an expansion of gut-associated CD4⁺FoxP3⁺T_{regs} was observed following schistosome infection, this could be a product of proliferating, naturally occurring (n)T_{regs} in response to auto-antigens (*e.g.* arising from disrupted intestinal barrier), or comprise an induced population of FoxP3⁺T_{regs} recognising schistosome antigens. However, in a syngenic adoptive transfer model, numbers of nT_{regs} did not expand relative to CD4⁺ effector cells within the mLN [24], suggesting that nT_{reg} expansion does not fully account for the heightened ratio of FoxP3⁺CD4⁺: CD4⁺ cells during chronic infection. The development of schistosome infection-induced T_{regs} is likely to be favoured by the constitutive production of intestinal signals such as TGF β and retinoic acid [29,30]. Intriguingly, SEA induces TGF β 1 secretion [31] and is critical for the development of auto-immune suppressing FoxP3⁺T_{regs} following *in vivo* SEA injection [32]. Since we observed

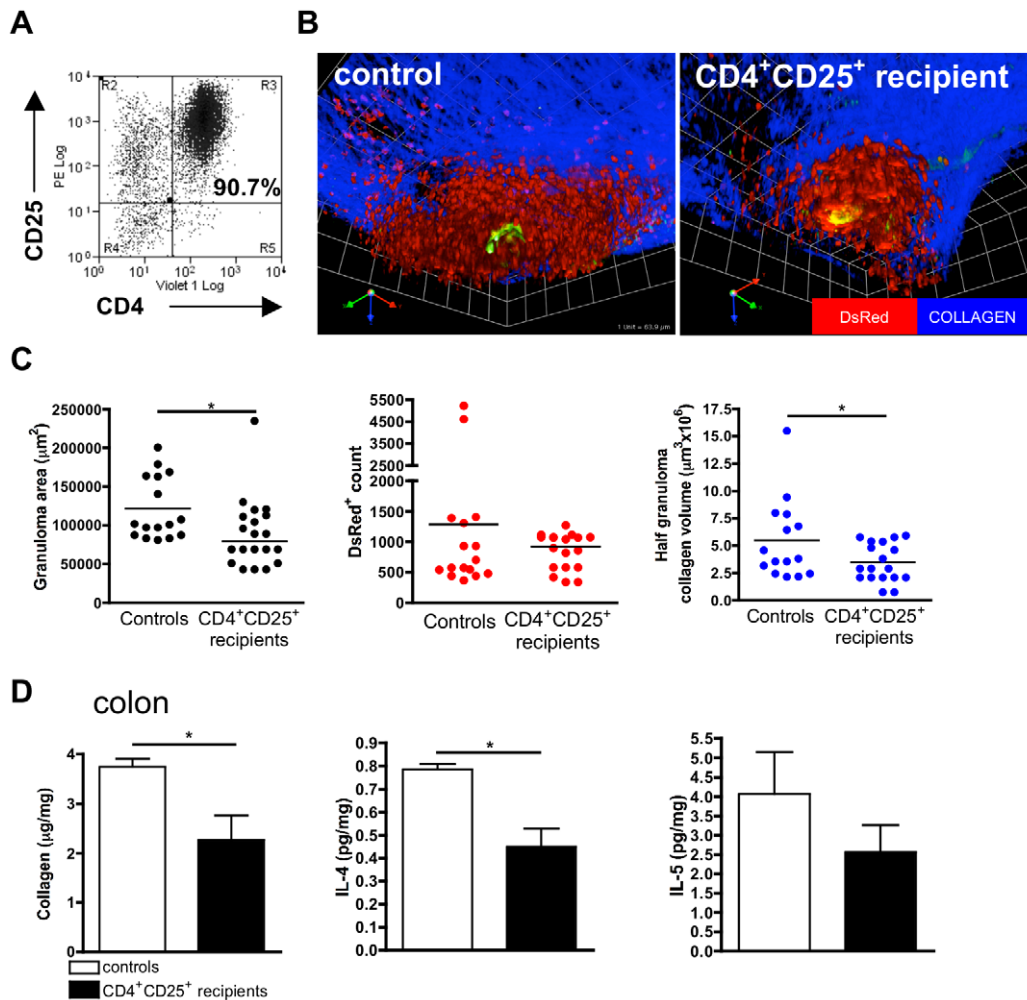


Figure 6. Transfer of schistosome-expanded $CD4^+CD25^+$ T_{regs} modulates the development of acute-stage granulomas. **A**) Isolated mLN $CD4^+CD25^+$ T_{regs} from mice with chronic infection used for transfer. **B**) 3D images of multiphoton confocal stacks of colonic tissue viewed *in situ* at the acute stage of infection in hCD2-VaDsRed-B.6 control mice, or recipients of infection-expanded $CD4^+CD25^+$ T_{regs} (2.5×10^6). Grid squares are $63.9 \mu m^2$. **C**) Granuloma area (left), DsRed⁺ cell counts (middle), and collagen half-volume (right) taken from confocal stacks (as above). Data are from 5–6 separate granulomas per mouse. Bars are mean / group ($n = 3$). **D**) Soluble collagen and cytokine in colonic extracts from infected control mice, or recipients of $CD4^+CD25^+$ cells. Bars are means / group ($n = 3$). Data is mean pg/ml (\pm SEM) per group. doi:10.1371/journal.pntd.0001269.g006

heightened bioactive TGF β 1 from mLN cells and within the colon at the chronic phase of infection, we speculate that TGF β 1 might have a role in the induction of gut-homing $CD4^+FoxP3^+T_{regs}$ from naïve precursors. Both proportions and absolute numbers of $CD4^+FoxP3^+T_{regs}$ significantly increased within the mLN during acute infection, when anti-egg Th2 responses are at their peak. Although inhibition of Treg induction by Th2 differentiation programs has been reported [33], our data would suggest that Th2 differentiation is insufficient to block outgrowth of a regulatory T cell phenotype during acute intestinal schistosomiasis.

$CD4^+FoxP3^+T_{regs}$ within the mLN of mice with a chronic infection expressed elevated levels of CD103 and CCR5, both of which are associated with homing to mucosal tissues during inflammation [20,21]. However, infection status did not alter the *in vitro* suppressive ability on a per-cell basis of $CD4^+CD25^+T_{regs}$ from the mLN, indicating that infection-expanded $CD4^+FoxP3^+T_{regs}$ and those from naïve mice share a common mechanism of Th2-effector cell suppression. Therefore, it is likely that the increased regulatory activity of $CD4^+FoxP3^+T_{regs}$ *in vivo* during chronic infection is a product of either increased numbers

trafficking to the site of egg deposition, or greater survival/retention in colonic granulomas. As recipients of schistosome-expanded $CD4^+CD25^+$ cells displayed selective suppression of Th2 activity at the enteric infection site versus the mLN, this may reflect a biased homing of T_{regs} in the colon that provides increased regulatory pressure on the local granulomatous response (*i.e.* collagen synthesis and eosinophil recruitment). Taken with the observation that ablation of $CD4^+CD25^+$ cells *in vivo* significantly restores the anti-egg IL-4 and proliferative mLN cell response, these data suggest that intestinal-associated, schistosome infection-expanded $FoxP3^+$ T_{regs} exert layers of Th2 suppression both within gut-draining lymph nodes and within the colonic infection site.

Our data are consistent with recent findings that $CD4^+FoxP3^+T_{regs}$ constitute a partial component of the modulation of granuloma development in the liver [14,24,34]. We show that depletion of $CD4^+CD25^+$ cells *in vivo* does not fully reverse the Th2 hypo-responsiveness, nor do chronic enteric granulomas in treated mice fully recover the florid cellularity of the acute phase. The use of transgenic mice deficient for *foxp3* [35] could help

further investigation of this phenomenon. The development of other regulatory cells such as alternatively-activated macrophages [36], IL-10-secreting non-FoxP3 cells [14], and regulatory B cells [37], may explain the partial role of CD4⁺FoxP3⁺T_{regs} in our system. T cell intrinsic energy, for instance mediated by GRAIL signalling [38], could also account for the remaining hypo-responsiveness after depletion of CD4⁺ CD25⁺ cells. However, because the micro-environment of the intestines is favourable towards the expansion of CD4⁺FoxP3⁺T_{regs}, it is possible that CD4⁺FoxP3⁺T_{reg}-mediated suppression of enteric granulomas is more apparent compared with their hepatic counterparts. This could explain why colonic granulomas modulate more rapidly versus hepatic granulomas after the acute stage, and why hepatic granulomas retain a greater size and cellularity than enteric granulomas during chronic disease [19].

During experimental helminth infections, CD4⁺CD25⁺T_{reg}-mediated suppression of Th2 effector responses confer a permissive state by stifling effective Th2-mediated worm attrition [23,25,26]. In the case of gut-helminths, this potentially operates by regulating Th2 cytokine signalling on smooth muscle contractility and epithelial cell turnover [39,40]. Paradoxically, during *S. mansoni* infections, while Th2 granulomatous responses protect and aid survival of the host [11,41], intact Th2 responses are also essential to propagate the parasite's life cycle, as egg transmission from the gut is impaired in the absence of T cells [42,43,44], IL-4 [41], or IL-4R α signalling [45]. Nevertheless, transfer of CD4⁺CD25⁺ cells to acute stage recipients does not impinge on egg transmission in spite of a modulatory effect on IL-4 in the colon. In fact, during the chronic stage of infection, where Th2 inflammation in the colon becomes modulated, egg excretion rates are unaffected (our unpublished observations). How intestinal schistosome parasites mediate sustained, chronic egg transmission in the face of marked Th2 hypo-responsiveness remains to be identified.

Enteric helminth infections, or products derived from helminths, are gaining prominence as potential therapies to reverse the effects of inflammatory bowel disease [46]. Some ameliorating capacities may be attributed to antagonism of Th1 processes by induction of IL-4-secreting cells within intestinal tissues rather than by induction of T_{regs}. Indeed, most experimental studies demonstrating the modulatory action of helminth infection used trinitrobenzene sulphate as a haptens agent to drive Th1 colitis reminiscent of Crohn's Disease [46]. Initiation of Th2/NKT colitis by oxazolone in conjunction with Th2-promoting helminth infection exacerbated pathology [47], further supporting that the mechanism of helminth suppression of colitis is based on Th2/Th1

antagonism. Thus, from a clinical perspective, helminth-based therapies might be considered inappropriate for ulcerative colitis or other intestinal disorders with Th2 aetiology. However, our data demonstrates that helminth infection-expanded FoxP3⁺ T_{regs} clearly regulate coincident pro-fibrotic Th2 processes in the colon. Indeed, distinct molecules released by schistosome eggs deliver triggers that polarize naive CD4⁺ T cells towards Th2, or a regulatory phenotype [8,9,10,48]. Potentially, the release of somatic molecules with regulatory potential from degrading eggs that fail to breach colonic tissues could favour T_{reg} expansion during chronic disease in the context of a TGF β -enriched microenvironment. Exploitation of such regulatory molecules may be of benefit in the treatment of intestinal schistosomiasis or other Th2-based intestinal disorders via the expansion of T_{regs} with bystander potential.

Supporting Information

Video S1 Acute colonic granuloma shown in Fig 1F.

(MOV)

Video S2 Chronic colonic granuloma shown in Fig 1F.

(MOV)

Video S3 Control recipient, colonic granuloma shown in Fig. 6B.

(MOV)

Video S4 CD4⁺CD25⁺ recipient, colonic granuloma shown in Fig. 6B.

(MOV)

Table S1 Acute-stage schistosome egg tissue burdens and excretion following adoptive transfer of chronic infection-expanded CD4⁺CD25⁺ cells.

(DOC)

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Author Contributions

Conceived and designed the experiments: APM GRJ KGH JDT. Performed the experiments: APM GRJ KGH PCC SA RAP JDT. Analyzed the data: APM JDT KGH GRJ. Contributed reagents/materials/analysis tools: MC. Wrote the paper: JDT APM.

References

- Chitsulo L, Loverde P, Engels D (2004) Schistosomiasis. *Nat Rev Microbiol* 2: 12–13.
- Steinmann P, Keiser J, Bos R, Tanner M, Utzinger J (2006) Schistosomiasis and water resources development: systematic review, meta-analysis, and estimates of people at risk. *Lancet Infect Dis* 6: 411–425.
- Gryseels B, Polman K, Clerinx J, Kestens L (2006) Human schistosomiasis. *Lancet* 368: 1106–1118.
- Wilson MS, Mentink-Kane MM, Pesce JT, Ramalingam TR, Thompson R, et al. (2007) Immunopathology of schistosomiasis. *Immunol Cell Biol* 85: 148–154.
- King CL (2001) Initiation and regulation of disease in schistosomiasis. In: Mahmoud A, ed. *Schistosomiasis*. London: Imperial College Press. pp 213–264.
- King CH, Dickman K, Tisch DJ (2005) Reassessment of the cost of chronic helminthic infection: a meta-analysis of disability-related outcomes in endemic schistosomiasis. *Lancet* 365: 1561–1569.
- Rocklin RE, Brown AP, Warren KS, Pelley RP, Houba V, et al. (1980) Factors that modify the cellular-immune response in patients infected by *Schistosoma mansoni*. *J Immunol* 125: 1916–1923.
- Steinfeldt S, Andersen JF, Cannons JL, Feng CG, Joshi M, et al. (2009) The major component in schistosome eggs responsible for conditioning dendritic cells for Th2 polarization is a T2 ribonuclease (omega-1). *J Exp Med* 206: 1681–1690.
- Everts B, Perona-Wright G, Smits HH, Hokke CH, van der Ham AJ, et al. (2009) Omega-1, a glycoprotein secreted by *Schistosoma mansoni* eggs, drives Th2 responses. *J Exp Med* 206: 1673–1680.
- Schramm G, Mohrs K, Wodrich M, Doenhoff MJ, Pearce EJ, et al. (2007) Cutting edge: IPSE/alpha-1, a glycoprotein from *Schistosoma mansoni* eggs, induces IgE-dependent, antigen-independent IL-4 production by murine basophils in vivo. *J Immunol* 178: 6023–6027.
- Fallon PG, Richardson EJ, McKenzie GJ, McKenzie AN (2000) Schistosome infection of transgenic mice defines distinct and contrasting pathogenic roles for IL-4 and IL-13: IL-13 is a profibrotic agent. *J Immunol* 164: 2585–2591.
- Fallon PG, Dunne DW (1999) Tolerization of mice to *Schistosoma mansoni* egg antigens causes elevated type 1 and diminished type 2 cytokine responses and increased mortality in acute infection. *J Immunol* 162: 4122–4132.
- Hoffmann KF, Cheever AW, Wynn TA (2000) IL-10 and the dangers of immune polarization: excessive type 1 and type 2 cytokine responses induce distinct forms of lethal immunopathology in murine schistosomiasis. *J Immunol* 164: 6406–6416.

14. Hesse M, Piccirillo CA, Belkaid Y, Prufer J, Mentink-Kane M, et al. (2004) The pathogenesis of schistosomiasis is controlled by cooperating IL-10-producing innate effector and regulatory T cells. *J Immunol* 172: 3157–3166.
15. Taylor JJ, Mohrs M, Pearce EJ (2006) Regulatory T cell responses develop in parallel to Th responses and control the magnitude and phenotype of the Th effector population. *J Immunol* 176: 5839–5847.
16. Weinstock JV, Boros DL (1983) Organ-dependent differences in composition and function observed in hepatic and intestinal granulomas isolated from mice with Schistosomiasis mansoni. *J Immunol* 130: 418–422.
17. Hogg KG, Kumkate S, Anderson S, Mountford AP (2003) Interleukin-12 p40 secretion by cutaneous CD11c+ and F4/80+ cells is a major feature of the innate immune response in mice that develop Th1-mediated protective immunity to Schistosoma mansoni. *Infect Immun* 71: 3563–3571.
18. Abe M, Harpel JG, Metz CN, Nunes I, Loskutoff DJ, et al. (1994) An assay for transforming growth factor-beta using cells transfected with a plasminogen activator inhibitor-1 promoter-luciferase construct. *Anal Biochem* 216: 276–284.
19. Weinstock JV, Boros DL (1981) Heterogeneity of the granulomatous response in the liver, colon, ileum, and ileal Peyer's patches to schistosome eggs in murine schistosomiasis mansoni. *J Immunol* 127: 1906–1909.
20. Izcue A, Coombes JL, Powrie F (2006) Regulatory T cells suppress systemic and mucosal immune activation to control intestinal inflammation. *Immunol Rev* 212: 256–271.
21. Luangsay S, Kasper LH, Rachinel N, Minns LA, Mennechet EJ, et al. (2003) CCR5 mediates specific migration of Toxoplasma gondii-primed CD8 lymphocytes to inflammatory intestinal epithelial cells. *Gastroenterology* 125: 491–500.
22. Wilson MS, Taylor MD, Balic A, Finney CA, Lamb JR, et al. (2005) Suppression of allergic airway inflammation by helminth-induced regulatory T cells. *J Exp Med* 202: 1199–1212.
23. Taylor MD, LeGoff L, Harris A, Malone E, Allen JE, et al. (2005) Removal of regulatory T cell activity reverses hyporesponsiveness and leads to filarial parasite clearance in vivo. *J Immunol* 174: 4924–4933.
24. Baumgart M, Tompkins F, Leng J, Hesse M (2006) Naturally occurring CD4+Foxp3+ regulatory T cells are an essential, IL-10-independent part of the immunoregulatory network in Schistosoma mansoni egg-induced inflammation. *J Immunol* 176: 5374–5387.
25. Taylor MD, van der Werf N, Harris A, Graham AL, Bain O, et al. (2009) Early recruitment of natural CD4+ Foxp3+ Treg cells by infective larvae determines the outcome of filarial infection. *Eur J Immunol* 39: 192–206.
26. D'Elia R, Behnke JM, Bradley JE, Else KJ (2009) Regulatory T cells: a role in the control of helminth-driven intestinal pathology and worm survival. *J Immunol* 182: 2340–2348.
27. Couper KN, Blount DG, de Souza JB, Suffia I, Belkaid Y, et al. (2007) Incomplete depletion and rapid regeneration of Foxp3+ regulatory T cells following anti-CD25 treatment in malaria-infected mice. *J Immunol* 178: 4136–4146.
28. Setiady YY, Coccia JA, Park PU (2010) In vivo depletion of CD4+FOXP3+ Treg cells by the PC61 anti-CD25 monoclonal antibody is mediated by FcγRIIIb phagocytes. *Eur J Immunol* 40: 780–786.
29. Chen W, Jin W, Hardegen N, Lei KJ, Li L, et al. (2003) Conversion of peripheral CD4+CD25- naive T cells to CD4+CD25+ regulatory T cells by TGF-beta induction of transcription factor Foxp3. *J Exp Med* 198: 1875–1886.
30. Sun CM, Hall JA, Blank RB, Bouladoux N, Oukka M, et al. (2007) Small intestine lamina propria dendritic cells promote de novo generation of Foxp3 T reg cells via retinoic acid. *J Exp Med* 204: 1775–1785.
31. Zaccane P, Burton OT, Gibbs S, Miller N, Jones FM, et al. (2010) Immune modulation by Schistosoma mansoni antigens in NOD mice: effects on both innate and adaptive immune systems. *J Biomed Biotechnol* 2010:795210. pp 795210.
32. Zaccane P, Burton O, Miller N, Jones FM, Dunne DW, et al. (2009) Schistosoma mansoni egg antigens induce Treg that participate in diabetes prevention in NOD mice. *Eur J Immunol* 39: 1098–1107.
33. Dardalhon V, Awasthi A, Kwon H, Galileos G, Gao W, et al. (2008) IL-4 inhibits TGF-beta-induced Foxp3+ T cells and, together with TGF-beta, generates IL-9+ IL-10+ Foxp3(-) effector T cells. *Nat Immunol* 9: 1347–1355.
34. Walsh CM, Smith P, Fallon PG (2007) Role for CTLA-4 but not CD25+ T cells during Schistosoma mansoni infection of mice. *Parasite Immunol* 29: 293–308.
35. Lahl K, Sparwasser T (2011) In vivo depletion of FoxP3+ Tregs using the DEREK mouse model. *Methods Mol Biol* 707: 157–172.
36. Pesce JT, Ramalingam TR, Wilson MS, Mentink-Kane MM, Thompson RW, et al. (2009) Retna (relmalph/fizz1) suppresses helminth-induced Th2-type immunity. *PLoS Pathog* 5: e1000393.
37. Amu S, Saunders SP, Kronenberg M, Mangan NE, Atzberger A, et al. (2010) Regulatory B cells prevent and reverse allergic airway inflammation via FoxP3-positive T regulatory cells in a murine model. *J Allergy Clin Immunol* Mar 19. epublished ahead of print.
38. Taylor JJ, Krawczyk CM, Mohrs M, Pearce EJ (2009) Th2 cell hyporesponsiveness during chronic murine schistosomiasis is cell intrinsic and linked to GRAIL expression. *J Clin Invest* 119: 1019–1028.
39. Cliffe LJ, Humphreys NE, Lane TE, Potten CS, Booth C, et al. (2005) Accelerated intestinal epithelial cell turnover: a new mechanism of parasite expulsion. *Science* 308: 1463–1465.
40. Zhao A, Urban JF Jr., Anthony RM, Sun R, Stiltz J, et al. (2008) Th2 cytokine-induced alterations in intestinal smooth muscle function depend on alternatively activated macrophages. *Gastroenterology* 135: 217–225. e211.
41. Fallon PG, Richardson EJ, Smith P, Dunne DW (2000) Elevated type 1, diminished type 2 cytokines and impaired antibody response are associated with hepatotoxicity and mortalities during Schistosoma mansoni infection of CD4-depleted mice. *Eur J Immunol* 30: 470–480.
42. Doenhoff M, Musallam R, Bain J, McGregor A (1978) Studies on the host-parasite relationship in Schistosoma mansoni-infected mice: the immunological dependence of parasite egg excretion. *Immunology* 35: 771–778.
43. Cheever AW, Eltoum IA, Andrade ZA, Cox TM (1993) Biology and pathology of Schistosoma mansoni and Schistosoma japonicum infections in several strains of nude mice. *Am J Trop Med Hyg* 48: 496–503.
44. Karanja DM, Colley DG, Nahlen BL, Ouma JH, Secor WE (1997) Studies on schistosomiasis in western Kenya. I. Evidence for immune-facilitated excretion of schistosome eggs from patients with Schistosoma mansoni and human immunodeficiency virus coinfections. *Am J Trop Med Hyg* 56: 515–521.
45. Herbert DR, Holscher C, Mohrs M, Arendse B, Schwegmann A, et al. (2004) Alternative macrophage activation is essential for survival during schistosomiasis and downmodulates T helper 1 responses and immunopathology. *Immunity* 20: 623–635.
46. Ruysers NE, De Winter BY, De Man JG, Loukas A, Herman AG, et al. (2008) Worms and the treatment of inflammatory bowel disease: are molecules the answer? *Clin Dev Immunol* 567314 2008: 567314.
47. Hunter MM, Wang A, McKay DM (2007) Helminth infection enhances disease in a murine TH2 model of colitis. *Gastroenterology* 132: 1320–1330.
48. van der Kleij D, Latz E, Brouwers JF, Kruize YC, Schmitz M, et al. (2002) A novel host-parasite lipid cross-talk. Schistosomal lyso-phosphatidylserine activates toll-like receptor 2 and affects immune polarization. *J Biol Chem* 277: 48122–48129.