

On the pathway to better birth outcomes? A systematic review of azithromycin and curable sexually transmitted infections

Expert Rev. Anti Infect. Ther. Early online, 1–30 (2013)

R Matthew Chico*¹,
Berkin B Hack²,
Melanie J Newport²,
Enesia Ngulube¹ and
Daniel Chandramohan¹

¹London School of Hygiene and Tropical Medicine Keppel Street, London, WC1E 7HT, UK

²Brighton and Sussex Medical School, Brighton, East Sussex, BN1 9PX, UK

*Author for correspondence:

Tel.: +44 20 7636 8636 ext. 2841

Fax: +44 207 927 2918

matthew.chico@lshtm.ac.uk

The WHO recommends the administration of sulfadoxine-pyrimethamine (SP) to all pregnant women living in areas of moderate (stable) to high malaria transmission during scheduled antenatal visits, beginning in the second trimester and continuing to delivery. Malaria parasites have lost sensitivity to SP in many endemic areas, prompting the investigation of alternatives that include azithromycin-based combination (ABC) therapies. Use of ABC therapies may also confer protection against curable sexually transmitted infections and reproductive tract infections (STIs/RTIs). The magnitude of protection at the population level would depend on the efficacy of the azithromycin-based regimen used and the underlying prevalence of curable STIs/RTIs among pregnant women who receive preventive treatment. This systematic review summarizes the efficacy data of azithromycin against curable STIs/RTIs.

KEYWORDS: azithromycin • bacterial vaginosis • Chlamydia • gonorrhoea • malaria • pregnancy • reproductive tract infections • sexually transmitted infections • sub-Saharan Africa • syphilis • trichomoniasis

The WHO recommends the administration of sulfadoxine-pyrimethamine (SP) to all pregnant women who live in areas of moderate (stable) to high malaria transmission during scheduled antenatal care (ANC) visits, beginning in the second trimester and continuing to delivery [1]. This intervention, known as intermittent preventive treatment of malaria in pregnancy (IPTp), is national policy in 36 countries worldwide, 35 of which are in sub-Saharan Africa [2]. The objective of IPTp-SP is to reduce the incidence of low birthweight and maternal anemia attributable to malaria. In recent years, however, malaria parasites have developed resistance to SP such that IPTp no longer reduces the incidence of low birthweight in some epidemiological settings, particularly in East Africa [3]. Evidence suggests that in areas where parasites express the 581G *dhps* mutation that is associated with SP resistance, the administration of IPTp-SP may even harm fetal growth [4–6]. Thus, the urgency to replace SP has never been greater and azithromycin-based combination

(ABC) therapies are among leading candidates to do so.

Azithromycin is a slow-acting analog of erythromycin in the macrolide (azalide) class of drugs, which targets the ribosomal subunit of susceptible microorganisms and causes cellular death by inhibiting protein synthesis [7]. It has *in vitro* and *in vivo* antimalarial properties [8] and can be safely administered during pregnancy [9]. Two human challenge studies have published results of azithromycin monotherapy treatment against *Plasmodium falciparum* infection. The first study reported a protective effect of 40% (n = 10; 95% CI: 12–74%) among immunologically naïve patients who received 250 mg azithromycin daily for 2 weeks prior to inoculation and for 1 week more following exposure [10]. When the same regimen was used for one additional week post-inoculation, treatment efficacy was 100% (n = 10) [11]. Despite this finding, comparable results have not been replicated in endemic settings where patients often have

mixed and multiple infections. However, *in vitro* evidence suggests that azithromycin may be combined with antimalarial partner drugs to prevent or to cure *P. falciparum* infection [12], the malaria species most prevalent in sub-Saharan Africa and which uniquely adhere to the placenta of pregnant women. In addition to reducing the burden of malaria infection, ABC therapies may also protect against adverse birth outcomes attributable to curable sexually transmitted and reproductive tract infections (STIs/RTIs). This could offer considerable public health impact. A recent meta-analysis suggests that curable STIs/RTIs are as prevalent as malaria parasitemia, if not more so, among pregnant women who attend ANC facilities in sub-Saharan Africa [13]. Five curable STIs/RTIs – *Treponema pallidum*, *Neisseria gonorrhoeae*, *Chlamydia trachomatis*, *Trichomonas vaginalis* and bacterial vaginosis – are associated with adverse birth outcomes that include spontaneous abortion [14–18], stillbirth [19–21], intrauterine growth retardation [20,22,23], premature rupture of membranes [24–26], preterm birth [17,22,23,26–33] and low birthweight (TABLE 1) [20,23,24,28,29,33–35]. This paper summarizes azithromycin efficacy and sensitivity against these curable STIs/RTIs and highlights important issues for policymakers to consider while determining the potential use of ABC therapies in IPTp.

Methodology

Between April and May 2013, PubMed, MEDLINE and EMBASE were searched using Medical Subject Headings and free-text terms for publications specific to the curable STIs/RTIs noted above. With each query, the infection and causal organism were used together, for example, ‘Syphilis’ AND ‘*Treponema pallidum*’, and then combined with search terms ‘azithromycin’ OR ‘macrolide’. Because the evidence base is limited with respect to azithromycin and some curable STIs/RTIs, both ‘azithromycin’ and ‘macrolide’ were used as filters. We had particular interest in randomized clinical trials (RCTs) that compared azithromycin against the current first-line treatments for curable STIs/RTIs in pregnancy, noting that azithromycin is the WHO-recommended treatment for pregnant women infected with *C. trachomatis*. Searches were limited to the English language and strict inclusion and exclusion criteria were applied so as to narrow the number of papers retained. Reference lists were also reviewed for additional documents. Excluded records and full-text articles were in seven categories:

- **‘Unrelated outcomes’** were studies that reported nonclinical aspects of azithromycin use such as cost-effective analysis, noncommunicable diseases such as heart disease or pharmacological outcomes involving a route of administration that is not applicable to this review (e.g., intravenous);
- **‘Unrelated organisms’** were papers dedicated to microbes that are not the focus of this review;
- **‘Not specific to STI/RTI’** were articles on the subject of same genus of interest, for example, *Chlamydia*, but were not specific to the genital tract, for example, *Chlamydia pneumoniae*;

- **‘Not related to azithromycin or close macrolide family’** were papers that did not contain macrolides in their analysis or outcomes, but focused on different antimicrobials against the organisms in question;
- **‘Sequential observations from same source’** involved surveillance reports from which most recent data set was used;
- **‘General discussion papers’** contained information pertinent to the search, but failed to provide specific data for STIs/RTIs.
- **‘Contraindicated in pregnancy’** were papers that reported outcomes of azithromycin combined with antimicrobial compounds that are considered unsafe in pregnancy.

A total of 122 articles met our primary inclusion criteria (FIGURE 1).

Results

Treponema pallidum

In vivo evidence

The WHO recommends treating pregnant women with syphilis infection using 2.4 million units of benzathine penicillin G (BPG) administered by intramuscular injection [36]. Thus, we summarize the results of the six clinical trials that reported outcomes among nonpregnant adults following treatment with BPG, azithromycin or a combination of BPG and azithromycin (TABLE 2). The oldest data are from a trial in the USA (1993–1997) in which individuals who discovered they had been exposed to infectious stage syphilis through sexual intercourse in the preceding 30 days were given either 1 g azithromycin (n = 40) or BPG (n = 23). Three months post-treatment, rapid plasma reagin (RPR) and fluorescent treponemal antibody absorption tests (FTA-ABS) were negative for all participants in both treatment groups [37]. Another trial in the USA during the same time period was designed to measure treatment outcomes in a population at high risk of contracting STIs/RTIs. Although diagnostic methods were not reported, the trial was suspended after two of the first 12 patients were provided 1 g azithromycin failed their test of cure while all 13 participants were cured using BPG (p = 0.18) [38]. A three-arm trial of early syphilis in the USA then compared treatment outcomes among patients given BPG, or 2 g azithromycin once or 2 g azithromycin two-times with 1 week in between doses. RPR and FTA-ABS testing showed that cure was achieved in 85.7% (n = 14; 95% CI: 60.0–95.7%) of patients given BPG, 94.1% (n = 17; 95% CI: 72.7–98.6%) among recipients of 2 g azithromycin once and 82.8% (n = 29; 95% CI: 65.3–92.3%) in participants who twice received 2 g azithromycin [39].

In sub-Saharan Africa, three trials have investigated BPG versus azithromycin, the first being a community-randomized trial in Uganda (1994–1998) among nonpregnant adults with serological syphilis. Diagnosis and test of cure were based on toluidine red unheated serum tests (TRUSTs) and *Treponema pallidum* hemagglutination assays. Treatment efficacy varied across regimens depending on TRUST titers at enrolment. Among patients with initial titers <1:2, BPG cured 71.0% (n = 93; 95% CI: 61.0–79.2%) of cases compared with 58.5%

Table 1. Effect of curable STIs/RTIs on pregnancy outcomes.

Study (year)	Country	Year(s)	Spontaneous abortion	Stillbirth	IUGR	PROM	Preterm	Low birthweight	Ref.
<i>Treponema pallidum</i>									
Watson-Jones et al. (2002)	Tanzania	1998–2000	NR	18 (5.5–59.6) RR	2.1 (1.0–4.2) RR	NR	6.1 (2.5–15.3) RR	3.3 (2.0–5.4) [†]	[21]
Temmerman et al. (1995)	Kenya	1991	NR	3.34 RR	NR	NR	NR	4.01 [†]	[120]
McDermott et al. (1993)	Malawi	1987–1990	NR	10.98	NR	NR	NR	NR	[19]
Donders et al. (1993)	South Africa	1988	NR	NR	NR	NR	33%; 5 of 15 cases	NR	[34]
Elliott et al. (1990)	Kenya	1985	NR	NR	NR	NR	1.4 (0.5–4.1)	NR	[121]
Ratnam et al. (1982)	Zambia	NR	42% of cases	NR	NR	NR	NR	NR	[15]
Williams et al. (1923)	USA	1923	40% of cases	NR	NR	NR	NR	NR	[14]
<i>Neisseria gonorrhoeae</i>									
Johnson et al. (2011)	USA	1996–2002	NR	NR	NR	NR	2.0 (1.0–4.0)	0.8 (0.3–2.3)	[122]
Donders et al. (1993)	South Africa	1988	NR	NR	NR	NR	56%; 5 of 9 cases	p < 0.005	[34]
Elliott et al. (1990)	Kenya	1985	NR	NR	NR	NR	3.2 (1.3 to 8.4)	NR	[121]
<i>Chlamydia trachomatis</i>									
Rours et al. (2011)	Netherlands	2003–2005	NR	NR	NR	NR	4.4 (1.3–15.2) [†] ; 2.7 (1.1–6.5) [‡] ; 1.17 (0.6–2.4) [§]	1.0 (0.4–2.2)	[32]
Silveira et al. (2009)	USA	2005–2008	NR	NR	NR	NR	0.7 (0.4–1.4)	NR	[123]
Wilkowska-Trojnieł et al. (2009)	Poland	2003–2006	12 versus 2% p = 0.029	NR	NR	NR	NR	NR	[18]
Bias et al. (2007)	USA	2003	NR	NR	NR	1.5 (1.0–2.2) RR	1.5 (1.1–2.0) RR	1.1 (0.7–1.7)	[124]
Odendaal et al. (2006)	South Africa	2002–2003	NR	NR	NR	NR	22.2%; 8 of 36 cases versus 10.4%; 32 of 307 cases; p = 0.037	NR	[30]

Results reported as odds ratios unless otherwise noted and 95% confidence intervals in parentheses.

[†]Preterm delivery before 32 weeks.

[‡]Preterm delivery before 35 week.

[§]Preterm delivery before 37 weeks.

^{||}Bacterial vaginosis at 16–20 weeks.

^{**}Bacterial vaginosis at 28–32 weeks.

^{††}Intermediate flora (Nugent scores 4–7) and bacterial vaginosis (Nugent scores 7–10).

IUGR: Intrauterine growth retardation; NR: Not reported; PROM: Premature rupture of membranes; RR: Risk ratio.

Table 1. Effect of curable STIs/RTIs on pregnancy outcomes (cont.).

Study (year)	Country	Year(s)	Spontaneous abortion	Stillbirth	IUGR	PROM	Preterm	Low birthweight	Ref.
<i>Chlamydia trachomatis</i> (cont.)									
Johnson et al. (2011)	USA	1996–2002	NR	NR	NR	NR	1.0 (0.6–2.0)	2.1 (1.0–4.2)	[122]
Kovacs et al. (1998)	Hungary	1994–1995	NR	NR	7.3 versus 5.8% p > 0.05	20 versus 21% p > 0.05	NR	15.5 versus 13.2% p > 0.05	[125]
Donders et al. (1993)	South Africa	1988	NR	NR	NR	NR	27%; 6 of 22 cases	NR	[34]
Elliott et al. (1990)	Kenya	1985	NR	NR	NR	NR	0.7 (0.4–1.4)	NR	[121]
Johns Hopkins et al. (1989)	USA	1983–1985	NR	NR	2.4 (1.3–4.2)	NR	1.6 (1.0–4.2)	NR	[126]
Gravett et al. (1986)	USA	1983	NR	NR	NR	2.4 (1.7–5.4)	NR	2.7 (1.3–5.7)	[24]
<i>Trichomonas vaginalis</i>									
Johnson et al. (2011)	USA	1996–2002	NR	NR	NR	NR	1.4 (0.7–2.8)	1.5 (0.9–2.6)	[122]
Meis et al. (1995)	USA	1992–1994	NR	NR	NR	NR	1.5 (0.1–8.1) week 24; 0.9 (0.2–3.6) week 28	NR	[127]
Sutton et al. (1999)	DR Congo	1989–1990	NR	NR	NR	NR	NR	2.1 (1.0–4.7)	[35]
Minkoff et al. (1984)	USA	NR	NR	NR	NR	p < 0.03	NR	NR	[128]
Cotch et al. (1997)	USA	1984–1989	NR	NR	NR	NR	1.3 (1.1–1.4)	1.3 (1.1–1.5)	[28]
<i>Bacterial vaginosis</i>									
Johnson et al. (2011)	USA	1996–2002	NR	NR	NR	NR	1.3 (0.9–2.1)	1.1 (0.6–1.8)	[122]
Svare et al. (2006)	Denmark	1998–2002	NR	NR	NR	NR	2.5 (1.6–3.9)	2.0 (1.3–2.9)	[29]
Watson-Jones et al. (2007)	Tanzania	1997–2000	NR	NR	NR	NR	3.0 (1.3–6.6)	NR	[31]
Leitch et al. (2003)	Multiple	Multiple	9.9 (2.0–49.3)	NR	NR	NR	2.2 (1.5–3.1)	NR	[17]
Meis et al. (1995)	USA	1992–1994	NR	NR	NR	NR	1.4 (0.9–2.05) week 24; 1.8 (1.2–3.0) week 28	NR	[127]

Results reported as odds ratios unless otherwise noted and 95% confidence intervals in parentheses.

[†]Preterm delivery before 32 weeks.

[‡]Preterm delivery before 35 week.

[§]Preterm delivery before 37 weeks.

[¶]Bacterial vaginosis at 16–20 weeks.

^{**}Bacterial vaginosis at 28–32 weeks.

^{††}Intermediate flora (Nugent scores 4–7) and bacterial vaginosis (Nugent scores 7–10).

IUGR: Intrauterine growth retardation; NR: Not reported; PROM: Premature rupture of membranes; RR: Risk ratio.

Table 1. Effect of curable STIs/RTIs on pregnancy outcomes (cont.).

Study (year)	Country	Year(s)	Spontaneous abortion	Stillbirth	IUGR	PROM	Preterm	Low birthweight	Ref.
<i>Bacterial vaginosis (cont.)</i>									
McGregor et al. (1995)	USA	1991–1992	NR	NR	NR	3.5 (1.4–8.9) RR	1.9 (1.2–3.0); 1.5 (0.7–3.0) [§] RR	NR	[25]
Hillier et al. (1995)	USA	1984–1989	NR	NR	NR	1.1 (0.8–1.6)	1.4 (1.1–1.8)	1.5 (1.2–1.7)	[129]
Hay et al. (1994)	UK	NR	5.5 (2.3–13.3) ^{††}	NR	NR	NR	13.1 (4.0–42.6) ^{**}	NR	[16]
Elliott et al. (1990)	Kenya	1985	NR	NR	NR	NR	1.0 (0.6–1.8)	NR	[121]
Gravett et al. (1986)	USA	1983	NR	NR	NR	2.0 (1.1–3.7)	NR	1.5 (0.8–2.0)	[24]

Results reported as odds ratios unless otherwise noted and 95% confidence intervals in parentheses.
^{*}Preterm delivery before 32 weeks.
[†]Preterm delivery before 35 weeks.
[‡]Preterm delivery before 37 weeks.
[§]Bacterial vaginosis at 16–20 weeks.
^{||}Bacterial vaginosis at 28–32 weeks.
^{**}Intermediate flora (Nugent scores 4–7) and bacterial vaginosis (Nugent scores 7–10).
^{††}IUGR: Intrauterine growth retardation; NR: Not reported; PROM: Premature rupture of membranes; RR: Risk ratio.

(n = 94; 95% CI: 48.4–68.0%) among recipients of 1 g azithromycin and 70.6% (n = 313; 95% CI: 65.3–75.4%) of participants given 1 g azithromycin plus BPG. If titers at enrolment were >1:4, the efficacy of BPG was reduced to 41.3% (n = 75; 95% CI: 30.9–52.7%). Treatment efficacy was also lower among groups given azithromycin but higher than BPG alone. Recipients of 1 g azithromycin alone had a cure rate of 53.3% (n = 71; 95% CI: 42.0–64.7%), whereas 1 g azithromycin plus BPG cured 54.7% of cases (n = 309; 95% CI: 49.1–60.2%) [40].

These results were followed by a trial carried out in Tanzania (2000–2003) among patients who were recruited by screening high-risk populations. All 328 subjects had a titer of at least 1:8 on RPR test; 106 had baseline titers of >1:64, levels indicative of active syphilitic lesions. Confirmed by RPR test and *T. pallidum* particle agglutination assay, serological cure was observed in 97.5% (n = 163; 95% CI: 93.9–99.0%) of participants given 2 g azithromycin versus 95.2% (n = 165; 95% CI: 90.7–97.5%) in the BPG group [41].

The most recent study comparing the efficacy of azithromycin versus BPG is a multicenter trial (2000–2007) in Madagascar (n = 421) and North America (n = 94) among HIV-negative patients with early syphilis. Based on RPR testing, serological cure was reported in 77.6% of subjects given 2 g azithromycin (n = 232; 95% CI: 71.8–82.5%) and 78.5% (n = 237; 95% CI: 72.8–83.3%) in the BPG group. Nonserious adverse events were reported by 61.5% (n = 174; 95% CI: 55.7–67.0%) of individuals treated with 2 g azithromycin, most of whom had self-limiting gastrointestinal discomfort, whereas 46.1% (95% CI: 40.6–52.1%) of BPG recipients reported nonserious adverse events [42].

In vitro evidence

Fourteen *in vitro* studies met our inclusion criteria, seven with isolates from low-risk populations (TABLE 3) and seven from high-risk or mixed-risk groups (TABLE 4). A report from San Francisco in 2001 was the first to associate azithromycin treatment failure with A→G mutations at the 2,058 position of the 23S rRNA gene of *T. pallidum* [43]. Retrospective analysis of samples revealed that 4.0% (n = 25; 95% CI: 0.9–19.6%) of isolates had A→G mutations between 1999 and 2002. In 2003, the proportion of isolates with A→G mutations increased to 36.7% (n = 30; 95% CI: 21.9–54.6%) [44]; by 2004, 56.1% (n = 66; 95% CI: 44.0–67.3%) had selected for resistance [43]. However, in Dublin, 88.2% (n = 17; 95% CI: 65.3–96.4%) of isolates already had A→G mutations by 2002 [44].

Macrolide resistance is strongly associated with use by an individual in the previous year. Isolates from Seattle (2001–2005) were two-times more likely to be resistant if patients had been treated with macrolides in the past 12 months (RR: 2.2; 95% CI: 1.1–4.4; p = 0.02) [45]. This relationship persisted over the decade. A2058G and A2059G mutations, which are associated with clinical failures of azithromycin, were found in 88.9% (n = 36; 95% CI: 74.6–95.5%) of isolates from 2001 to 2010 among patients exposed to macrolides in the preceding

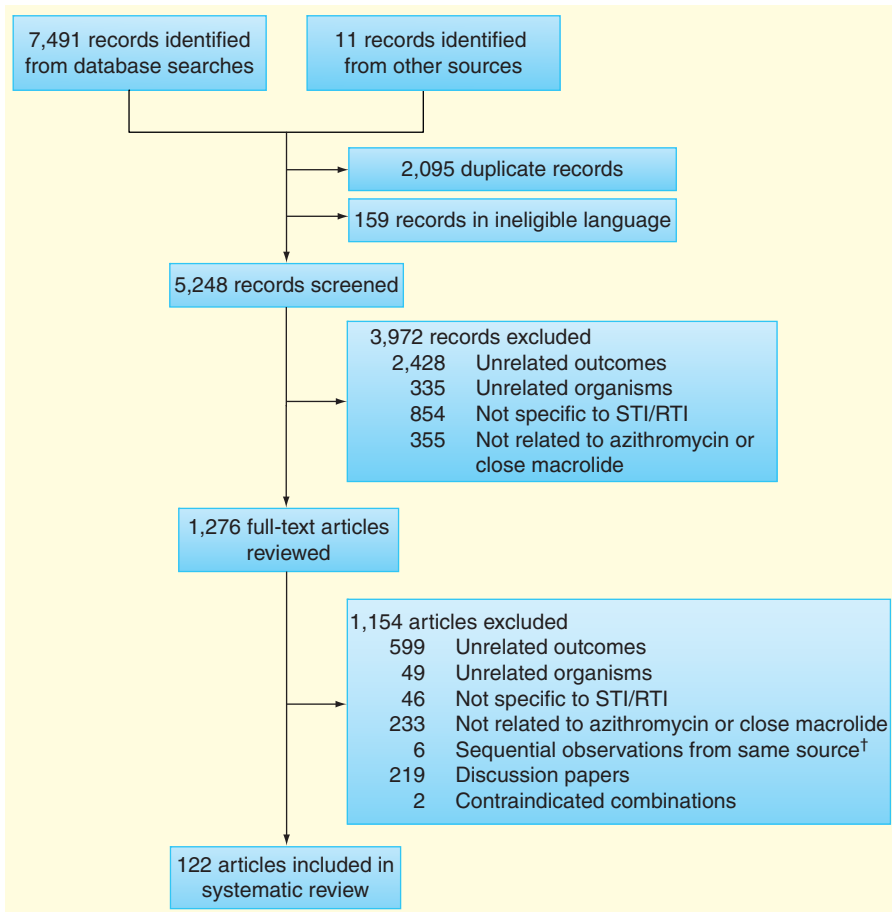


Figure 1. Identification, screening and eligibility of studies included in systematic review.

†Surveillance reports from which the most recent data set was used.

STI/RTI: Sexually transmitted infection and reproductive tract infection.

12 months, whereas 61.2% ($n = 98$; 95% CI: 51.3–70.4%) of isolates from patients who had not received prior macrolide treatment contained the same mutations [46]. Similar mutations were found among strains of *T. pallidum* in eight cities across China (2008–2011). A2058G was present in 97.0% individuals who had taken macrolides in the previous 12 months versus 62.5% of patients who had not ($n = 211$; OR: 19.65; 95% CI: 5.8–66.9) [47]. The opposite was found in Taiwan (2009–2011) where no single A2058G or A2059G mutation was seen among 211 isolates tested from a population where only one person had been given macrolide therapy in the previous year [48]. Similarly, there was no evidence of resistance among 141 amplified samples from HIV-negative heterosexual patients in Madagascar [49]. Although use of macrolides in the previous year was not reported, the Essential Drugs List of the Malagasy Ministry of Health does not include macrolides [301].

Neisseria gonorrhoeae

In vivo evidence

The WHO recommends treating pregnant women with *Neisseria gonorrhoeae* infection using 400 mg cefixime as a single dose

or 125 mg ceftriaxone by intramuscular injection [50]. However, azithromycin has been used for the treatment of gonorrhoea among nonpregnant adults during the past two decades. Eleven trials were identified through our review (TABLE 5). Nine trials conducted between the late 1980s and 1999 investigated the use of 1 g azithromycin among individuals attending sexually transmitted infection (STI) clinics. Of these, three were open label without comparators [51–53] and six were two-arm trials that compared azithromycin to ciprofloxacin and/or doxycycline [54–59]. The pooled efficacy of azithromycin against *N. gonorrhoeae*, estimated using random effects models [60], was 97.0% ($n = 539$; 95% CI: 95.5–98.5%). This is slightly higher than 96.5% ($n = 539$; 95% CI: 94.3%–97.6%) reported in a 2010 review [61] that added numerators and divided the sum of denominators among the same nine trials. Such an approach does not account for heterogeneity across study populations and gives equal weight to all trials regardless of their precision. Regardless of pooling methods, it is unlikely that the same efficacy would be observed today using 1 g azithromycin in high-income countries following 25 years of cumulative drug pressure. However, the epidemiological context in sub-Saharan Africa is likely different

where azithromycin use has been almost exclusively limited to trachoma eradication campaigns [62].

We identified two RCTs that investigated the use of 2 g azithromycin among patients at STI clinics. The first was a multicenter trial in the USA (1991–1992) in which 98.9% ($n = 374$; 95% CI: 97.3–99.6%) of patients were cured [63]. A similar RCT in New Delhi (2005–2006) involved 42 participants; loss to follow-up was high, 52.4%, but all 22 subjects who returned for a test of cure had their *N. gonorrhoeae* infections cured [64].

In vitro evidence

Over the past decade, *in vitro* studies have documented the loss of *N. gonorrhoeae* sensitivity to azithromycin. There are no standard breakpoints of minimum inhibitory concentrations (MICs) used to categorize *N. gonorrhoeae* resistance to azithromycin, but $>1 \mu\text{g/ml}$ [65] and $>2 \mu\text{g/ml}$ [66] have both been used. In this section, we summarize the key regional observations from 36 *in vitro* studies (TABLES 6 & 7).

The Public Health Agency of Canada reported that 0.17% ($n = 40,875$; 95% CI: 0.001–0.002%) of *N. gonorrhoeae*

Table 2. Randomized clinical trials of azithromycin versus benzathine penicillin G for the treatment of *Treponema pallidum*

Study (year)	Country	Year(s)	Regimen	Number cured of treated	Percent cured	95% CI	Diagnostic methods used	Follow up	Stage of infection	Ref.
<i>Specified low-risk populations</i>										
Hook et al. (2010)	Madagascar, USA	2000–2007	2 g azithromycin 2.4 mu BPG	180/232 186/237	77.6% 78.5%	71.8–82.5% 72.8–83.2%	RPR and FTA-ABS	6 months	Primary, secondary, early latent syphilis	[42]
Kiddugavu et al. (2005)	Uganda	1994–1998	1 g azithromycin 2.4 mu BPG 1 g azithromycin plus 2.4 mu BPG	55/94 66/93 221/313	58.5% 71.0% 70.6%	48.4–68.0% 61.0–79.2% 65.3–75.4%	TRUST and TPHA (initial TRUST titers ≤ 1:2)	10 months	Serologic syphilis	[130]
			1 g azithromycin 2.4 mu BPG 1 g azithromycin plus 2.4 mu BPG	38/71 31/75 169/309	53.3% 41.3% 54.7%	42.0–64.7% 30.9–52.7% 49.1–60.2%	TRUST and TPHA (initial TRUST titers ≥ 1:4)	10 months	Serologic syphilis	
<i>Unspecified low-risk populations</i>										
Klausner et al. (2006)	USA	2004	1 g azithromycin 2.4 mu BPG	10/12 13/13	83.3% 100%	54.6–95.0% NA	NR	NR	Exposed to infectious syphilis	[38]
Hook et al. (2002)	USA	1995–1997	2 g azithromycin 2 g azithromycin (two-times) week apart 2.4 mu BPG	14/14 19/22	100% 86.4%	NA 66.4–95.0%	RPR and MHA-TP or FTA-ABS	12 months	Primary, secondary, early latent syphilis	[39]
Hook et al. (1999)	USA	1995–1997	1 g azithromycin 2.4 mu BPG	40/40 23/23	100% 100%	NA NA	RPR and FTA-ABS	3 months	Exposed to infectious syphilis	[131]
<i>High-risk population</i>										
Riedner et al. (2005)	Tanzania	2000–2003	2 g azithromycin 2.4 mu BPG	159/163 157/165	97.5% 95.2%	93.9–99.0% 90.7–97.5%	RPR and PCR	9 months	Primary, secondary, higher titer latent syphilis	[132]

2.4 mu BPG: 2.4 million units benzathine penicillin G; FTA-ABS: Fluorescent treponemal antibody absorption; MHA-TP: Microhemagglutination assay-*Treponema pallidum*; NA: Not applicable; NR: Not reported; RPR: Rapid plasma reagin; TPHA: *Treponema pallidum* hemagglutination; TRUST: Toluene red unheated serum test.

Table 3. Sensitivity testing of *T. pallidum* isolates to azithromycin and other macrolides.

Study (year)	Country	Year(s)	Specimen source	Sample size	Sample	Resistant mutation	Number resistant	Percent resistant	95% CI	Additional details	Ref.
<i>Low-risk populations</i>											
Chen et al. (2013)	USA	2007–2009	Surveillance	129		2058A 2058G 2059G	83 67 17	64.3% 51.9% 13.2%	55.8–72.1% 43.4–60.4% 8.4–20.1%	From patients with primary of secondary syphilis attending STI clinics	[133]
Van Damme et al. (2009)	Madagascar	NR	Randomized clinical trial	186		23S rRNA	0	0.0%	NA	DNA of <i>T. pallidum</i> was detected in 141 samples; 61% of patients were male; 98% were heterosexual	[49]
<i>Unspecified low-risk populations</i>											
Miller et al. (2012)	South Africa Lesotho	2005–2010	Surveillance	100		A2058G A2059G	1 0	1.0% 0.0%	0.0–5.4%	117 ulcer specimens collected of which 100 were positive for <i>T. pallidum</i>	[134]
A2058G Prevalence Workgroup (2012)	USA	2007–2009	Surveillance	141		A2058G	75	53.1%	45.0–61.2%	From patients with primary of secondary syphilis attending STI clinics; MSM were nearly 6 times more likely to have resistant syphilis compared with heterosexual women and men	[135]
Matejková et al. (2009)	Czech Republic	2005–2008	Passive case detection	22		23S rRNA A2058G A2059G	8 4 4	36.4% 18.2% 18.2%	19.7–57.3% 7.5–38.8% 7.5–38.8%		[136]
Martin et al. (2009)	China	2007–2008	Passive case detection	38		A2058G	38	100%	NA	Patients presenting to STI clinic with symptoms compatible with primary syphilis	[137]
Lukehart et al. (2004)	USA	1999–2002 2003 2001–2003 1998–2000	Surveillance	25 30 23 19		23S rRNA	1 11 3 2	4.0% 36.7% 13.0% 10.5%	1.0–19.6% 21.9–54.6% 4.7–32.3% 3.2–31.7%	PCR to detect 23S rRNA gene mutations; confirmation of azithromycin resistance was conducted through intradermal rabbit inoculation	[44]
	Ireland										
	Dublin	2002	Surveillance	17		23S rRNA	15	88.2%	65.3–96.4%		
	Multiple locations	1912–1987	Historical	18		23S rRNA	1	5.6%	1.3–2.6%		

DNA: Deoxyribonucleic acid; MSM: Men who has sex with men; NA: Not applicable; NR: Not reported; STI: Sexually transmitted infection.

Table 4. Sensitivity testing of *T. pallidum* isolates to azithromycin and other macrolides.

Study (year)	Country	Year(s)	Specimen source	Sample size	Resistant mutation	Number resistant	Percent resistant	95% CI	Additional details	Ref.
<i>High-risk populations</i>										
Tipple et al. (2011)	UK	2006–2008	Cross-sectional survey	18	23S rRNA A2058G	12	66.6%	43.4–83.7%	Specimens from men, 94.1% were MSM	[138]
Rekart et al. (2003)	Canada	2000	Mass drug administration	25	23S rRNA	0	0.0%	NA	1.8 g azithromycin given to sex workers and clients (n = 4,384)	[139]
<i>Mixed-risk populations</i>										
Chen et al. (2012)	China	2008–2011	Cross-sectional survey	211	A2058G	194	91.9%	87.2–95.1%	391 samples collected; 6.1% from FSW, 71.8% reported sex with FSW and 1.4% were MSM	[47]
Muldoon et al. (2012)	Ireland	2009–2010	Cross-sectional survey	29	A2058G	27	93.1%	77.9–97.9%	34.6% (36/104) of samples were positive for <i>T. pallidum</i> by PCR; 29 sequenced	[140]
Martin et al. (2010)	Canada	2007–2009	Cross-sectional survey	43	A2058G	7	16.3%	8.2–30.1%	449 samples collected from 374 patients; 43 were positive for <i>T. pallidum</i> and sequenced	[141]
Mitchell et al. (2006)	USA	2000–2002 2003 2004	Retrospective study	25 32 66	23S rRNA	1 13 37	4.0% 40.6% 56.1%	1.0–19.6% 25.5–57.9% 44.0–67.4%	Patients (n = 1,308) diagnosed primary or secondary syphilis; all treatment failure and resistance in MSM/bisexual patients	[43]
Morshed et al. (2006)	Canada	2000–2003 2004	Retrospective study	47 9	23S rRNA	1 4	2.1% 44.4%	0.5–11.1% 18.7–73.8%	MSM patients presenting to STI clinic with primary or secondary syphilis	[142]

FSW: Female sex worker; MSM: Men who have sex with men; NA: Not applicable; STI: sexually transmitted infection.

Table 5. Randomized clinical trials of azithromycin for the treatment of *Neisseria gonorrhoeae*

Study (year)	Country	Year(s)	Azithromycin dose	Number cured of evaluated	Percent cured	95% CI	Diagnostic methods	Additional details	Ref.
<i>Dose of 2 g azithromycin</i>									
Khaki et al. (2007)	India	2005–2006	2 g	22/22	100%	NA	>Gram stain TOC days 5–7		[143]
Handsfield et al. (1994)	USA	1991–1992	2 g	370/374	98.9%	97.3–99.6%			[63]
<i>Dose of 1 g azithromycin</i>									
Rustomjee et al. (2002)	South Africa	1999	1 g	30/31	96.8%	83.8–99.2%	LCR for NG/CT TOC day 14	100% (n = 21) NG infections cured 100% (n = 14) CT infections cured 90.0% (n = 10; 95% CI: 58.7–97.8%) co-infections cured	[60]
Swanston et al. (2001)	Trinidad and Tobago	1996	1 gram	125/127	98.4%	94.5–99.5%	ELISA for NG culture for CT; TOC days 7–10	100% (n = 115) NG infections cured 95.7% (n = 23; 95% CI: 78.9–99.0%) CT infections cured 88.3% (n = 12; 95% CI: 54.6–95.0%) co-infections cured	[53]
Gruber et al. (1997)	Croatia	1994–1995	1 g	48/50	96.0%	86.5–98.8%	Culture and gram stain for NG; TOC day 14		[58]
Gruber et al. (1995)	Croatia	1991–1993	1 g	24/24	96.0%	80.4–99.1%	Culture and gram stain for NG; TOC day 14		[57]
Steingrímsson et al. (1994)	Iceland	NR	1 g	27/28	96.4%	82.4–99.2%	Culture for NG w/ DFA; Culture for CT; TOC day 28	92.4% (n = 79; 84.4–96.4%) CT infections cured	[56]

CT: *Chlamydia trachomatis*; DFA: Direct fluorescent antibody; ELISA: Enzyme-linked immunosorbent assay; LCR: Ligase chain reaction; mg: milligrams; NA: Not applicable; NG: *Neisseria gonorrhoeae*; NR: Not reported; TOC: Test of cure.

Table 5. Randomized clinical trials of azithromycin for the treatment of *Neisseria gonorrhoeae*

Study (year)	Country	Year(s)	Azithromycin dose	Number cured of evaluated	Percent cured	95% CI	Diagnostic methods	Additional details	Ref.
<i>Dose of 1 g azithromycin (cont.)</i>									
Waugh <i>et al.</i> (1993)	UK	1990–1991	1 g	85/89	95.5%	89.0–98.2%	Culture and gram stain for NG; culture for CT; TOC day 10	100% (n = 22) NG/CT co-infections cured	[52]
Odugbemi <i>et al.</i> (1993)	Nigeria	1989–1990	1 g	114/120	95.0%	89.5–97.6%	Culture for NG; TOC day 14		[51]
Steingrímsson <i>et al.</i> (1990)	Iceland	NR	1 g (day 0)	11/12	91.7%	64.0–98.1%	Culture for NG/CT; TOC day 28	97.7% (n = 44; 88.2–99.5%) CT infections cured	[55]
			500 mg (day 0 x 2)	7/8	87.5%	51.8–97.2%		96.3% (n = 27; 81.7–99.1%) CT infections cured	
			500 mg (day 0) 250 mg (days 1 and 2)	7/7	100%	NA		88.0% (n = 25; 69.8–95.6%) CT infections cured	
Lassus <i>et al.</i> (1990)	Finland	NR	1 g (day 0)	15/15	100%	NA	Culture and gram stain for NG/CT w/ DFA; TOC day 14	100% (n = 12) CT infections cured	[54]
			500 mg (day 0) 250 mg (days 1 and 2)	14/14	100%	NA		100% (n = 5) co-infections cured	
								100% (n = 9) CT infections cured	
								83.3% (n = 6; 95% CI: 42.1–96.3%) co-infections cured	

CT: *Chlamydia trachomatis*; DFA: Direct fluorescent antibody; EUSA: Enzyme-linked immunosorbent assay; LCR: Ligase chain reaction; mg: milligrams; NA: Not applicable; NG: *Neisseria gonorrhoeae*; NR: Not reported; TOC: Test of cure.

Table 6. Sensitivity testing of *Neisseria gonorrhoeae* isolates to azithromycin.

Study (year)	Country	Year(s)	Sample size	MIC range ($\mu\text{g/ml}$)	Percent strains susceptible	Additional details	Ref.
Olsen <i>et al.</i> (2013)	Vietnam	2011	108	NR	62%	11% isolates fully resistant, 29% intermediate susceptibility	[144]
Lahra <i>et al.</i> (2012)	Australia	2011	3,293	≤ 4	98.1%	Isolates from all states in Australia	[145]
Sethi <i>et al.</i> (2013)	India/Pakistan/Bhutan	2007–2011	65	0.016–4.0	76.9%	7.7% isolates fully resistant, 15.4% intermediate susceptibility	[72]
Lo <i>et al.</i> (2012)	Hong Kong	2010	485	<0.25 to >256	69.7%		[146]
Lefebvre <i>et al.</i> (2011)	Canada	2010	831	≤ 16	98.7%		[147]
Hottes <i>et al.</i> (2013)	Canada	2006–2011	1,837	0.064–16	99%	Elevated MIC showed increasing trend over time	[148]
CDC (2011)	USA	2002–2009	87,566		99.9%	39 (0.04%) had MICs ≥ 8 $\mu\text{g/ml}$ (25 with 8 $\mu\text{g/ml}$; 14 with 16 $\mu\text{g/ml}$)	[69]
Yuan <i>et al.</i> (2011)	China	2008–2009	318	NR	94.7%		[149]
Takahashi <i>et al.</i> (2013)	Japan	2007–2009	52	0.016–1	100%	100% of isolates from men [†]	[150]
Herciline <i>et al.</i> (2010)	USA	2006–2008	286	0.032–1.0	99.0%	Median MIC 0.125 $\mu\text{g/ml}$	[151]
Cole <i>et al.</i> (2010)	Europe (17 countries)	2006–2008	3,528	≤ 256	2.0–7.0%	High resistance (>256 $\mu\text{g/ml}$) in 4 isolates from Scotland and 1 in Ireland	[68]
Olsen <i>et al.</i> (2012)	Guinea-Bissau	2006–2008	31	NR	100%	Of resistant strains, two had MIC >64 $\mu\text{g/ml}$	[152]
Tanaka <i>et al.</i> (2011)	Japan	2001–2009	242	0.004–1.0	99.9%	Modal shift of MIC was 0.25–0.5 $\mu\text{g/ml}$	[153]
Martin <i>et al.</i> (2011)	Canada	2000–2009	40,875	≤ 64	99.8%	100% isolates susceptible also to sefexime, ceftriaxone and spectinomycin.	[66]
Bala <i>et al.</i> (2011)	India	2000–2009	274	NR	99.7%	One isolate was resistant to azithromycin, quinolones and penicillin	[154]
Chisholm <i>et al.</i> (2008)	UK	2001–2007	108	<0.03 to >256	94.0%	6/108 isolates had MIC >256 $\mu\text{g/ml}$; shift to high level resistance	[155]
Khaki <i>et al.</i> (2007)	India	2004–2005	60	0.016–0.25	100%		[156]
Enders <i>et al.</i> (2006)	Southern Germany	2004–2005	65	0.016–5.0	100%	100% of isolates susceptible to azithromycin	[157]

Unspecified low-risk populations.

[†]Sexual practices were not reported; isolates are assumed not to be from men who have sex with men;

[‡]Dislocated males workers are among high-risk populations.

NR: Not reported.

Table 6. Sensitivity testing of *Neisseria gonorrhoeae* isolates to azithromycin (cont.).

Study (year)	Country	Year(s)	Sample size	MIC range ($\mu\text{g/ml}$)	Percent strains susceptible	Additional details	Ref.
Vorobieva <i>et al.</i> (2007)	Russia	2004	76	NR	100%	Although all susceptible, reduced susceptibility seen in 14%	[158]
Sutrisna <i>et al.</i> (2006)	Indonesia	2004	163	NR	100%	53% resistant to ciprofloxacin	[159]
Martin <i>et al.</i> (2004)	Western Europe	2004	965	NR	91.8%	Variation from 31.2% (Austria 30/96) to 0% (France 0/101, Greece 0/79, Portugal 0/17)	[160]
Chaudhary <i>et al.</i> (2005)	Nepal	2003	16	0.008–0.5	100%	No isolates resistant, but 3/16 (19%) had reduced susceptibility	[161]
Chen <i>et al.</i> (2009)	Taiwan	1999–2004	65	NR	100%		[162]
Hsueh <i>et al.</i> (2005)	Taiwan	1999–2003	55	0.03–9.0	72.7%		[163]
Aydin <i>et al.</i> (2005)	Turkey	1998–2002	78	0.004–0.25	100%	100% of isolates from men [†]	[164]
Moodley <i>et al.</i> (2001)	South Africa	1995–2000	58	0.015–1.0	100%	37% (37/100) strains resistant to penicillin; tetracyclines had reduced susceptibility.	[165]
Kobayashi <i>et al.</i> (2003)	Japan	1995–1999	699	0.015–1	100%	100% of isolates from men [†]	[166]
Llanes <i>et al.</i> (2003)	Cuba	1995–1999	52	0.064–0.5	76.9%	23.1% intermediate susceptibility: MIC= 0.125 (10 isolates) MIC= 0.5 (2 isolates)	[167]
Sosa <i>et al.</i> (2003)	Cuba	1995–1999	91	0.063–4.0	90.1%	Isolates with reduced susceptibility also resistant to penicillin and tetracycline	[168]
Dillon <i>et al.</i> (2001)	Brazil	1998	81	0.032–0.5	100%	28% reduced susceptibility	[169]
Zarantonelli <i>et al.</i> (1999)	Uruguay	1996–1997	51	0.032–0.5	100%	Decreased susceptibility (MIC 0.025 to 0.5) in 72%; isolates from men [†]	[170]
Young <i>et al.</i> (1997)	Scotland	1996	67	0.023–0.75	100%	Isolates randomly selected from with penicillin MIC \geq 1	[171]
Mehaffey <i>et al.</i> (1996)	USA	NR	105	0.06–2.0	100%	Two tests compared. Data from agar dilution method not Etest.	[172]
Dillon <i>et al.</i> (2001)	Guyana & St Vincent	1992–1996	136	0.032–8.0	85.5 and 97.0%	Two isolates had MIC = 8 $\mu\text{g/l}$. 49% (67/137) reduced susceptibility (combined)	[173]
Van Rijsoort-Vos <i>et al.</i> (1995)	Netherlands	1991–1993	114	0.03–1.0	100%	One isolate reduced susceptibility	[174]
Ison <i>et al.</i> (1993)	South Africa	1989–1990	192	0.03–1.0	100%	Study in migrant mine workers (men only) [†]	[175]

Unspecified low-risk populations.

[†]Sexual practices were not reported; isolates are assumed not to be from men who have sex with men;

[‡]Dislocated males workers are among high-risk populations.

NR: Not reported.

Table 7. Sensitivity testing of *N. gonorrhoeae* isolates to azithromycin.

Study (year)	Country	Year(s)	Sample size	MIC range ($\mu\text{g/ml}$)	Percent of strains susceptible	Additional details	Ref.
<i>High-risk populations</i>							
CDC (2011)	USA	2009	55	NR	90.9%	9.1% resistant (95% CI: 4.0 to 19.6%); of 5 resistant (all from MSM), 3 had MIC = 8 $\mu\text{g/ml}$ and two had MIC = 16 $\mu\text{g/ml}$	[69]
Starnino <i>et al.</i> (2009)	Italy	2007–2008	219	1.0–256.0	90.0%	72.7% (95% CI: 51.6–86.8%) of resistant isolates from MSM	[176]
Donegan <i>et al.</i> (2004)	Bali	2004	147	0.013–0.512	100%	Study in FSWs; prevalence of NG estimated to be 35–60%	[177]
Morris <i>et al.</i> (2009)	USA	2000–2002	79	0.03–0.5	100%	Increased MIC values seen in MSM subject isolates	[178]
Leven <i>et al.</i> (2003)	Indonesia	1996	267	0.032–0.5	100%	Study in FSWs; prevalence of NG estimated to be 18–44%	[179]
CDC (2000)	USA	1999	12	1.0–4.0	NR	Median MIC was 2.0 $\mu\text{g/ml}$. 6 of 12 samples were from men who had sex with a CSW; 2 of 12 were from HIV positive men	[180]
<i>Mixed-risk populations</i>							
Bruck <i>et al.</i> (2012)	UK	2005–2006	147	NR	99.3%	Mixed male heterosexual, MSM and female heterosexual isolates	[181]
Dan <i>et al.</i> (2010)	Israel	2002–2007	406	0.023–8.0	91.8%	Mixed male heterosexual, MSM and female heterosexual; resistance to azithromycin did not appear to rise over 5 year period	[182]
McLean <i>et al.</i> (2004)	USA	1999–2001	1,248	≤ 4	97.4%	Mixed high- and low-risk population. Median MIC was 2.0 $\mu\text{g/ml}$	[183]
Arreaza <i>et al.</i> (2003)	Spain	1992–2001	63	0.03–4.0	96.8%	58.7% of strains had reduced susceptibility (0.25–1.0 $\mu\text{g/ml}$). 50% of resistant isolates were from FSW	[184]

BV: Bacterial vaginosis; CSW: Commercial sex worker (gender not specified); FSWs: Female sex workers; GI: Gastrointestinal; MIC: Minimum inhibitory concentration; MSM: Men who have sex with men; NG: *Neisseria gonorrhoeae*; NR: Not reported; PROM: Preterm premature rupture of the membranes; SD: Standard deviation; TOC: Test of cure.

samples were resistant to azithromycin between 2000 and 2009, although the modal value of the MIC shifted from 0.25 $\mu\text{g/ml}$ in 2001 to 0.5 $\mu\text{g/ml}$ between 2007 and 2009 [67]. During the same 10-year period, the Centers for Disease Control and Prevention in the USA reported that 0.04% (n = 87,566; 95% CI: 0.03–0.06%) of *N. gonorrhoeae* isolates tested had MICs $\geq 8 \mu\text{g/ml}$ (including 25 with 8 $\mu\text{g/ml}$ and 14 with 16 $\mu\text{g/ml}$) [68]. This did not include five cases of azithromycin-resistant *N. gonorrhoeae* found between August and October 2009 among men who have sex with men; three had MICs of 8 $\mu\text{g/ml}$ and two had 16 $\mu\text{g/ml}$ [69]. Resistance may have appeared in Europe slightly before North America. Analysis of isolates from 17 European countries found that 3.2% (n = 836;

95% CI: 0.02–0.05%) of gonococcal isolates were resistant to azithromycin in 2006. By 2007, 6.8% (n = 973; 95% CI: 0.05–0.09) of samples were resistant. The overall proportion of resistant isolates declined in 2008 to 1.8% (n = 940; 95% CI: 0.01–0.03%), although only 5.2% (95% CI: 4.1–6.8%) of strains tested in the same year were fully susceptible to azithromycin and ciprofloxacin. Four isolates from Scotland and one from Ireland exhibited MICs $>256 \text{ mg/l}$ [68].

Gonococcal isolates examined from South America and Cuba exhibited high but stable levels of resistance between 2000 and 2009 in most settings [70]. Collectively, azithromycin resistance was 13.0% (n = 8,373; 95% CI: 12.3–13.7%) based on data from six countries including Chile, an outlier.

Averaged over the decade, 26.7% (n = 3,116; 95% CI 25.2–28.3%) of samples from Chile were resistant, rising to 45.6% (n = 463; 95% CI: 41.1–50.1%) according to the most recent data from 2009. Removing Chile from the regional summary, 4.4% (n = 5,257; 95% CI: 3.9–5.1%) of isolates were resistant over the decade.

All 60 gonococcal isolates from India between 2004 and 2005 were susceptible to azithromycin [71]. Pooled analysis of samples collected from India, Pakistan and Bhutan between 2007 and 2011 found that 76.9% (n = 65; 95% CI: 65.3–85.5%) were susceptible. Results were not stratified by country and, therefore, it is not known whether the sensitivity of isolates from India had changed [72]. Applying the more conservative breakpoint of >1 µg/ml to the *in vitro* studies identified in this review, 35% (7 of 20) of the *in vitro* studies reported upper range MICs that included gonococcal isolates resistant to azithromycin. This percentage does not include 16 studies we identified and included in TABLES 6 & 7 that did not report MICs.

Chlamydia trachomatis

In vivo evidence

The WHO recommends treating pregnant women with *Chlamydia trachomatis* infection using 1 g azithromycin as a single oral dose [50]. We found eight RCTs in the literature that reported the treatment efficacy of 1 g azithromycin among pregnant women (TABLE 8) [73–80]. Using random effect models, we estimate the pooled treatment efficacy to be 92.1% (n = 268; 95% CI: 88.4–95.7%). The estimated efficacy would be higher if we excluded two trials that were conducted in the USA. The first trial (1995–1997) reported a 3-week test of cure rate to be 88.1% (n = 42; 95% CI: 74.9–94.7%) [76], whereas the second trial (1998–2000) was terminated early due to poor efficacy, 63.3% (n = 55; 95% CI: 50.4–75.1%), based on test of cure ≥ 4 weeks post-treatment [74]. These results need to be interpreted with caution because no distinction was made between treatment failures and new infections, sex partners were not treated by trial staff, but were referred to a treatment center, and only 35% of women were seen within 7 days of the scheduled test of cure.

Studies investigating sexual activity following treatment offer some perspectives on post-treatment infections and the extent to which they may be failures or *de novo* infections. A trial in Seattle (1998–2003) found that persistent or recurrent chlamydial or gonorrheal infection occurred in 7.6% (n = 289; 95% CI: 5.1–4.9%) of female patients who reported no sexual intercourse after treatment [81]. Another study reported that 19.0% (n = 79; 95% CI 11.9–29.0%) of women were positive for *C. trachomatis* 3 months after treatment using 1 g azithromycin. Of these women, 13.3% (n = 15; 95% CI: 4.0–38.3%) reported being sexually inactive during the post-treatment period [82]. These findings may be attributable to false reporting of sexual contact, or treatment failure or may lend credence to the hypothesis that *C. trachomatis* enters a latent asymptomatic state that is undetectable by culture or, possibly, Nucleic Acid Amplification Tests, but can later reactivate [83].

In vitro evidence

Thresholds for antimicrobial susceptibility and resistance of *C. trachomatis* are not universally standardized, although MICs >4 µg/ml are often used to characterize therapeutic failure [84–87]. The lowest concentration of antimicrobial compound needed to inhibit chlamydial formation is between 0.03 and 0.125 µg/ml, whereas the minimum bactericidal concentration (MBC; also referred to as the minimum chlamydicidal concentration, or MCC) is between 0.06 and 0.5 µg/ml [88,89]. The published *in vitro* studies of azithromycin collectively suggest the persistence of high and widespread treatment efficacy (TABLE 9). One noted exception is the study of six isolates from three patients who experienced treatment failure in Russia (2000–2002); four isolates were resistant to azithromycin, doxycycline and ofloxacin at MICs and MBCs >5.12 µg/ml [90]. Not surprisingly, *in vitro* resistance appears to be more common in individuals with greater severity of disease or recurrent disease. A study in the USA during the early 1990s described decreased susceptibility and emerging resistance to azithromycin and doxycycline in isolates from women with mucopurulent cervicitis but not in isolates from women with asymptomatic infections [91]. Similar observations were reported in 2010 from India; six of eight isolates with modified susceptibility had been obtained from recurrently infected individuals, whereas the remaining two were from nonrecurrently infected patients. MICs and MBCs for azithromycin were 8 µg/ml for two of the patients from which the modified susceptibility isolates were taken. One individual had chronic cervicitis and the other had pelvic inflammatory disease [92].

Trichomonas vaginalis

In vivo evidence

Trichomonas vaginalis is a protozoal infection which causes cervicitis and nongonococcal urethritis. The WHO recommends treating pregnant women with *T. vaginalis* infection after the first trimester using 2 g metronidazole orally as a single dose, or 400–500 mg twice daily for 7 days or 300 mg clindamycin orally twice a day for 7 days [36]. If treatment is imperative during the first trimester of pregnancy, the single-dose regimen of 2 g metronidazole orally is recommended [36]. Azithromycin has not been used directly for prevention or treatment purposes because *T. vaginalis* is anaerobic. Nevertheless, azithromycin has demonstrated protection against *T. vaginalis* in studies of mass STI/RTI treatment (TABLES 10 & 11).

In Kenya (1998–2002), 1 g azithromycin or placebo was given once per month to 466 HIV-negative female sex workers [93]. At the end of the trial, HIV incidence was the same across treatment groups, the primary endpoint, but the incidence of *T. vaginalis* was reduced significantly among those given azithromycin versus placebo (RR: 0.56; 96% CI: 0.40–0.78; p < 0.001). A similar observation was made in a three-arm IPTp trial in Malawi (2003–2006) [94]. Pregnant women received standard IPTp-SP, or monthly IPTp-SP or monthly IPTp-SP plus 1 g azithromycin during two antenatal visits; the prevalence of *T. vaginalis* at delivery was 16.7% (n = 411;

Table 8. Treatment efficacy studies of 1 g azithromycin for the treatment of *Chlamydia trachomatis* in pregnant women.

Study (year)	Country	Year(s)	Number cured of evaluated	Percent cured	95% CI	Diagnostic method	Birth outcomes	Additional details	Ref.
Kacmar <i>et al.</i> (2001)	USA	1998–2000	18/19	94.7%	84.4–105.1%	Ligase chain reaction; TOC 28–42 days	NR	52.6% (n = 19; 95% CI: 31.5–72.8%) had side effects; passive or active solicitation not reported; 13.6 weeks (± 8.0 SD) mean gestational age at enrollment	[73]
Jacobson <i>et al.</i> (2001)	USA	1998–2000	35/55	63.6%	47.7–79.6%	DNA LCx STD assay; TOC 28 days	13.3% (6/45) preterm	10.1% (n = 55; 95% CI: 5.2–21.9%) had side effects; passive or active solicitation not reported; 20.6 weeks (± 8.8 SD) mean gestational age at enrollment	[74]
Wehbeh <i>et al.</i> (1998)	USA	NR	26/27	96.3%	89.0–103.6%	Culture	NR	100% (n = 27) of partners treated	[75]
Adair <i>et al.</i> (1998)	USA	1995–1997	37/42	88.1%	77.7–98.5%	DNA assay; TOC 28 days	NR	11.9% (n = 5; 95% CI: 5.3–25.1%) had side effects; passive or active solicitation not reported; 21.6 weeks (± 9.5 SD) mean gestational age at enrollment; 54.8% (n = 23) of partners treated	[76]
Edwards <i>et al.</i> (1996)	USA	1993–1994	61/65	93.8%	87.8–99.9%	DNA assay; TOC 14 days	9.2% (6/65) preterm, 3 due to PROM; mean birth age 38.8 weeks ± 1.6	20.4 weeks mean gestational age at enrollment	[78]
Rosenn <i>et al.</i> (1995)	USA	1994–1995	21/23	91.3%	79.3–103.4%	PCR (Amplicor); TOC 21 days	NR	19.3 weeks (± 3.5 SD) mean gestational age at enrollment	[79]
Gunter <i>et al.</i> (1996)	USA	NR	22/22	100%	NA	DNA assay; TOC 14 days	NR	13.6% (n = 3; 95% CI: 5.0–33.6%) had gastrointestinal side effects; passive or active solicitation not reported	[77]
	USA	NR	15/15	100%	NA		NR		[80]

NR: Not reported; TOC: Test of cure.

Table 8. Treatment efficacy studies of 1 g azithromycin for the treatment of *Chlamydia trachomatis* in pregnant women (cont.).

Study (year)	Country	Year(s)	Number cured of evaluated	Percent cured	95% CI	Diagnostic method	Birth outcomes	Additional details	Ref.
Bush & Rosa (1994)						DNA assay; TOC 14 days		0% of women experienced side effects; 100% (n = 15) of partners treated	
Rahangdale et al. (2006)	USA	1999–2000	137/141	97.2%	94.4–99.9%	DNA assay; TOC days 7–20; 21–34; 35–55; >56	7.5% (16/221 'any' preterm azithromycin)	13.1% (n = 191; 95% CI: 9.0–18.6%) also had BV 14.1 weeks (± 6.3 SD) mean gestational age at enrollment	[185]
Miller et al. (1995)	USA	1993–1994	132/138	95.6%	92.2–99.1%	DNA assay; TOC 10–14 days	15.2% (19/125) preterm	5.5% (n = 146; 95% CI: 2.8–10.4%) had side effects; 23.9% (n = 138; 17.6–31.7%) women also had NG; 17.4% (4/17) reported side effects (all GI); mean gestational age at enrollment not reported	[186]

NR: Not reported; TOC: Test of cure.

95% CI: 13.5–20.7%), 15.1% (n = 411; 95% CI: 12.0–18.9%) and 11.0% (n = 419; 95% CI: 8.3–14.3%), respectively. Thus, women who received azithromycin had 35% (RR: 0.65; 95% CI: 0.46–0.93; p = 0.02) fewer *T. vaginalis* infections at delivery compared with monthly recipients of IPTp-SP.

A cluster randomized trial in Uganda (1994–1998) compared the incidence of HIV infections among nonpregnant adults who received 1 g azithromycin, 250 mg ciprofloxacin and 2 g metronidazole versus multivitamins plus antihelmintics [95]. Although the trial was terminated early for lack of protection against the primary endpoint, the incidence of several curable STIs/RTIs was lower in the control group, most notably *T. vaginalis*. The cumulative incidence of newly diagnosed *T. vaginalis* infection was 4.8/100 person-years (116/2,397 person-years) in the intervention group compared with 9.1/100 person-years (182/1,993 person-years) in the control group (RR: 0.52; 95% CI: 0.35–0.79).

The same combination of antimicrobials was provided to female sex workers in rural Zimbabwe as a one-time treatment followed by 3 monthly check-ups [96]. The prevalence of *T. vaginalis* was just under 20% at baseline, decreased to approximately 5% at visit 2, rose to nearly 13% at visit 3 and lowered again to just over 10%, that is, one-half of the pretreatment levels.

Bacterial vaginosis

The WHO recommends treating bacterial vaginosis in pregnant women, preferably after the first trimester, with 200 or 250 mg metronidazole three-times per day for 7 days, or 5 g metronidazole gel (0.75%) applied intravaginally twice a day for 5 days or 300 mg clindamycin 300 mg orally twice a day for 7 days [36]. As with *T. vaginalis*, if treatment is imperative during the first trimester of pregnancy, 2 g metronidazole orally is recommended [36]. Bacterial vaginosis has no single causative agent, but is thought to result from destabilization of *Lactobacillus* species (spp.) with secondary colonization of anaerobic organisms that include *Gardnerella vaginalis*, *Bacteroides* spp, *Mobiluncus* spp. and *Mycoplasma hominis* alongside an increase in vaginal pH [97,98].

In vivo evidence

Our review identified no trials that have attempted to measure the treatment efficacy of azithromycin alone against bacterial vaginosis. Only one study in the USA (2002–2005) investigated the use of azithromycin as a partner drug with metronidazole for the treatment of symptomatic bacterial vaginosis. Nonpregnant women received one of four treatments: 750 mg metronidazole once per day for 7 days, or metronidazole once per day for 7 days plus 1 g azithromycin on days 1 and 3, or metronidazole for 14 days or metronidazole for 14 days plus azithromycin on days 1 and 3 [99]. No additional benefit of cure was observed among women who received metronidazole plus azithromycin compared with metronidazole alone.

Antibiotic treatment for bacterial vaginosis is challenging, in part, because it is a syndrome that involves multiple

Table 9. Sensitivity testing of *Chlamydia trachomatis* isolates to azithromycin.

Study (year)	Country	Year(s)	Azithromycin			Other macrolides			Additional details	Ref.
			MIC (µg/ml)	MBC/MCC (µg/ml)	MIC (µg/ml)	MBC (µg/ml)				
<i>No resistance observed</i>										
Ljubin-Sternak <i>et al.</i> (2012)	Croatia	2010	0.064 to 0.125	0.064–2.0	Doxycycline: 0.016–0.064	0.032–1.0	0.032–1.0	24 urogenital strains assessed	[187]	
Donati <i>et al.</i> (2010)	Italy	2005–2006	0.25–0.5	0.5–1.0	Doxycycline: 0.03–0.06 Erythromycin: 0.5–1.0	0.06–0.125 1.0–2.0	0.06–0.125 1.0–2.0	All serovars had comparable susceptibilities. Azithromycin and doxycycline bactericidal with MBC at one to two-times the MIC. (50 strains)	[188]	
Hong <i>et al.</i> (2009)	Ethiopia	2006	0.25–0.5	0.25–1.0	Doxycycline: 0.015–0.03	0.03–0.06	0.03–0.06	Azithromycin unchanged between pre- and post-mass biannual treatment of trachoma (10 strains)	[189]	
Samra <i>et al.</i> (2001)	Israel	1997–1999	0.06–0.125	0.06–0.25	Doxycycline: 0.125–0.25 Tetracyclines: 0.25–0.5	0.125–4.0 0.25–4.0	0.125–4.0 0.25–4.0	Smallest MBC and MIC difference in azithromycin versus doxycycline (4 dilutions differences; 50 isolates)	[84]	
Lefèvre <i>et al.</i> (1993)	France	NR	0.06–0.125	0.25–0.5	Clarithromycin: 0.008 Erythromycin : 0.06–0.125 Tetracyclines: 0.125–0.25	0.03–0.125 0.25–2.0 1.0–4.0	0.03–0.125 0.25–2.0 1.0–4.0	15 clinical isolates tested	[190]	
Agacifidan <i>et al.</i> (1993)	United States	1993	≤0.06–1	0.12–2.0	Doxycycline: 0.015–0.06 Tetracyclines: 0.03–0.12	0.015–0.06 0.06–0.12	0.015–0.06 0.06–0.12	Azithromycin highly active against CT in isolates from urethral and cervical samples (azithromycin 10 strains, doxycycline 22 strains, tetracycline 22 strains)	[191]	
Scieux <i>et al.</i> (1990)	UK	1990	0.064–0.25	2.0–8.0	Doxycycline: 0.016–0.064 Erythromycin: 0.064–0.128	0.5–8.0 0–64.0	0.5–8.0 0–64.0	10 strains used from the USA	[192]	
<i>Resistance observed</i>										
Bhengraj <i>et al.</i> (2010)	India	2006–2007	0.12–8	≤8.0	Doxycycline: 0.025–8.0	≤8.0	≤8.0	Decreased antimicrobial susceptibility in recurrently infected female patients (21 isolates)	[92]	
Misyurina <i>et al.</i> (2004)	Russia	2000–2002	>5.12	>5.12	Erythromycin: >5.12	>5.12	>5.12	Isolates from salpingitis, endocervicitis, and urethritis showed resistance (6 isolates)	[90]	
Somani <i>et al.</i> (2000)	USA	1995–1998	0.5 to >4.0	>4.0	Doxycycline: 0.125–>4.0	>4.0	>4.0	Resistance of strains causing relapsing or persistent infection in 3 patients (3 isolates)	[193]	
Rice <i>et al.</i> (1995)	USA	1995	0.125–2.0	0.5 to >4.0	Doxycycline: 0.008–0.06	0.015–4.0	0.015–4.0	Isolates susceptible to azithromycin and doxycycline in asymptotic infection	[91]	

Unspecified low-risk populations.
 CT: *Chlamydia trachomatis*; MIC: Minimum bactericidal concentration; MCC: Minimum chlamydical concentration; MBC: Minimum inhibitory concentrations; NR: Not reported; PID: Pelvic inflammatory disease.

Table 10. Trials using azithromycin alone and in combination with other drugs reporting protection curable STIs/RTIs among pregnant women.

Study (year)	Country	Year(s)	Regimen	Number of cases post-treatment (%)					Additional details	Ref.
				Treponema pallidum†	Neisseria gonorrhoeae	Chlamydia trachomatis	Trichomonas vaginalis	Bacterial vaginosis		
Luntamo et al. (2010)	Malawi	2003–2006	Intervention 1 g AZ two-times + SP monthly	NR	0.5% (2/391)	0.3% (1/391)	11.0% (46/419)	NR	Significant protection against <i>T. vaginalis</i> p = 0.05	[94]
			Intervention SP monthly	NR	2.1% (8/384)	0.3% (1/384)	15.1% (62/411)	NR		
			Control SP two-times	NR	0.7% (3/391)	0.3% (1/391)	16.7% (69/411)	NR		
van den Broek et al. (2009)	Malawi	2004–2005	Intervention 1 g AZ two-times + SP two-times	NR	NR	NR	NR	NR	No difference in preterm birth (16.8% versus 17.4%); potential explanatory factors include use of sub-optimal syphilis treatment [196,197]	[194]
			Control SP two-times	NR	NR	NR	NR	NR		
Gray et al. (2001)	Uganda	1994–1998	Intervention 1 g AZ + 250 mg CIPX + 2 g MTZ	3.4% (57/1,677)	0.9% (14/1,503)	1.1% (16/1,503)	4.7% (41,779)	36.3% (645/1,779)	Neonatal death RR: 0.83; 95% CI: 0.71–0.97; Low birth weight RR: 0.68; 95% CI: 0.55–0.86; Preterm delivery RR: 0.77; 95% CI: 0.56 to 1.05. Vertical transmission of HIV was no different between intervention and control groups	[104]
			Control Iron-folate + 100 mg MBZ two-times	3.3% (46/1,376)	1.7% (24/1,394)	2.7% (38/1,394)	15.9% (248/1,569)	48.5% (764/1,576)		
Wawer et al. (1999)	Uganda	1994–1998	Intervention 1 g AZ + 250 mg CIPX + 2 g MTZ	6.0% (80/1,323)	1.0% (8/770)	1.2% (9/770)	5.3% (72/1,350)	39.1% (533/1,364)	Vertical transmission of HIV was no different between intervention and control groups	[95]
Control Iron-folate + 100 mg MBZ two-times	7.1% (75/1,056)	2.1% (15/714)	3.5% (25/714)	17.4% (198/1,137)	52.8% (609/1,154)					

Low-risk populations

†2,4 mu benzathine Penicillin G was administered to pregnant women in all treatment groups per national guidelines (exception being [194].
AZ: Azithromycin; CIPX: Ciprofloxacin; NR: Not reported; RR: Risk ratio; SP: Sulfadoxine-pyrimethamine.

Table 11. Trials using azithromycin alone and in combination with other antimicrobial therapies not contraindicated in pregnancy and reporting protection curable STIs/RTIs among commercial sex workers.

Study (year)	Country	Year(s)	Regimen	Proportion of cases cured (cases pre-treatment/cases post-treatment)				Additional details	Ref.
				Treponema pallidum [†]	Neisseria gonorrhoeae	Chlamydia trachomatis	Trichomonas vaginalis		
Kaul <i>et al.</i> (2004)	Kenya	1998–2002	Intervention 1 g AZ monthly (multi-yr pd)	3.9%	2.6%	1.1%	11.3%	53.0%	Incidence reported per 100 women-years [93]
			Control Placebo	3.8%	5.7%	6.5%	20.4%	57.4%	
Labbe <i>et al.</i> (2012)	Benin Ghana	2001	Intervention 1 g AZ or 500 g CIPX monthly for 9 months alternating AZ & CIPX	NR	5.6% (7/126)	1.6% (2/126)	NR	NR	Significant protection against <i>N. gonorrhoeae</i> p = 0.05 [197]
Cowan <i>et al.</i> (2005)	Zimbabwe	NR	Control Placebo	NR	12.5% (14/112)	2.7% (3/112)	NR	NR	[96]
			Intervention 1 g AZ+2 g MTZ+500 mg CIPX	Base: 5.0% (2.8–8.7%); V2, V3, V4: NR in any form	Base: 1.9% (0.5–3.4%); V2, V3, V4: visible inaccuracies in graphs; cannot estimate	Base: 1.7% (0.3–3.0%); V2, V3, V4: visible inaccuracies in graphs; cannot estimate	Base: 19.3% (15.2–23.4%); V2: 4.3% (7.7–2.2%); V3: 12.6% (17.7–8.8%); V4: 11.5% (15.7–7.8%)		
Wi <i>et al.</i> (2006)	Philippines	2001	Intervention 1 g AZ one time	NR	18.3% (207/1,130) 11.9% (82/687)	28.6% (323/1,130) 15.1% (104/687)	NR	NR	[198]
			Control Placebo	Prior to intervention 1 month post-intervention	NR	18.3% (207/1,130) 11.9% (82/687)	28.6% (323/1,130) 15.1% (104/687)	NR	
Williams <i>et al.</i> (2003)	South Africa	1998–2000	Intervention 1 g AZ nine-times in 9 months	9.8% (68/691) 18.7% (166/893)	6.9% (48/691) 8.6% (77/893)	7.9% (55/691) 13.8% (123/893)	NR	NR	HIV prevalence among CSW in the mining community was 68.6% [199]
Steen <i>et al.</i> (2000)	South Africa	1996–1997	Intervention 1 g AZ every month for 9 months	NR	17.3% (70/407) 4.7% (5/108)	14.3% (58/407) 0.9% (1/108)	NR	NR	Pre-intervention NG and/or CT = 24.9% (101/407); Post-intervention NG and/or CT = 5.7% (6/108) [200]

Italicized values are approximate based on enlarged graphs published in Cowan *et al.* and percentages in parentheses reflect the 95% confidence intervals [96].
[†]2, 4 mg benzathine Penicillin G was administered to commercial sex workers who tested positive for syphilis in all treatment groups per national guidelines.
 AZ: Azithromycin; NR: Not reported; CIPX: Ciprofloxacin; CT: C. trachomatis; CSW: Commercial sex worker; MTZ: Metronidazole; NG: Neisseria gonorrhoeae.

Table 12. Sensitivity of macrolides and structurally related agents against key causative organisms in bacterial vaginosis

Study (year)	Country	Year(s)	Minimum inhibitory concentrations of specific macrolides (µg/ml)							Ref.
			Azithromycin	Erythromycin	Clarithromycin	Roxithromycin	Clindamycin [†]	Telithromycin [†]		
<i>Gardnerella vaginalis</i>										
Jones <i>et al.</i> (1998)	UK	NR	<0.03–0.125	<0.03–0.06	NR	NR	NR	NR	NR	[201]
Shanker <i>et al.</i> (1982)	Australia	NR	NR	0.007–0.06	NR	NR	0.007–0.06	NR	NR	[202]
King [†] <i>et al.</i> (1987)		NR	NR	0.008–0.016	NR	0.016	NR	NR	NR	[203]
<i>Bacteroides</i> species										
Jones <i>et al.</i> (1998)	UK	NR	0.06–16	<0.03–32	NR	NR	NR	NR	NR	[201]
Dubreuil <i>et al.</i> (1987)	England, France, Germany, Japan	NR	NR	0.003–>64	NR	0.003–>64	NR	NR	NR	[204]
Maskell <i>et al.</i> (1990)	UK	NR	0.5–>16	<0.25–16	NR	NR	NR	NR	NR	[205]
Chang <i>et al.</i> (1995)	Taiwan	1989–1992	1–>256	0.25–>256	≤0.03–>256	0.25–>256	NR	NR	NR	[206]
Ednie <i>et al.</i> (1997)	USA	NR	1–>64	0.5–>64	0.5–>64	2–>64	≤0.06–>64	NR	NR	[207]
Mikamo <i>et al.</i> (2003)	Japan	2000	0.125–32	0.125–32	0.063 to 16	NR	NR	0.032–16	NR	[208]
Marina <i>et al.</i> (2009)	Bulgaria	1983–2007	NR	0.5–>64	NR	NR	0.125–32	NR	NR	[209]
Chen <i>et al.</i> (1992)	Australia	1986–1991	0.5 to 128	0.25–128	NR	NR	NR	NR	NR	[210]
Wexler <i>et al.</i> (2001)	USA	NR	NR	NR	NR	NR	NR	NR	0.25–>64	[211]
<i>Mycoplasma hominis</i>										
Ridgway <i>et al.</i> (1987)	UK	NR	NR	>32	NR	NR	8 to 16	NR	NR	[203]
<i>Mobinculus</i> species										
Spiegel <i>et al.</i> (1987)	USA	NR	NR	≤0.2–>200	NR	NR	NR	≤0.015–4	NR	[212]

Unspecified low-risk population.

[†]Not a macrolide but has similar mechanism of action and included for comparability.
NR: Not reported.

microorganisms rather than a single etiological agent. Comparable data from other macrolides suggest potential therapeutic value for azithromycin against bacterial vaginosis (TABLE 12). Analysis of azithromycin against the anaerobic and carboxyphilic bacteria that replace the normal vaginal flora may provide a better understanding as to the potential role of azithromycin against bacterial vaginosis.

Discussion

Azithromycin has been used against curable STIs/RTIs for 25 years. It has been an attractive option for preventive and curative treatment because it is efficacious as a single dose and offers reasonable tolerability against *T. pallidum*, *N. gonorrhoeae* and *C. trachomatis*. During the 1990s, and prior to the advent of antiretroviral therapies for HIV, the management of curable STIs/RTIs received renewed importance, particularly as trials showed that treatment of *N. gonorrhoeae*, *C. trachomatis* and *T. vaginalis* reduced the genital viral load of HIV among men and women [100–103]. Groups at high risk for transmitting HIV have since been targeted by treatment campaigns using 1 g azithromycin. Thus, it is not a surprise that changes in azithromycin sensitivity within high-income settings have often been observed first among members of high-risk groups. Pregnant women attending ANC facilities in sub-Saharan Africa do not have the same risk profile. Thus, on this basis alone, it is less likely that the use of ABC therapies in IPTp would be a catalyst for the rapid emergence of azithromycin resistance, although its emergence cannot be ruled out. The potential benefits of ABC therapies may be best viewed through prior experience with mass drug administration among pregnant women. In the context of the AIDS epidemic and before the age of antiretroviral therapies, researchers attempted to prevent maternal-to-child transmission (MTCT) of HIV by providing pregnant women in Uganda 1 g azithromycin, in combination with 250 mg ciprofloxacin and 2 g metronidazole [104]. The data safety monitoring board suspended the trial early for reasons of futility, despite having cut neonatal deaths by 17% (RR: 0.83; 95% CI: 0.71–0.97), decreased the incidence of low birth-weight by 32% (RR: 0.68; 95% CI: 0.53–0.86), and reduced the incidence of preterm delivery by 23% (RR: 0.77; 95% CI: 0.56–1.05). These impressive results were achieved at a time when neither IPTp-SP nor insecticide treated bed nets for the control of malaria in pregnancy had been deployed.

If ABC therapies are used in IPTp, then there are several key factors to consider that are pathogen specific. Regarding syphilis, 1 g azithromycin should be used alongside 2.4 mu BPG for three reasons: combination therapy has been shown to achieve higher rates of cure than either therapy alone [105]; use of ABC therapy with BPG would likely reduce selection of the A→G mutation associated with azithromycin and preserve *T. pallidum* sensitivity; and only BPG can be expected to cure congenital infection if the placenta has been invaded by spirochetes [106]. As for *N. gonorrhoeae*, 1 g azithromycin may be just above the MIC of fully susceptible strains. Thus, ABC therapies containing >1 g azithromycin may be preferable from

the standpoint of reducing selection pressure. However, a single 2 g dose may not be well tolerated as 6 in 10 patients reported self-limiting gastrointestinal discomfort when treated for syphilis infection with such a regimen [42]. The dose could be split over 2 days to improve tolerability. ABC therapies that contain 2 g azithromycin, either a single- or split-dose, would be protective against *C. trachomatis*. Although the data are limited and the mechanism of action is not understood, ABC therapies that have 1 g azithromycin may protect against *T. vaginalis* based on reports from Malawi among pregnant women [107] and commercial sex workers in Kenya [93]. It is curious, however, that *T. vaginalis* infection during pregnancy is associated with adverse birth outcomes, but the first-line treatment of 2 g metronidazole does not always improve birth outcomes. A trial in Uganda reported that pregnant women treated for *T. vaginalis* infection were 2.5-times more likely to deliver a low birth-weight infant than untreated women (RR: 2.49; 95% CI: 1.12–5.50) [108]. The authors suggest that this may be attributable to metronidazole exposure. Another trial in the USA reported an increase in the risk of preterm delivery among pregnant women exposed to metronidazole for the treatment of asymptomatic trichomoniasis compared with those who were not treated (RR: 1.8; 95% CI: 1.2–2.7) [109]. In contrast to these findings from high-income settings, data from a multicenter trial in sub-Saharan Africa indicate that treatment of *T. vaginalis* infection using metronidazole does not increase the chances of preterm birth [110]. Apart from bacterial vaginosis, which is not transmitted through sexual contact, re-infection will remain a risk for pregnant women and, therefore, providers should continue to offer education and screening as appropriate.

None of the studies identified in this review indicate that azithromycin offers preventive or curative effect against bacterial vaginosis, the most prevalent of curable STIs/RTIs. Antibiotic therapy has only been shown to reduce the risk of preterm delivery by one-half (RR: 0.53; 95% CI: 0.34–0.84) among pregnant women with bacterial vaginosis (Nugent scores 7–10) or intermediate flora (Nugent scores 4–6) [111]. A Nugent score of 0–3 is considered normal [112] for which no protection against adverse birth outcomes has been observed.

Conclusions

ABC therapies are among leading candidates to replace SP for use in IPTp and may offer important public health benefits by also reducing the burden of curable STIs/RTIs in pregnancy. Evidence from nonpregnant adults suggests that ABC therapies containing 1 g azithromycin may cure maternal *T. pallidum* infection. BPG should still be administered with azithromycin because the combination has been shown to be more efficacious in nonpregnant adults than either treatment alone. Moreover, evidence from pregnant women indicates that eradication of congenital syphilis may require BPG treatment. *Neisseria gonorrhoeae* infection among pregnant women in sub-Saharan Africa, where strains are likely to be fully sensitive to azithromycin, is likely to be cured by ABC therapies containing 1 g azithromycin. However, 2 g may be needed to reduce persistent and/or

recurrent infection, and opportunities for the emergence of drug resistance. ABC therapy containing 1 g azithromycin would be curative of *C. trachomatis* infection, whereas some protection against *T. vaginalis* infection could be expected with the same dose. It remains unknown whether ABC therapies could offer protection against bacterial vaginosis if administered during the first half of pregnancy. ABC therapies merit investigation for the use in IPTp given their potential to reduce the dual burden of malaria and curable STIs/RTIs in pregnancy and improve maternal, fetal and neonatal health.

Expert commentary

Current strategies for addressing the dual-burden of malaria and curable STIs/RTIs in pregnancy are suboptimal. In West Africa, IPTp-SP continues to provide protection against the effects of malaria infection but, as previously noted, malaria parasites in East Africa have developed resistance so that IPTp-SP no longer protects against the malaria attributable fraction of low birthweight [3]. Some evidence suggests that IPTp-SP may even be harmful in areas where parasites express the 581G *dhps* mutation [4–6]. ABC therapies are likely to be more efficacious against malaria parasites in these settings. However, there is an urgent need for trials of ABC therapies to be conducted by independent researchers for policymakers to review alongside the results of trials produced and reported by industry.

In the case of curable STIs/RTIs, the focused ANC package recommended by the WHO includes screening for syphilis and the provision of BPG to women who test positive [50]. Screening would need to continue even if ABC therapies were used in IPTp. The WHO currently recommends the use of rapid point of care tests for syphilis in the ANC setting [113]. Using such tests will expedite case finding and treatment with BPG because results are available during the same consultation. As for the four other curable STIs/RTIs of this review, health care providers are limited to the use of a syndrome-based management algorithm to diagnose and to treat suspected infections. However, 80% of gonococcal and 70–75% chlamydial infections in women are asymptomatic [114] and, therefore, rarely recognized using the syndromic approach. As a result, the diagnostic algorithm has a low sensitivity (30–80%) and specificity (40–80%) for *N. gonorrhoeae* and *C. trachomatis* among pregnant women [115–117]. The sensitivity for *T. vaginalis* (54–83%) and bacterial vaginosis (51–69%) is slightly higher, with moderate specificity for *T. vaginalis* (40–54%) and bacterial vaginosis (40–58%) [118]. Given the evidence of this review, ABC therapies used in IPTp could be expected to mitigate a considerable proportion of this unattended burden of curable STIs/RTIs.

Much is debated about the utility of a syndrome-based approach to diagnosing and treating many STIs/RTIs. Its shortcomings, as described above, illuminate a much-needed area for research. Specifically, more refined definitions of diagnosis need to be used when characterizing adverse birth outcomes. This is particularly so with *T. vaginalis* for which successful treatment does not necessarily reduce the risk of adverse birth outcomes. Similarly with bacterial vaginosis, treatment of women who have

Nugent scores of 1–3 has not reduced the incidence of preterm birth. With both of these infections, is the recommended regimen of metronidazole inadequate for radical cure? Is it administered too late in pregnancy to alter the course of events? Or are asymptomatic infections simply much less virulent and, therefore, treatment has a marginal effect on selected downstream measures of the adverse birth outcome? Studies of descriptive epidemiology are needed to understand better the extent to which symptomatic versus asymptomatic curable STIs/RTIs contribute to adverse birth outcomes. Such descriptive epidemiology, however, would be incomplete if the prevalence of co-infections were not also considered. The apparent failure to reduce the incidence of adverse birth outcomes following treatment for one infection may be masked by the presence of co-infection(s) that will only be mitigated with the use of combination therapies and consideration of downstream outcomes. The trial in Uganda that failed to reduce the incidence of MTCT of HIV is illustrative. HIV transmission was not interrupted for providing combination treatment against curable STIs/RTIs, but significant reductions were observed in the incidence of neonatal deaths by 17% (RR: 0.83; 95% CI: 0.71–0.97) and low birthweight by 32% (RR: 0.68; 95% CI: 0.53–0.86) [104].

Five-year view

Discussion of the future of IPTp needs to be placed in the context of broader malaria elimination efforts. IPTp-SP has long been considered a malaria control intervention that can be expected to protect less against the fraction of low birthweight attributable to malaria infection as malaria transmission decreases. Recent evidence suggests that IPTp-SP continues to protect against low birthweight among multigravidae in areas where the prevalence of malaria parasitemia measured in children is between 7 and 8%, whereas protection is conferred by IPTp-SP among paucigravidae until very low levels of transmission [119]. Unpublished results from a recently completed multicenter trial in West Africa, where there remain malaria parasites sensitive to SP, indicate that an approach of intermittent screening and treatment (IST) of malaria in pregnancy is noninferior to IPTp-SP (Manuscript under review/personal communication with D. Chandramohan). Thus, there is an urgent need for clinical trials in an area of high SP resistance in East Africa, designed to compare ABC therapies versus IST versus IPTp-SP. ABC therapies, given their action against malaria and curable STIs/RTIs, would be superior to IST and IST would be superior to IPTp-SP, potentially paving the way for adoption of an integrated malaria and curable STI/RTI control package that employs the use of combination treatment.

Financial & competing interests disclosure

RM Chico receives funding from Medicines for Malaria Venture, a non-profit foundation based in Geneva, Switzerland. E Ngulube is a Commonwealth Scholar. The authors have no other relevant affiliations or financial involvement with any organization or entity with a financial interest in or financial conflict with the subject matter or materials discussed in the manuscript apart from those disclosed.

No writing assistance was utilized in the production of this manuscript.

Key issues

- Use of azithromycin-based combination (ABC) therapies may have a transformative effect on maternal, fetal and newborn health by mitigating the dual-burden of malaria and curable sexually transmitted infections and reproductive tract infections sexually transmitted infections and reproductive tract infections (STIs/RTIs) in pregnancy.
- ABC therapies containing two or more grams of azithromycin may be less likely to select for resistance when exposed to *Treponema pallidum*, *Neisseria gonorrhoeae* and potentially, *Chlamydia trachomatis*.
- In the absence of evidence that azithromycin is curative of congenital syphilis, not simply maternal infection, benzathine penicillin G (BPG) will still need to be administered to pregnant women who have a syphilis infection; however, the combination of azithromycin plus BPG is more efficacious than BPG alone.
- ABC therapies may be preventive of *Treponema vaginalis* infection during pregnancy, although its impact on birth outcomes needs to be investigated.
- The most prevalent of all STIs/RTIs, bacterial vaginosis, may or may not be mitigated by the use of ABC therapies.
- Studies of descriptive epidemiology are needed to understand better the extent to which symptomatic versus asymptomatic curable STIs/RTIs contribute to adverse birth outcomes. There is an urgent need for clinical trials in an area of high sulfadoxine-pyrimethamine resistance in East Africa, designed to compare ABC therapies versus IPTp-SP versus providing IST for malaria in pregnancy during ANC visits.

References

- 1 WHO. WHO Policy Recommendation: Intermittent Preventive Treatment of Malaria in Pregnancy using Sulfadoxine-Pyrimethamine (IPTp-SP). 11 April 2013 (2013).
- 2 World Health Organization. *World malaria report: 2012*. World Health Organization, Geneva (2012).
- 3 Chico RM and Chandramohan D. Intermittent preventive treatment of malaria in pregnancy: at the crossroads of public health policy. *Trop. Med. Int. Health* 16(7), 774–785 (2011).
- 4 Harrington WE, Mutabingwa TK, Muehlenbachs A *et al.* Competitive facilitation of drug-resistant *Plasmodium falciparum* malaria parasites in pregnant women who receive preventive treatment. *Proc. Natl Acad. Sci. USA* 106(22), 9027–9032 (2009).
- 5 Harrington WE, Mutabingwa TK, Kabemela E, Fried M and Duffy PE. Intermittent treatment to prevent pregnancy malaria does not confer benefit in an area of widespread drug resistance. *Clin. Infect. Dis.* 53(3), 224–230 (2011).
- 6 Harrington WE, Morrison R, Fried M and Duffy PE. Intermittent preventive treatment in pregnant women is associated with increased risk of severe malaria in their offspring. *PLoS ONE*, 8(2), e56183 (2013).
- 7 Whitman MS and Tunkel AR. Azithromycin and clarithromycin: overview and comparison with erythromycin. *Infect. Control Hosp. Epidemiol.* 13(6), 357–368 (1992).
- 8 Chico RM, Pittrof R, Greenwood B and Chandramohan D. Azithromycin-chloroquine and the intermittent preventive treatment of malaria in pregnancy. *Malar. J.* 7(1), 255 (2008).
- 9 Sarkar M, Woodland C, Koren G and Einarson AR. Pregnancy outcome following gestational exposure to azithromycin. *BMC Pregnancy Childbirth* 6, 18 (2006).
- 10 Kuschner R, Heppner D, Andersen S *et al.* Azithromycin prophylaxis against a chloroquine resistant strain of *Plasmodium falciparum*. *Lancet* 343(8910), 1396–1397 (1994).
- 11 Anderson S, Berman J, Kuschner R *et al.* Prophylaxis of *Plasmodium falciparum* malaria with azithromycin administered to volunteers. *Ann. Intern. Med.* 123, 771–773 (1995).
- 12 Ohrt C, Willingmyre G, Lee P, Knirsch C and Milhous W. Assessment of azithromycin in combination with other antimalarial drugs against *Plasmodium falciparum* in vitro. *Antimicrob. Agents Chemother.* 46, 2518–2524 (2002).
- 13 Chico R M, Mayaud P, Ariti C, Mabey D, Ronsmans C, Chandramohan D. Prevalence of malaria and sexually transmitted and reproductive tract infections in pregnancy in sub-saharan Africa. *JAMA* 307(19), 2079–2086 (2012).
- 14 Williams JW. *A textbook of obstetrics*. D Appleton & Co, New York, USA 1923.
- 15 Ratnam AV, Din SN, Hira SK *et al.* Syphilis in pregnant women in Zambia. *Br. J. Vener Dis.* 58(6), 355–358 (1982).
- 16 Hay PE, Lamont RF, Taylor-Robinson D, Morgan DJ, Ison C and Pearson J. Abnormal bacterial colonisation of the genital tract and subsequent preterm delivery and late miscarriage. *BMJ* 308(6924), 295–298 (1994).
- 17 Leitich H, Bodner-Adler B, Brunbauer M, Kaider A, Egarter C, Husslein P. Bacterial vaginosis as a risk factor for preterm delivery: a meta-analysis. *Am. J. Obstet. Gynecol.* 189(1), 139–147 (2003).
- 18 Wilkowska-Trojnieł M, Zdrodowska-Stefanow B, Ostaszewska-Puchalska I, Redzko S, Przepiesc J, Zdrodowski M. The influence of *Chlamydia trachomatis* infection on spontaneous abortions. *Adv. Med. Sci.* 54(1), 86–90 (2009).
- 19 McDermott J, Steketee R, Larsen S and Wirima J. Syphilis-associated perinatal and infant mortality in rural Malawi. *Bull. World Health Organ.* 71(6), 773–780 (1993).
- 20 Temmerman M, Gichangi P, Fonck K *et al.* Effect of a syphilis control programme on pregnancy outcome in Nairobi, Kenya. *Sex. Transm. Infect.* 76(2), 117–121 (2000).
- 21 Watson-Jones D CJ, Balthazar G, Weiss H *et al.* Syphilis in pregnancy in Tanzania. I. Impact of maternal syphilis on outcome of pregnancy. *J. Infect. Dis.* 186, 940–947 (2002).
- 22 Association of *Chlamydia trachomatis* and *Mycoplasma hominis* with intrauterine growth retardation and preterm delivery. The John Hopkins Study of Cervicitis and Adverse Pregnancy Outcome. *Am. J. Epidemiol.* 129(6), 1247–1257 (1989).
- 23 Watson-Jones D, Changalucha J, Gumodoka B *et al.* Syphilis in pregnancy in Tanzania. I. Impact of maternal syphilis on

- outcome of pregnancy. *J. Infect. Dis.* 186(7), 940–947 (2002).
- 24 Gravett MG, Nelson HP, DeRouen T, Critchlow C, Eschenbach DA and Holmes KK. Independent associations of bacterial vaginosis and Chlamydia trachomatis infection with adverse pregnancy outcome. *JAMA* 256(14), 1899–1903 (1986).
- 25 McGregor JA, French JI, Parker R *et al.* Prevention of premature birth by screening and treatment of common genital tract infections: results of a prospective controlled evaluation. *Am. J. Obstet. Gynecol.* 173(1), 157–167 (1995).
- 26 Blas MM, Canchihuaman FA, Alva IE, Hawes SE. Pregnancy outcomes in women infected with Chlamydia trachomatis: A population-based cohort study in Washington State. *Sex. Transm. Infect.* 83(4), 314–318 (2007).
- 27 Elliott B, Brunham RC, Laga M *et al.* Maternal gonococcal infection as a preventable risk factor for low birth weight. *J. Infect. Dis.* 161(3), 531–536 (1990).
- 28 Cotch MF, Pastorek JG 2nd, Nugent RP *et al.* Trichomonas vaginalis associated with low birth weight and preterm delivery. The vaginal infections and prematurity study group. *Sex. Transm. Dis.* 24(6), 353–360 (1997).
- 29 Svare JA, Schmidt H, Hansen BB, Lose G. Bacterial vaginosis in a cohort of Danish pregnant women: prevalence and relationship with preterm delivery, low birthweight and perinatal infections. *BJOG* 113(12), 1419–1425 (2006).
- 30 Odendaal HJ, Schoeman J, Grove D *et al.* The association between Chlamydia trachomatis genital infection and spontaneous preterm labour. *S. Afr. J. Obstet. Gynaecol.* 12(3), 146–149 (2006).
- 31 Watson-Jones D, Weiss HA, Changalucha JM *et al.* Adverse birth outcomes in United Republic of Tanzania—impact and prevention of maternal risk factors. *Bull. World Health Organ.* 85(1), 9–18 (2007).
- 32 Rours GI, Duijts L, Moll HA *et al.* Chlamydia trachomatis infection during pregnancy associated with preterm delivery: a population-based prospective cohort study. *Eur. J. Epidemiol.* 26(6), 493–502 (2011).
- 33 Johnson HL, Ghanem KG, Zenilman JM, Erbdelding EJ. Sexually transmitted infections and adverse pregnancy outcomes among women attending inner city public sexually transmitted diseases clinics. *Sex. Transm. Dis.* 38(3), 167–171 (2011).
- 34 Donders GG, Desmyter J, De Wet DH, Van Assche FA. The association of gonorrhoea and syphilis with premature birth and low birthweight. *Genitourin. Med.* 69(2), 98–101 (1993).
- 35 Sutton MY, Sternberg M, Nsuami M, Behets F, Nelson AM, St Louis ME. Trichomoniasis in pregnant human immunodeficiency virus-infected and human immunodeficiency virus-uninfected congolese women: prevalence, risk factors, and association with low birth weight. *Am J Obstet Gynecol.* 181(3), 656–662 (1999).
- 36 Organization WH. *Guidelines for the Management of Sexually Transmitted Infections.* Switzerland (2003).
- 37 Hook EW 3rd, Stephens J, Ennis DM. Azithromycin compared with penicillin G benzathine for treatment of incubating syphilis. *Ann. Intern. Med.* 131(6), 434–437 (1999).
- 38 Klausner JD, Kohn RP, Kent CK. Azithromycin versus penicillin for early syphilis. *N. Engl. J. Med.* 354(2), 203–205, author reply 203–205 (2006).
- 39 Hook EW, 3rd, Martin DH, Stephens J, Smith BS and Smith K. A randomized, comparative pilot study of azithromycin versus benzathine penicillin G for treatment of early syphilis. *Sex. Transm. Dis.* 29(8), 486–490 (2002).
- 40 Kiddugavu MG, Kiwanuka N, Wawer MJ *et al.* Effectiveness of syphilis treatment using azithromycin and/or benzathine penicillin in Rakai, Uganda. *Sex. Transm. Dis.* 32(1), 1–6 (2005).
- 41 Riedner G, Rusizoka M, Todd J *et al.* Single-dose azithromycin versus penicillin G benzathine for the treatment of early syphilis. *N. Engl. J. Med.* 353(12), 1236–1244 (2005).
- 42 Hook EW, 3rd, Behets F, Van Damme K *et al.* A phase III equivalence trial of azithromycin versus benzathine penicillin for treatment of early syphilis. *J. Infect. Dis.* 201(11), 1729–1735 (2010).
- 43 Mitchell SJ, Engelman J, Kent CK, Lukehart SA, Godornes C, Klausner JD. Azithromycin-resistant syphilis infection: San Francisco, California, 2000–2004. *Clin. Infect. Dis.* 42(3), 337–345 (2006).
- 44 Lukehart SA, Godornes C, Molini BJ *et al.* Macrolide resistance in Treponema pallidum in the United States and Ireland. *N. Engl. J. Med.* 351(2), 154–158 (2004).
- 45 Marra CM, Colina AP, Godornes C *et al.* Antibiotic selection may contribute to increases in macrolide-resistant Treponema pallidum. *J. Infect. Dis.* 194(12), 1771–1773 (2006).
- 46 Grimes M, Sahi SK, Godornes BC *et al.* Two mutations associated with macrolide resistance in Treponema pallidum: increasing prevalence and correlation with molecular strain type in Seattle, Washington. *Sex. Transm. Dis.* 39(12), 954–958 (2012).
- 47 Chen XS, Yin YP, Wei WH *et al.* High prevalence of azithromycin resistance to Treponema pallidum in geographically different areas in China. *Clin. Microbiol. Infect.* 8(10), 1469–0691 (2012).
- 48 Wu H, Chang SY, Lee NY *et al.* Evaluation of macrolide resistance and enhanced molecular typing of Treponema pallidum in patients with syphilis in Taiwan: a prospective multicenter study. *J. Clin. Microbiol.* 50(7), 2299–2304 (2012).
- 49 Van Damme K, Behets F, Ravelomanana N *et al.* Evaluation of azithromycin resistance in Treponema pallidum specimens from Madagascar. *Sex. Transm. Dis.* 36(12), 775–776 (2009).
- 50 World Health Organization. *Sexually transmitted and other reproductive tract infections: Guide to essential practice.* Department of Reproductive Health and Research, World Health Organization, Geneva (2005).
- 51 Odugbemi T, Oyewole F, Isichei CS, Onwukeme KE and Adeyemi-Doro FA. Single oral dose of azithromycin for therapy of susceptible sexually transmitted diseases: a multicenter open evaluation. *West Afr. J. Med.* 12(3), 136–140 (1993).
- 52 Waugh MA. Open study of the safety and efficacy of a single oral dose of azithromycin for the treatment of uncomplicated gonorrhoea in men and women. *J. Antimicrob. Chemother.* 31(Suppl. E), 193–198 (1993).
- 53 Swanston WH, Prabhakar P, Barrow L, Mahabir BS, Furlonge C. Single dose (direct observed) azithromycin therapy for Neisseria gonorrhoeae and Chlamydia trachomatis in STD clinic attendees with genital discharge in Trinidad and Tobago. *West Indian Med. J.* 50(3), 198–202 (2001).
- 54 Lassus A. Comparative studies of azithromycin in skin and soft-tissue infections and sexually transmitted infections by Neisseria and Chlamydia species. *J. Antimicrob. Chemother.* 25(Suppl. A), 115–121 (1990).
- 55 Steingrimsson O, Olafsson JH, Thorarinnsson H, Ryan RW, Johnson RB and Tilton RC. Azithromycin in the

- treatment of sexually transmitted disease. *J. Antimicrob. Chemother.* 25(Suppl. A), 109–114 (1990).
- 56 Steingrimsson O, Olafsson JH, Thorarinnsson H, Ryan RW, Johnson RB and Tilton RC. Single dose azithromycin treatment of gonorrhoea and infections caused by *C. trachomatis* and *U. urealyticum* in men. *Sex. Transm. Dis.* 21(1), 43–46 (1994).
- 57 Gruber F, Grubisic-Greblo H, Jonjic A *et al.* Treatment of gonococcal and chlamydial urethritis with azithromycin or doxycycline. *Chron. Derm. (Roma)* 5, 213–218 (1995).
- 58 Gruber F, Brajac I, Jonjic A, Grubisic-Greblo H, Lenkovic M, Stasic A. Comparative trial of azithromycin and ciprofloxacin in the treatment of gonorrhoea. *J. Chemother.* 9(4), 263–266 (1997).
- 59 Rustomjee R, Kharsany AB, Connolly CA, Karim SS. A randomized controlled trial of azithromycin versus doxycycline/ciprofloxacin for the syndromic management of sexually transmitted infections in a resource-poor setting. *J. Antimicrob. Chemother.* 49(5), 875–878 (2002).
- 60 DerSimonian R, Laird N. Meta-analysis in clinical trials. *Control Clin. Trials* 7(3), 177–188 (1986).
- 61 Bignell C, Garley J. Azithromycin in the treatment of infection with *Neisseria gonorrhoeae*. *Sex. Transm. Infect.* 86(6), 422–426 (2010).
- 62 World Health Organization, London School of Hygiene and Tropical Medicine and International Trachoma Initiative. *Trachoma control: A guide for programme managers*. World Health Organization, Switzerland (2006).
- 63 Handsfield HH, Dalu ZA, Martin DH, Douglas JM Jr, McCarty JM, Schlossberg D. Multicenter trial of single-dose azithromycin vs. ceftriaxone in the treatment of uncomplicated gonorrhoea. Azithromycin Gonorrhoea Study Group. *Sex. Transm. Dis.* 21(2), 107–111 (1994).
- 64 Khaki P, Bhalla P, Sharma A, Kumar V. Correlation between In vitro susceptibility and treatment outcome with azithromycin in gonorrhoea: a prospective study. *Indian J. Med. Microbiol.* 25(4), 354–357 (2007).
- 65 Johnson SR, Sandul AL, Parekh M, Wang SA, Knapp JS, Trees DL. Mutations causing in vitro resistance to azithromycin in *Neisseria gonorrhoeae*. *Int. J. Antimicrob. Agents* 21(5), 414–419 (2003).
- 66 Martin I, Jayaraman G, Wong T, Liu G, Gilmour M, Canadian Public Health Laboratory N. Trends in antimicrobial resistance in *Neisseria gonorrhoeae* isolated in Canada: 2000–2009. *Sex. Transm. Dis.* 38(10), 892–898 (2011).
- 67 Martin I, Jayaraman G, Wong T, Liu G and Gilmour M. Trends in antimicrobial resistance in *Neisseria gonorrhoeae* isolated in Canada: 2000–2009. *Sex. Transm. Dis.* 38(10), 892–898. (2011).
- 68 Cole MJ, Chisholm SA, Hoffmann S *et al.* European surveillance of antimicrobial resistance in *Neisseria gonorrhoeae*. *Sex. Transm. Infect.* 86(6), 427–432 (2010).
- 69 Centers for Disease Control and Prevention. *Neisseria gonorrhoeae* with reduced susceptibility to azithromycin — San Diego County, California, 2009. *MMWR Morb. Mortal. Wkly Rep.* 60(18), 579–581 (2011).
- 70 Starnino S, Galarza P, Carvallo ME *et al.* Retrospective analysis of antimicrobial susceptibility trends (2000–2009) in *Neisseria gonorrhoeae* isolates from countries in Latin America and the Caribbean shows evolving resistance to ciprofloxacin, azithromycin and decreased susceptibility to ceftriaxone. *Sex. Transm. Dis.* 39(10), 813–821. (2012).
- 71 Khaki P, Bhalla P, Sharma P, Chawla R, Bhalla K. Epidemiological analysis of *Neisseria gonorrhoeae* isolates by antimicrobial susceptibility testing, auxotyping and serotyping. *Indian J. Med. Microbiol.* 25(3), 225–229 (2007).
- 72 Sethi S, Golparian D, Bala M *et al.* Antimicrobial susceptibility and genetic characteristics of *Neisseria gonorrhoeae* isolates from India, Pakistan and Bhutan in 2007–2011. *BMC Infect. Dis.* 13, 35 (2013).
- 73 Kacmar J, Cheh E, Montagno A and Peipert JF. A randomized trial of azithromycin versus amoxicillin for the treatment of Chlamydia trachomatis in pregnancy. *Infect. Dis. Obstet. Gynecol.* 9(4), 197–202 (2001).
- 74 Jacobson GF, Autry AM, Kirby RS, Liverman EM, Motley RU. A randomized controlled trial comparing amoxicillin and azithromycin for the treatment of Chlamydia trachomatis in pregnancy. *Am. J. Obstet. Gynecol.* 184(7), 1352–1354, discussion 1354–1356 (2001).
- 75 Wehbeh HA, Rugeirio RM, Shahem S, Lopez G, Ali Y. Single-dose azithromycin for Chlamydia in pregnant women. *J. Reprod. Med.* 43(6), 509–514 (1998).
- 76 Adair CD, Gunter M, Stovall TG, McElroy G, Veille JC, Ernest JM. Chlamydia in pregnancy: a randomized trial of azithromycin and erythromycin. *Obstet. Gynecol.* 91(2), 165–168 (1998).
- 77 Gunter ME AC, Ernest JM, McElroy G. Azithromycin powder versus erythromycin in the treatment of chlamydial cervicitis in pregnancy. *Infect. Dis. Obstet. Gynecol.* 4(53) (1996).
- 78 Edwards MS, Newman RB, Carter SG, Leboeuf FW, Menard MK, Rainwater KP. Randomized clinical trial of azithromycin vs. erythromycin for the treatment of chlamydia cervicitis in pregnancy. *Infect. Dis. Obstet. Gynecol.* 4(6), 333–337 (1996).
- 79 Rosenn MF, Macones GA and Silverman NS. Randomized trial of erythromycin and azithromycin for treatment of chlamydial infection in pregnancy. *Infect Dis Obstet Gynecol.* 3(6), 241–244 (1995).
- 80 Bush MR and Rosa C. Azithromycin and erythromycin in the treatment of cervical chlamydial infection during pregnancy. *Obstet. Gynecol.* 84(1), 61–63 (1994).
- 81 Golden MR, Whittington WL, Handsfield HH *et al.* Effect of expedited treatment of sex partners on recurrent or persistent gonorrhoea or chlamydial infection. *N. Engl. J. Med.* 352(7), 676–685 (2005).
- 82 Katz BP FD, Orr DP. Factors affecting chlamydial persistence or recurrence one and three months after treatment. Chlamydial infections. *Proceedings of the ninth international symposium on human chlamydial infection*. Stephens RS, Byrne GI, Christiansen G *et al.*(Eds). International Chlamydia Symposium, USA 35–38 (1998).
- 83 Horner P. The case for further treatment studies of uncomplicated genital Chlamydia trachomatis infection. *Sex. Transm. Infect.* 82(4), 340–343 (2006).
- 84 Samra Z, Rosenberg S, Soffer Y, Dan M. In vitro susceptibility of recent clinical isolates of Chlamydia trachomatis to macrolides and tetracyclines. *Diag. Microbiol. Infect. Dis.* 39(3), 177–179 (2001).
- 85 Somani J, Bhullar VB, Workowski KA, Farshy CE, Black CM. Multiple drug-resistant Chlamydia trachomatis associated with clinical treatment failure. *J. Infect. Dis.* 181(4), 1421–1427 (2000).
- 86 Lefevre JC and Lepargneur JP. Comparative in vitro susceptibility of a tetracycline-resistant Chlamydia trachomatis strain isolated in Toulouse (France). *Sex. Transm. Dis.* 25(7), 350–352 (1998).
- 87 Jones RB, Van der Pol B, Martin DH, Shepard MK. Partial characterization of Chlamydia trachomatis isolates resistant to

- multiple antibiotics. *J. Infect. Dis.* 162(6), 1309–1315 (1990).
- 88 Dreses-Werringloer U, Padubrin I, Zeidler H, Kohler L. Effects of azithromycin and rifampin on Chlamydia trachomatis infection in vitro. *Antimicrob. Agents Chemother.* 45(11), 3001–3008 (2001).
- 89 Donati M, Rodriguez Fermepin M, Olmo A, D'Apote L, Cevenini R. Comparative in-vitro activity of moxifloxacin, minocycline and azithromycin against Chlamydia spp. *J. Antimicrob. Chemother.* 43(6), 825–827 (1999).
- 90 Misyurina OY, Chipitsyna EV, Finashutina YP *et al.* Mutations in a 23S rRNA gene of Chlamydia trachomatis associated with resistance to macrolides. *Antimicrob. Agents Chemother.* 48(4), 1347–1349 (2004).
- 91 Rice RJ, Bhullar V, Mitchell SH, Bullard J, Knapp JS. Susceptibilities of Chlamydia trachomatis isolates causing uncomplicated female genital tract infections and pelvic inflammatory disease. *Antimicrob. Agents Chemother.* 39(3), 760–762 (1995).
- 92 Bhengraj AR, Vardhan H, Srivastava P, Salhan S, Mittal A. Decreased susceptibility to azithromycin and doxycycline in clinical isolates of Chlamydia trachomatis obtained from recurrently infected female patients in India. *Chemotherapy* 56(5), 371–377 (2010).
- 93 Kaul R, Kimani J, Nagelkerke NJ *et al.* Monthly antibiotic chemoprophylaxis and incidence of sexually transmitted infections and HIV-1 infection in Kenyan sex workers: a randomized controlled trial. *JAMA* 291(21), 2555–2562 (2004).
- 94 Luntamo M, Kulmala T, Mbewe B, Cheung YB, Maleta K, Ashorn P. Effect of repeated treatment of pregnant women with sulfadoxine-pyrimethamine and azithromycin on preterm delivery in Malawi: a randomized controlled trial. *Am. J. Trop. Med. Hyg.* 83(6), 1212–1220 (2010).
- 95 Wawer MJ, Sewankambo NK, Serwadda D *et al.* Control of sexually transmitted diseases for AIDS prevention in Uganda: a randomised community trial. Rakai Project Study Group. *Lancet* 353(9152), 525–535 (1999).
- 96 Cowan FM, Hargrove JW, Langhaug LF *et al.* The appropriateness of core group interventions using presumptive periodic treatment among rural Zimbabwean women who exchange sex for gifts or money. *J. Acquir. Immune Defic. Syndr.* 38(2), 202–207 (2005).
- 97 Hay PE. Therapy of bacterial vaginosis. *J. Antimicrob. Chemother.* 41(1), 6–9 (1998).
- 98 Eschenbach DA. Bacterial vaginosis: resistance, recurrence, and/or reinfection? *Clin. Infect. Dis.* 44(2), 220–221 (2007).
- 99 Schwebke JR and Desmond RA. A randomized trial of the duration of therapy with metronidazole plus or minus azithromycin for treatment of symptomatic bacterial vaginosis. *Clin. Infect. Dis.* 44(2), 213–219 (2007).
- 100 Cohen MS, Hoffman IF, Royce RA *et al.* Reduction of concentration of HIV-1 in semen after treatment of urethritis: implications for prevention of sexual transmission of HIV-1. AIDSCAP Malawi Research Group. *Lancet* 349(9069), 1868–1873 (1997).
- 101 McClelland RS, Wang CC, Mandaliya K *et al.* Treatment of cervicitis is associated with decreased cervical shedding of HIV-1. *AIDS (Lond., Engl.)*, 15(1), 105–110 (2001).
- 102 Wang CC, McClelland RS, Reilly M *et al.* The effect of treatment of vaginal infections on shedding of human immunodeficiency virus type 1. *J. Infect. Dis.* 183(7), 1017–1022 (2001).
- 103 Price MA, Zimba D, Hoffman IF *et al.* Addition of treatment for trichomoniasis to syndromic management of urethritis in Malawi: a randomized clinical trial. *Sex. Transm. Dis.* 30(6), 516–522 (2003).
- 104 Gray RH, Wabwire-Mangen F, Kigozi G *et al.* Randomized trial of presumptive sexually transmitted disease therapy during pregnancy in Rakai, Uganda. *Am. J. Obstet. Gynecol.* 185(5), 1209–1217 (2001).
- 105 Aires FT, Soares RP and Bernardo WM. Efficacy of azithromycin on the treatment of syphilis. *Rev. Assoc. Med. Bras.* 56(5), 496 (2010).
- 106 Zhou P, Qian Y, Xu J, Gu Z and Liao K. Occurrence of congenital syphilis after maternal treatment with azithromycin during pregnancy. *Sex. Transm. Dis.* 34(7), 472–474 (2007).
- 107 Luntamo M, Kulmala T, Mbewe B, Cheung Y B, Maleta K, Ashorn P. Effect of Repeated Treatment of Pregnant Women with Sulfadoxine-Pyrimethamine and Azithromycin on Preterm Delivery in Malawi: A Randomized Controlled Trial. *Am. J. Trop. Med. Hyg.* 83, 1212–1220 (2010).
- 108 Kigozi GG, Brahmabhatt H, Wabwire-Mangen F *et al.* Treatment of Trichomonas in pregnancy and adverse outcomes of pregnancy: a subanalysis of a randomized trial in Rakai, Uganda. *Am. J. Obstet. Gynecol.* 189(5), 1398–1400 (2003).
- 109 Klebanoff MA, Carey JC, Hauth JC *et al.* Failure of metronidazole to prevent preterm delivery among pregnant women with asymptomatic Trichomonas vaginalis infection. *N. Engl. J. Med.* 345(7), 487–493 (2001).
- 110 Stringer E, Read JS, Hoffman I, Valentine M, Aboud S, Goldenberg RL. Treatment of trichomoniasis in pregnancy in sub-Saharan Africa does not appear to be associated with low birth weight or preterm birth. *Samj South Afr. Med. J.* 100(1), 58–64 (2010).
- 111 Brocklehurst P, Gordon A, Heatley E, Milan SJ. Antibiotics for treating bacterial vaginosis in pregnancy. *Cochrane Database Syst. Rev.* 1, CD000262 (2013).
- 112 Nugent RP, Krohn MA, Hillier SL. Reliability of diagnosing bacterial vaginosis is improved by a standardized method of gram stain interpretation. *J. Clin. Microbiol.* 29(2), 297–301 (1991).
- 113 World Health Organization and Special Programme for Research and Training in Tropical Diseases. *The use of rapid syphilis tests.* WHO, Geneva (2006).
- 114 World Health Organization. *Global Prevalence and Incidence of Selected Curable Sexually Transmitted Infections: Overview and Estimates.* World Health Organization, Geneva (2001).
- 115 Mayaud P, ka-Gina G, Cornelissen J *et al.* Validation of a WHO algorithm with risk assessment for the clinical management of vaginal discharge in Mwanza, Tanzania. *Sex. Transm. Infect.* 74(Suppl. 74), S77–S84 (1998).
- 116 Vuylsteke B, Laga M, Alary M *et al.* Clinical algorithms for the screening of women for gonococcal and chlamydial infection: evaluation of pregnant women and prostitutes in Zaire. *Clin. Infect. Dis.* 17(1), 82–88 (1993).
- 117 Costello Daly C, Wangel AM, Hoffman IF *et al.* Validation of the WHO diagnostic algorithm and development of an alternative scoring system for the management of women presenting with vaginal discharge in Malawi. *Sex. Transm. Infect.* 74(Suppl. 74), S50–S58 (1998).
- 118 Tann CJ, Mpairwe H, Morison L *et al.* Lack of effectiveness of syndromic management in targeting vaginal infections in pregnancy in Entebbe, Uganda. *Sex. Transm. Infect.* 82(4), 285–289 (2006).

- 119 Chico RM, Ariti C, Cano J, Chandramohan D, Greenwood B. Malaria transmission intensity and the protective effect of intermittent preventive therapy using sulphadoxine-pyrimethamine. *Evidence Review Group of the World Health Organization* (2013).
- 120 Temmerman M, Njagi E, Nagelkerke N, Ndinya-Achola J, Plummer FA, Meheus A. Mass antimicrobial treatment in pregnancy. A randomized, placebo-controlled trial in a population with high rates of sexually transmitted diseases. *J. Reprod. Med.* 40(3), 176–180 (1995).
- 121 Elliott B, Brunham RC, Laga M *et al.* Maternal gonococcal infection as a preventable risk factor for low birth weight. *J. Infect. Dis.* 161(3), 531–536 (1990).
- 122 Johnson HL, Ghanem KG, Zenilman JM, Erbelding EJ. Sexually transmitted infections and adverse pregnancy outcomes among women attending inner city public sexually transmitted diseases clinics. *Sex. Transm. Dis.* 38(3), 167–171 (2011).
- 123 Silveira MF, Ghanem KG, Erbelding EJ *et al.* Chlamydia trachomatis infection during pregnancy and the risk of preterm birth: A case-control study. *Int. J. STD AIDS* 20(7), 465–469 (2009).
- 124 Blas MM, Canchihuaman FA, Alva IE, Hawes SE. Pregnancy outcomes in women infected with Chlamydia trachomatis: A population-based cohort study in Washington State. *Sex. Transm. Infect.* 83(4), 314–318 (2007).
- 125 Kovacs L, Nagy E, Berik I, Meszaros G, Deak J, Nyari T. The frequency and the role of Chlamydia trachomatis infection in premature labor. *Int. J. Gynaecol. Obstet.* 62(1), 47–54 (1998).
- 126 Investigators of the Johns Hopkins Study of Cervicitis and Adverse Pregnancy Outcome. Association of Chlamydia trachomatis and Mycoplasma hominis with intrauterine growth retardation and preterm delivery. *Am. J. Epidemiol.* 129, 1247–1257 (1989).
- 127 Meis PJ, Goldenberg RL, Mercer B *et al.* The preterm prediction study: significance of vaginal infections. National Institute of Child Health and Human Development Maternal-Fetal Medicine Units Network. *Am. J. Obstet. Gynecol.* 173(4), 1231–1235 (1995).
- 128 Minkoff H, Grunebaum AN, Schwarz RH *et al.* Risk factors for prematurity and premature rupture of membranes: a prospective study of the vaginal flora in pregnancy. *Am. J. Obstet. Gynecol.* 150(8), 965–972 (1984).
- 129 Hillier SL, Nugent RP, Eschenbach DA *et al.* Association between bacterial vaginosis and preterm delivery of a low-birth-weight infant. The Vaginal Infections and Prematurity Study Group. *N. Engl. J. Med.* 333(26), 1737–1742 (1995).
- 130 Kiddugavu MG, Kiwanuka N, Wawer MJ *et al.* Effectiveness of syphilis treatment using azithromycin and/or benzathine penicillin in Rakai, Uganda. *Sex. Transm. Dis.* 32(1), 1–6 (2005).
- 131 Hook EW 3rd, Stephens J, Ennis DM. Azithromycin compared with penicillin G benzathine for treatment of incubating syphilis. *Ann. Int. Med.* 131(6), 434–437 (1999).
- 132 Riedner G, Rusizoka M, Todd J *et al.* Single-dose azithromycin versus penicillin G benzathine for the treatment of early syphilis. *N. Engl. J. Med.* 353(12), 1236–1244 (2005).
- 133 Chen CY, Chi KH, Pillay A, Nachamkin E, Su JR, Ballard RC. Detection of the A2058G and A2059G 23S rRNA gene point mutations associated with azithromycin resistance in Treponema pallidum by use of a TaqMan real-time multiplex PCR assay. *J. Clin. Microbiol.* 51(3), 908–913 (2013).
- 134 Muller EE, Paz-Bailey G and Lewis DA. Macrolide resistance testing and molecular subtyping of Treponema pallidum strains from southern Africa. *Sex. Transm. Infect.* 88(6), 470–474 (2012).
- 135 The A2058G Prevalence Workgroup. Prevalence of the 23S rRNA A2058G point mutation and molecular subtypes in Treponema pallidum in the United States, 2007 to 2009. *Sex. Transm. Dis.* 39(10), 794–798 (2012).
- 136 Matejkova P, Flasarova M, Zakoucka H *et al.* Macrolide treatment failure in a case of secondary syphilis: a novel A2059G mutation in the 23S rRNA gene of Treponema pallidum subsp. pallidum. *J. Med. Microbiol.* 58(Pt 6), 832–836 (2009).
- 137 Martin IE, Gu W, Yang Y, Tsang RS. Macrolide resistance and molecular types of Treponema pallidum causing primary syphilis in Shanghai, China. *Clin. Infect. Dis.* 49(4), 515–521 (2009).
- 138 Tipple C, McClure MO, Taylor GP. High prevalence of macrolide resistant Treponema pallidum strains in a London centre. *Sex. Transm. Infect.* 87(6), 486–488 (2011).
- 139 Rekart ML, Patrick DM, Chakraborty B *et al.* Targeted mass treatment for syphilis with oral azithromycin. *Lancet* 361(9354), 313–314 (2003).
- 140 Muldoon EG, Walsh A, Crowley B, Mulcahy F. Treponema pallidum azithromycin resistance in Dublin, Ireland. *Sex. Transm. Dis.* 39(10), 784–786. (2012).
- 141 Martin IE, Tsang RS, Sutherland K *et al.* Molecular typing of Treponema pallidum strains in western Canada: predominance of 14d subtypes. *Sex. Transm. Dis.* 37(9), 544–548 (2010).
- 142 Morshed MG and Jones HD. Treponema pallidum macrolide resistance in BC. *CMAJ* 174(3), 349 (2006).
- 143 Khaki P, Bhalla P, Sharma A, Kumar V. Correlation between In vitro susceptibility and treatment outcome with azithromycin in gonorrhoea: a prospective study. *Indian J. Med. Microbiol.* 25(4), 354–357 (2007).
- 144 Olsen B, Pham TL, Golparian D, Johansson E, Tran HK, Unemo M. Antimicrobial susceptibility and genetic characteristics of Neisseria gonorrhoeae isolates from Vietnam, 2011. *BMC Infect. Dis.* 13, 40 (2013).
- 145 Lahra MM. Annual report of the Australian Gonococcal Surveillance Programme, 2011. *Commun. Dis. Intell. Q Rep.* 36(2), E166–E173. (2012).
- 146 Lo JY, Ho KM, Lo AC. Surveillance of gonococcal antimicrobial susceptibility resulting in early detection of emerging resistance. *J. Antimicrob. Chemother.* 67(6), 1422–1426 (2012).
- 147 Lefebvre B, Bourgault AM. P1-S1.44 Antimicrobial susceptibility profile of Neisseria gonorrhoeae isolates in the Province of Quebec - 2010. *Sex. Transm. Infect.* 87 (Suppl. 87), A117 (2011).
- 148 Hottes TS, Lester RT, Hoang LM *et al.* Cephalosporin and azithromycin susceptibility in Neisseria gonorrhoeae isolates by site of infection, British Columbia, 2006 to 2011. *Sex. Transm. Dis.* 40(1), 46–51 (2013).
- 149 Yuan LF, Yin YP, Dai XQ *et al.* Resistance to azithromycin of Neisseria gonorrhoeae isolates from 2 cities in China. *Sex. Transm. Dis.* 38(8), 764–768 (2011).
- 150 Takahashi S, Kurimura Y, Hashimoto J *et al.* Antimicrobial susceptibility and penicillin-binding protein 1 and 2 mutations in Neisseria gonorrhoeae isolated from male urethritis in Sapporo, Japan. *J. Infect. Chemother.* 19(1), 50–56 (2013).
- 151 Herchline TE, Inkrott BP. Resistance trends in Neisseria gonorrhoeae in southwestern

- Ohio. *Sex. Transm. Dis.* 37(2), 121–122 (2010).
- 152 Olsen B, Mansson F, Camara C *et al.* Phenotypic and genetic characterisation of bacterial sexually transmitted infections in Bissau, Guinea-Bissau, West Africa: a prospective cohort study. *BMJ Open.* 2(2), e000636 (2012).
- 153 Tanaka M, Koga Y, Nakayama H *et al.* Antibiotic-resistant phenotypes and genotypes of *Neisseria gonorrhoeae* isolates in Japan: identification of strain clusters with multidrug-resistant phenotypes. *Sex. Transm. Dis.* 38(9), 871–875 (2011).
- 154 Bala M. Characterization of profile of multidrug-resistant *Neisseria gonorrhoeae* using old and new definitions in India over a decade: 2000–2009. *Sex. Transm. Dis.* 38(11), 1056–1058 (2011).
- 155 Chisholm SA, Neal TJ, Alawattagama AB, Birley HD, Howe RA, Ison CA. Emergence of high-level azithromycin resistance in *Neisseria gonorrhoeae* in England and Wales. *J. Antimicrob. Chemother.* 64(2), 353–358 (2009).
- 156 Khaki P, Bhalla P, Sharma P, Chawla R, Bhalla K. Epidemiological analysis of *Neisseria gonorrhoeae* isolates by antimicrobial susceptibility testing, auxotyping and serotyping. *Indian J. Med. Microbiol.* 225–229 (2007).
- 157 Enders M, Turnwald-Maschler A, Regnath T. Antimicrobial resistance of *Neisseria gonorrhoeae* isolates from the Stuttgart and Heidelberg areas of southern Germany. *Eur. J. Clin. Microbiol. Infect. Dis.* 25(5), 318–322 (2006).
- 158 Vorobieva V, Firsova N, Ababkova T *et al.* Antibiotic susceptibility of *Neisseria gonorrhoeae* in Arkhangelsk, Russia. *Sex. Transm. Infect.* 83(2), 133–135 (2007).
- 159 Sutrisna A, Soebjakto O, Wignall FS *et al.* Increasing resistance to ciprofloxacin and other antibiotics in *Neisseria gonorrhoeae* from East Java and Papua, Indonesia, in 2004 - implications for treatment. *J. Clin. Pathol.* 60(1), 90–91 (2007).
- 160 Martin IM, Hoffmann S, Ison CA. European Surveillance of Sexually Transmitted Infections (ESSTI): the first combined antimicrobial susceptibility data for *Neisseria gonorrhoeae* in Western Europe. *J. Antimicrob. Chemother.* 58(3), 587–593 (2006).
- 161 Chaudhary C, Hasan Chaudhary FA, Pandey AR *et al.* A pilot study on antimicrobial susceptibility of *Neisseria gonorrhoeae* isolates from Nepal. *Sex. Transm. Dis.* 32(10), 641–643 (2005).
- 162 Chen PL, Hsieh YH, Lee HC *et al.* Suboptimal therapy and clinical management of gonorrhoea in an area with high-level antimicrobial resistance. *Int. J. STD AIDS* 20(4), 225–228 (2009).
- 163 Hsueh PR, Tseng SP, Teng LJ, Ho SW. High prevalence of ciprofloxacin-resistant *Neisseria gonorrhoeae* in Northern Taiwan. *Clin. Infect. Dis.* 40(1), 188–192 (2005).
- 164 Aydin D, Kucukbasmaci O, Gonullu N, Aktas Z. Susceptibilities of *Neisseria gonorrhoeae* and *Ureaplasma urealyticum* isolates from male patients with urethritis to several antibiotics including telithromycin. *Clin. Infect. Dis.* 40(11), 1608–1616 (2005).
- 165 Moodley P, Pillay C, Goga R, Kharsany AB, Sturm AW. Evolution in the trends of antimicrobial resistance in *Neisseria gonorrhoeae* isolated in Durban over a 5 year period: impact of the introduction of syndromic management. *J. Antimicrob. Chemother.* 48(6), 853–859 (2001).
- 166 Kobayashi I, Kanayama A, Saika T *et al.* Tendency toward increase in the frequency of isolation of beta-lactamase-nonproducing *Neisseria gonorrhoeae* exhibiting penicillin resistance, and recent emergence of multidrug-resistant isolates in Japan. *Pediatrics* 112(1 Pt 1), 87–95 (2003).
- 167 Llanes R, Sosa J, Guzman D *et al.* Antimicrobial susceptibility of *Neisseria gonorrhoeae* in Cuba (1995–1999): implications for treatment of gonorrhoea. *Sex. Transm. Dis.* 30(1), 25–29 (2003).
- 168 Sosa J, Ramirez-Arcos S, Ruben M *et al.* High percentages of resistance to tetracycline and penicillin and reduced susceptibility to azithromycin characterize the majority of strain types of *Neisseria gonorrhoeae* isolates in Cuba, 1995–1998. *Sex. Transm. Dis.* 30(5), 443–448 (2003).
- 169 Dillon JA, Rubabaza JP, Benzaken AS *et al.* Reduced susceptibility to azithromycin and high percentages of penicillin and tetracycline resistance in *Neisseria gonorrhoeae* isolates from Manaus, Brazil, 1998. *Sex. Transm. Dis.* 28(9), 521–526 (2001).
- 170 Zaranonelli L, Borthagaray G, Lee EH, Shafer WM. Decreased azithromycin susceptibility of *Neisseria gonorrhoeae* due to *mtrR* mutations. *J. Antimicrob. Chemother.* 44(3), 411–414 (1999).
- 171 Young H, Moyes A, McMillan A. Azithromycin and erythromycin resistant *Neisseria gonorrhoeae* following treatment with azithromycin. *J. Antimicrob. Chemother.* 39(5), 623–630. (1997).
- 172 Mehaffey PC, Putnam SD, Barrett MS, Jones RN. Evaluation of in vitro spectra of activity of azithromycin, clarithromycin, and erythromycin tested against strains of *Neisseria gonorrhoeae* by reference agar dilution, disk diffusion, and Etest methods. *Clin. Infect. Dis.* 22(2), 233–239. (1996).
- 173 Dillon JA, Li H, Sealy J, Ruben M, Prabhakar P. Antimicrobial susceptibility of *Neisseria gonorrhoeae* isolates from three Caribbean countries: Trinidad, Guyana, and St. Vincent. *Sex. Transm. Dis.* 28(9), 508–514 (2001).
- 174 van Rijsoort-Vos JH, Stolz E, Verbrugh HA, Kluytmans JA. In-vitro activity of a new quinolone (CP-99,219) compared with ciprofloxacin, pefloxacin, azithromycin and penicillin against *Neisseria gonorrhoeae*. *J. Antimicrob. Chemother.* 36(1), 215–218 (1995).
- 175 Ison CA, Roope NS, Dangor Y, Radebe F, Ballard R. Antimicrobial susceptibilities and serotyping of *Neisseria gonorrhoeae* in southern Africa: influence of geographical source of infection. *Epidemiol. Infect.* 110(2), 297–305 (1993).
- 176 Starnino S, Stefanelli P. Azithromycin-resistant *Neisseria gonorrhoeae* strains recently isolated in Italy. *J. Antimicrob. Chemother.* 63(6), 1200–1204 (2009).
- 177 Donegan EA, Wirawan DN, Muliawan P *et al.* Fluoroquinolone-resistant *Neisseria gonorrhoeae* in Bali, Indonesia: 2004. *Sex. Transm. Dis.* 33(10), 625–629 (2006).
- 178 Morris SR, Moore DF, Hannah PB *et al.* Strain typing and antimicrobial resistance of fluoroquinolone-resistant *Neisseria gonorrhoeae* causing a California infection outbreak. *J. Clin. Microbiol.* 47(9), 2944–2949 (2009).
- 179 Ieven M, Van Looveren M, Sudigdoadi S *et al.* Antimicrobial susceptibilities of *Neisseria gonorrhoeae* strains isolated in Java, Indonesia. *Sex. Transm. Dis.* 30(1), 25–29 (2003).
- 180 Centers for Disease C and Prevention. Fluoroquinolone-resistance in *Neisseria gonorrhoeae*, Hawaii, 1999, and decreased susceptibility to azithromycin in *N. gonorrhoeae*, Missouri, 1999. *MMWR Morb. Mortal. Wkly Rep.* 49(37), 833–837 (2000).
- 181 Bruck PE, Robertson C, Allan PS. Management of *Neisseria gonorrhoeae* infection over 12 months in a genitourinary medicine setting against British Association for Sexual Health and HIV auditable outcome measures. *Int. J. STD AIDS* 23(3), e30–e32 (2012).

- 182 Dan M, Mor Z, Gottlieb S, Sheinberg B, Shohat T. Trends in antimicrobial susceptibility of *Neisseria gonorrhoeae* in Israel, 2002 to 2007, with special reference to fluoroquinolone resistance. *Sex. Transm. Dis.* 37(7), 451–453 (2010).
- 183 McLean CA, Wang SA, Hoff GL *et al.* The emergence of *Neisseria gonorrhoeae* with decreased susceptibility to Azithromycin in Kansas City, Missouri, 1999 to 2000. *Sex. Transm. Dis.* 31(2), 73–78 (2004).
- 184 Arreaza L, Vazquez F, Alcalá B, Otero L, Salcedo C, Vazquez JA. Emergence of gonococcal strains with resistance to azithromycin in Spain. *J. Antimicrob. Chemother.* 51(1), 190–191 (2003).
- 185 Rahangdale L, Guerry S, Bauer HM *et al.* An observational cohort study of Chlamydia trachomatis treatment in pregnancy. *Sex. Transm. Dis.* 33(2), 106–110 (2006).
- 186 Miller JM. Efficacy and tolerance of single-dose azithromycin for treatment of chlamydial cervicitis during pregnancy. *Infect. Dis. Obstet. Gynecol.* 3(5), 189–192 (1995).
- 187 Ljubin-Sternak S, Mestrovic T, Vilibic-Cavlek T *et al.* In vitro susceptibility of urogenital *Chlamydia trachomatis* strains in a country with high azithromycin consumption rate. *Folia Microbiol.* 29, 29 (2012).
- 188 Donati M, Di Francesco A, D'Antuono A *et al.* In vitro activities of several antimicrobial agents against recently isolated and genotyped *Chlamydia trachomatis* urogenital serovars D through K. *Antimicrob. Agents Chemother.* 54(12), 5379–5380 (2010).
- 189 Hong KC, Schachter J, Moncada J, Zhou Z, House J, Lietman TM. Lack of macrolide resistance in *Chlamydia trachomatis* after mass azithromycin distributions for trachoma. *Emerg. Infect. Dis.* 15(7), 1088–1090 (2009).
- 190 Lefevre JC, Escaffre MC, Courdil M, Lareng MB. In vitro evaluation of activities of azithromycin, clarithromycin and sparfloxacin against *Chlamydia trachomatis*. *Pathol. Biol.* 41(4), 313–315 (1993).
- 191 Agacfidan A, Moncada J, Schachter J. In vitro activity of azithromycin (CP-62,993) against *Chlamydia trachomatis* and *Chlamydia pneumoniae*. *Antimicrob. Agents Chemother.* 37(9), 1746–1748 (1993).
- 192 Scieux C, Bianchi A, Chappey B, Vassias I, Perol Y. In-vitro activity of azithromycin against *Chlamydia trachomatis*. *J. Antimicrob. Chemother.* 7–10 (1990).
- 193 Somani J, Bhullar VB, Workowski KA, Farshy CE, Black CM. Multiple drug-resistant *Chlamydia trachomatis* associated with clinical treatment failure. *J. Infect. Dis.* 181(4), 1421–1427 (2000).
- 194 van den Broek NR, White SA, Goodall M *et al.* The APPLe Study: A Randomized, Community-Based, Placebo-Controlled Trial of Azithromycin for the Prevention of Preterm Birth, with Meta-Analysis. *PLoS Med.* 6(12), e1000191 (2009).
- 195 Luntamo M, Kulmala T, Cheung YB, Maleta K, Ashorn P. The effect of antenatal monthly sulphadoxine-pyrimethamine, alone or with azithromycin, on foetal and neonatal growth faltering in Malawi: a randomised controlled trial. *Trop. Med. Int. Health* 18(4), 386–397 (2013).
- 196 Chico RM, Chandramohan D. Azithromycin plus chloroquine: combination therapy for protection against malaria and sexually transmitted infections in pregnancy. *Expert Opin. Drug Metab. Toxicol.* 7(9), 1153–1167 (2011).
- 197 Labbe AC, Pepin J, Khonde N *et al.* Periodical antibiotic treatment for the control of gonococcal and chlamydial infections among sex workers in Benin and Ghana: a cluster-randomized placebo-controlled trial. *Sex. Transm. Dis.* 39(4), 253–259 (2012).
- 198 Wi T, Ramos ER, Steen R *et al.* STI declines among sex workers and clients following outreach, one time presumptive treatment, and regular screening of sex workers in the Philippines. *Sex. Transm. Infect.* 82(5), 386–391 (2006).
- 199 Williams BG, Taljaard D, Campbell CM *et al.* Changing patterns of knowledge, reported behaviour and sexually transmitted infections in a South African gold mining community. *AIDS* 17(14), 2099–2107 (2003).
- 200 Steen R, Vuylsteke B, DeCoito T *et al.* Evidence of declining STD prevalence in a South African mining community following a core-group intervention. *Sex. Transm. Dis.* 27(1), 1–8 (2000).
- 201 Jones BM, Kinghorn GR, Duerden BI. In vitro activity of azithromycin and erythromycin against organisms associated with bacterial vaginosis and chancroid. *Eur. J. Clin. Microbiol. Infect. Dis.* 7(4), 551–553 (1988).
- 202 Shanker S, Toohey M, Munro R. In vitro activity of seventeen antimicrobial agents against *Gardnerella vaginalis*. *Eur. J. Clin. Microbiol.* 1(5), 298–300 (1982).
- 203 Ridgway GL. A review of the in-vitro activity of roxithromycin against genital pathogens. *J. Antimicrob. Chemother.* 20 (Suppl. B), 7–11 (1987).
- 204 Dubreuil L. In-vitro comparison of roxithromycin and erythromycin against 900 anaerobic bacterial strains. *J. Antimicrob. Chemother.* 20(Suppl. B), 13–19 (1987).
- 205 Maskell JP, Sefton AM, Williams JD. Comparative in-vitro activity of azithromycin and erythromycin against Gram-positive cocci, *Haemophilus influenzae* and anaerobes. *J. Antimicrob. Chemother.* 25(Suppl. A), 19–24 (1990).
- 206 Chang SC, Chen YC, Luh KT, Hsieh WC. Macrolides resistance of common bacteria isolated from Taiwan. *Diag. Microbiol. Infect. Dis.* 23(4), 147–154 (1995).
- 207 Ednie LM, Spangler SK, Jacobs MR, Appelbaum PC. Antianaerobic activity of the ketolide RU 64004 compared to activities of four macrolides, five beta-lactams, clindamycin, and metronidazole. *Antimicrob. Agents Chemother.* 41(5), 1037–1041 (1997).
- 208 Mikamo H, Yin XH, Ninomiya M, Tamaya T. In vitro and in vivo antibacterial activities of telithromycin. *Chemotherapy* 49(1–2), 62–65 (2003).
- 209 Marina M, Ivanova M, Kantardjiev T. Antimicrobial susceptibility of anaerobic bacteria in Bulgaria. *Anaerobe* 15(4), 127–132 (2009).
- 210 Chen SC, Gottlieb T, Palmer JM, Morris G, Gilbert GL. Antimicrobial susceptibility of anaerobic bacteria in Australia. *J. Antimicrob. Chemother.* 30(6), 811–820 (1992).
- 211 Wexler HM, Molitoris E, Molitoris D, Finegold SM. In vitro activity of telithromycin (HMR 3647) against 502 strains of anaerobic bacteria. *J. Antimicrob. Chemother.* 47(4), 467–469 (2001).
- 212 Spiegel CA. Susceptibility of *Mobiluncus* species to 23 antimicrobial agents and 15 other compounds. *Antimicrob. Agents Chemother.* 31(2), 249–252 (1987).

Website

- 301 ReMeD (Network for Medicine and Development). Essential Medicines: Madagascar. *World Health Organization* www.who.int/selection_medicines/country_lists/mdg/en/.