Biological and pathological mechanisms leading to the birth of a small vulnerable newborn

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1 Summary

2 The pathway to a thriving newborn begins pre-conception and continues in utero with a 3 healthy placenta and the right balance of nutrients and growth factors that are timed and 4 sequenced alongside hormonal suppression of labour until a mature infant is ready for birth. 5 Optimal nutrition that includes adequate quantities of quality protein, energy, essential fats 6 and an extensive range of vitamins and minerals not only supports fetal growth but may also 7 prevent preterm birth by supporting the immune system and alleviating oxidative stress. 8 Infection, illness, undernourishment, and harmful environmental exposures can alter this 9 trajectory leading to an infant who is too small due to either poor growth during pregnancy or 10 preterm birth. Systemic inflammation suppresses fetal growth by interfering with growth 11 hormone and its regulation of insulin-like growth factors. Evidence supports the prevention 12 and treatment of several maternal infections during pregnancy to improve newborn health. 13 However, microbes, such as Ureaplasma species, that are able to ascend the cervix and cause 14 membrane rupture and chorioamnionitis require new strategies for detection and treatment. 15 The surge in fetal cortisol late in pregnancy is essential to parturition at the right time, but 16 acute or chronically high maternal cortisol levels caused by psychological or physical stress 17 may also trigger labour onset prematurely. In every pathway to the small vulnerable newborn, 18 there is a possibility to change direction by supporting improved nutrition, protection against 19 infection, holistic maternal wellness, and healthy environments.

20 Keywords

Preterm birth, fetal growth restriction, small for gestational age, small vulnerable newborn,
pregnancy, nutrition, infection

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25 Key messages

Factors that influence fetal growth change over course of pregnancy, from the direct
 exposure to nutrients in maternal fluids during conception, to the formation and
 function of the placenta, to the timing of bone elongation and fat deposition. Thus, the
 timing and regulation of nutrient availability is critical in achieving fetal growth
 potential.

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Pregnancy is maintained by the active suppression of labour mechanisms by
 progesterone and other factors and by a long, closed cervix. Thus, there are physical
 and chemical "barriers" to the initiation of labour and birth that are overcome by
 signals that the infant is ready to be born. The barriers can be modulated by
 progesterone insufficiency, diet and environmental contaminants. In addition, high
 levels of maternal cortisol and severe inflammation can override the barrier leading to
 preterm labour and birth.

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3. Preterm birth and fetal growth restriction may be the endpoints of different pathways
but infection, undernourishment, psychological stress and environmental exposures
have the potential to act on both pathways through intermediates of oxidative stress,
inflammation, inadequate immune protection and placental dysfunction.

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4. New knowledge about the mechanisms of pregnancy continues to emerge providing a
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49 Embedded in the United Nations' Sustainable Development Goals is a roadmap to break the 50 cycle of poverty and disadvantage perpetuated by vulnerable childhood and adolescence 51 giving rise to vulnerable pregnancy and infancy. In this series, we examine the vulnerability conferred by small size at birth resulting from growth restriction and/or preterm birth. We 52 53 cover the prevalence, causes, consequences and possible routes to prevention, either by 54 accelerating existing strategies or considering new approaches. Approximately one in four infants worldwide is born either preterm, small-for-gestational-age or both.¹ Forty per cent of 55 56 global neonatal mortality occurs in preterm infants and 28% occurs in small-for-gestationalage infants born at term.¹ 57

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59 Despite global attention and targets set for reducing the prevalence of the small vulnerable 60 newborn, there has been little change over the last 10 years.¹ The slow progress can be 61 attributed in part to gaps in our common understanding of the mechanisms controlling fetal 62 growth and gestational duration. Multiple, often interacting, risk factors contribute to poor 63 health in women both before and during pregnancy (panel 1). However, connecting risk 64 factors to the biological processes leading to preterm birth and growth restriction remains a 65 challenge. For some of the most prevalent risk factors, the relationship with causal mechanisms may be indirect. For example, maternal iron deficiency anaemia is the largest 66 67 global population-attributable risk factor for spontaneous preterm and small-for-gestationalage births,^{2,3} however iron supplementation (which reduces maternal anaemia by 70%) has 68 69 not reduced he prevalence of these outcomes in most contexts.⁴ A similar conundrum is the 70 global prevalence of bacterial vaginosis and its association with spontaneous preterm birth; 71 25 years of trials with antibiotics during pregnancy show that treatment can reduce the prevalence of bacterial vaginosis but not the risk of spontaneous preterm birth.^{5,6} 72

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74 Within the series, this article reviews the pathway to the birth of a healthy thriving newborn 75 in order to provide a framework to describe what can go wrong. Knowledge of these 76 mechanisms is incomplete, however new information is constantly emerging, often from 77 disciplines outside of mammalian reproduction and development. Novel concepts emerging 78 from randomised controlled trials, animal models, observational studies and laboratory work 79 that recapitulates mechanisms in vitro have enabled connections to be made with biological 80 mechanisms in order to explain why some strategies for prevention are effective and some 81 require new approaches. This article will demonstrate that it is useful to consider preterm 82 birth and growth restriction together because many risk factors can contribute to both, albeit 83 through different pathways. Context-specific, targeted and even personalised intervention 84 strategies to prevent preterm and small-for-gestational-age births are possible and likely to 85 bring better health to the next generation.

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87 Born at the right size but how?

88 Factors influencing the growth and development of the fetus change over the course of 89 pregnancy. The first critical period begins around the time of conception and ends at 90 implantation. At this stage, the embryo can sense the concentrations of nutrients in the 91 surrounding fluids and calibrate of metabolic processes to compensate for overabundance, in the case of maternal obesity, or paucity, in the case of undernutrition.⁷ The subsequent 92 93 adaptations in embryonic gene expression and regulation can become "fixed" in the form of 94 heritable chromatin changes that can lead to dysregulated fetal growth and obesity and metabolic disease in adulthood.⁸ 95

96 The next critical period begins with implantation, which triggers a hormonal surge leading to 97 changes in maternal physiology to support placental development and the increased 98 metabolic demands of pregnancy. Fetal trophoblast cells invade the maternal endometrial

99 spiral arteries, displacing the vascular endothelium and directing larger, stronger versions to be rebuilt on the same tissue scaffold.⁹ Proliferating trophoblasts elaborate the basic placental 100 101 structure, which consists of finger-like villi that float in compartments of maternal blood 102 (Figure 1). Peak placental growth occurs at the end of the first trimester but remodelling of 103 the maternal vasculature continues for the duration of pregnancy (Figure 2). 104 As the placenta develops, it takes over the production of hormones that maintain pregnancy 105 and direct the production of growth factors (Figure 3). Thus, a physiological dialog ensues 106 between the placenta and fetus, and the placenta and pregnant woman. For example, 107 placentally produced hormones create a transient state of mild insulin resistance at the cellular level in the woman, presumably to free up more glucose for the infant.¹⁰ Excess 108 109 glucose is taken up and stored as glycogen by the placenta, possibly to buffer the effects of 110 transient moderate undernourishment or to prepare for accelerated weight gain later in gestation.¹¹ 111

112 The second trimester is the critical period of peak fetal length gain, largely driven by insulin-113 like growth factors (IGFs) and regulated by growth hormone and a system of six binding proteins and their proteases.¹² IGF-1 is involved in bone elongation and skeletal growth.¹³ 114 115 IGF-2 drives placental growth as well as the synthesis of other placentally-derived hormones.¹⁴ The last trimester sees peak fetal weight gain with the enlarging of muscle and 116 117 laying down of fat under the skin and around the organs. Fat deposition is controlled and regulated by insulin, leptin, adiponectin and other adipokines.¹⁵ Undernutrition during the 118 119 third trimester leads to an infant that is too thin at birth whereas mid trimester undernutrition leads to an infant that is overall too small.¹⁶ Due to resource allocation to head and brain 120 121 development (so-called "brain sparing") head growth can follow a normal growth trajectory even when the growth of the fetal body is faltering.¹⁷ Since maternal weight gain is steadier 122 123 than that of the infant, it should be possible to identify women who are not gaining adequate

weight and intervene to support nutrient intake ahead of peak fetal weight gain in the thirdtrimester.

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127 Born at the right time

Pregnancy is maintained by progesterone-mediated suppression of the processes of labour and by an impenetrable cervix (Figure 2). Progesterone inhibits the production of components involved in receiving signals to prepare the uterus for labour such as the estrogen and oxytocin receptors. In most mammals, plasma progesterone concentrations decrease towards the end of pregnancy. In contrast, levels remain high throughout human pregnancy, even during labour. Activation of labour systems is brought about instead by the functional inhibition of progesterone, possibly by a soluble "A" form of the progesterone receptor (PR-

135 A).¹⁸

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137 The uterine cervix remains long and closed for the duration of pregnancy due to its rigid 138 structure bestowed by the high collagen content of the extracellular matrix. Compared with 139 many other mammals, the human cervix needs to be strong enough to counteract the 140 downward pressure of weight attributable to the growing fetus during the time the woman spends in the upright position.¹⁹ Additionally, the cervix needs to be kept free of bacteria 141 142 ascending from the vagina. Cervical mucus provides a scaffold for immunoglobulins and antimicrobial peptides as it accumulates and forms the mucus plug.^{20,21} The cellular defence 143 144 of the cervix is mainly provided by neutrophils that populate the mucus having exited the maternal circulation.²² 145

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Events leading to labour and birth of humans are not fully understood. However, there arepathways observed in other mammals that are likely to operate similarly in humans. A

149 common view is that signals from the infant indicating that key late developmental 150 milestones have been achieved are also able to start the processes leading to labour and birth. 151 For example, one of the final steps in lung development is the release of surfactant to the 152 surface of the lung alveoli so that when they fill with air at birth, the surface tension will be 153 kept low. Since the lungs are full of amniotic fluid and the infant is performing breathing 154 movements in the womb, the surfactant diffuses throughout the amniotic fluid around the 155 infant. In rodents, the accumulation of surfactant in amniotic fluid acts as a trigger to start the birth process.^{23,24} 156

A similar process occurs with fetal cortisol and corticosteroids. Towards the end of pregnancy, the fetal brain signals to increase production of corticotropin releasing hormone which leads to an increase in cortisol and corticosteroids in the fetal circulation (Figure 2).²⁵ As the main steroid involved in the stress response, cortisol directs the release of glucose into the fetal bloodstream and increases blood flow to the brain. It may have the dual function of bringing new alertness and awareness to the infant as well as signalling that the infant is ready for parturition to begin.

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165 The first committed step toward labour occurs when cortisol and corticosteroids in the fetal circulation reach the threshold for the activation of the production of the cyclooxygenase 2 166 167 (COX2) in the fetal membranes (figure 2). COX2 converts long chain polyunsaturated fatty 168 acids (LCPUFAs) into prostaglandins. The essential LCPUFA for labour is arachidonic acid, 169 which selectively accumulates in the myometrium, cervix and fetal membranes over the course of pregnancy.²⁶ COX2 converts arachidonic acid into prostaglandins E2 and F2 α , 170 which trigger a gene and protein expression cascade, leading to the functional inhibition of 171 progesterone, the production of contraction-associated proteins and the recruitment of 172 monocytes and neutrophils to the uterus and cervix.²⁷ These cells produce matrix 173

174	metalloproteinases which dissolve the extracellular collagen matrix of the myometrium and
175	cervix causing the cervix to soften. ²⁸ Tight gap junctions form between the cells of the
176	myometrium, which then takes on the appearance and function of smooth muscle.
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178	Omega-3 LCPUFAs are also substrates of COX2 and may act as competitive inhibitors of
179	prostaglandin E2 and F2a production thus contributing to the maintenance of pregnancy and
180	the inhibition of labour. ²⁹ Women with lower circulating concentration of omega-3
181	LCPUFAs are at increased risk of preterm birth, ³⁰ suggesting that these compounds, like
182	progesterone, act to raise the threshold for the activation of labour processes. One of the
183	unintended consequences of supplementation with omega-3 LCPUFAs is an increase in the
184	rate of post term birth ³¹ suggesting that if the threshold is too high, signals from the fetus
185	can't overcome the inhibitory mechanisms and the pregnancy is prolonged.
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187	Good nutrition supports more than just growth
188	The impact of maternal nutrition before and during pregnancy is now understood to extend
189	well beyond birth and childhood into the life courses of future generations. ^{7,32} Physiological
190	changes in pregnancy enable women to meet the increased demand for energy, nutrients, and
191	oxygen to supply to the growing fetus (Table 1). However, women who begin a pregnancy
192	before having reached their own biological growth potential due to chronic
193	undernourishment, young age, or both, are at increased risk of being unable to meet these

194 demands. Among underweight women, partitioning of energy and nutrients may result in

195 limited provision to the fetus in favour of maternal requirements for her survival. Thus, it is

196 not surprising that underweight women, who may also have inadequate gestational weight

197 gain, are at higher risk of delivering a small-for-gestational-age infant.³⁹

198 Anaemia is a highly prevalent risk factor linked to a wide range of adverse pregnancy outcomes.⁴⁰ There are many causes of anaemia unrelated to nutrition including malaria and 199 200 other infectious/inflammatory conditions. However, iron supplementation during pregnancy independently reduces the prevalence of anaemia, suggesting that iron deficiency is a key 201 contributor.⁴ Anaemia, as a measurable risk factor, may also identify women with a wider 202 203 range of micronutrient deficiencies. Supplementation with a broad range of micronutrients is able to lower the risk of small-for-gestational age births,^{41,42} particularly among underweight 204 and anaemic women.⁴² in comparison to iron and folic acid alone. This positive effect on 205 206 growth without the provision of energy is likely conferred by the efficiency gained when 207 multiple metabolic processes are supported simultaneously. Provision of micronutrients may also lower the risk of preterm birth in underweight women.⁴² There are many mechanisms 208 209 that might contribute to this effect listed in Table 2. We will expand on the ability of good 210 nutrition to enhance immune responses and reduce damage caused by oxidative stress.

211 Damage to tissue caused by the accumulation of reactive oxygen species is both a threat to pregnancy and a natural consequence of oxygen regulation in the placenta.⁴⁷ Micronutrients 212 213 with antioxidant properties including vitamins C and E, carotenoids and long-chain 214 polyunsaturated fatty acids (LCPUFAs) can reduce oxidative stress. The body can dismantle 215 reactive oxygen species using enzymes such as superoxide dismutase, glutathione reductase 216 and various peroxidases that can catalyse their binding to antioxidant molecules. However, 217 once an antioxidant is peroxidated, it is removed from tissue leading to increased turnover and reduced bioavailability.⁴⁸ The pathway to spontaneous preterm birth caused by oxidative 218 219 stress may involve the increased turnover of LCPUFAs, particularly docosahexaenoic acid, 220 which, as previously discussed, may act as a natural inhibitor of labour. People who smoke cigarettes carry a higher burden of oxidative damage compared with non-smokers,⁴⁹ and have 221 lower levels of endogenous omega-3 LCPUFAs.⁵⁰ Thus, it is unsurprising that a trial 222

comparing omega-3 LCPUFA supplementation with placebo in pregnant women found
 spontaneous preterm birth reduced by almost one-half in smokers, whereas there was no
 benefit in non-smokers.⁵¹

226 Zinc is an essential co-factor for superoxide dismutase and a wide range of enzymes and 227 transcription factors, and its deficiency is associated with immune dysfunction and increased susceptibility to infection.⁵² White blood cells require tenfold more zinc in comparison to red 228 229 blood cells.⁵³ In a healthy pregnancy, there is an increase in white blood cell counts, largely due to the 50% increase in neutrophils.⁵⁴ As one of the first lines of defence against 230 231 pathogens, neutrophils are ubiquitous at points of entry into the body. In pregnancy, they are crucial to defending the cervix against ascending infection.²² Recent evidence supports 232 233 previously unknown roles for neutrophils in vascular and tissue remodelling.⁵⁵ The secretion 234 of matrix metalloproteinases, for which zinc is a cofactor, by neutrophils is likely to be essential for this latter role. Blocking neutrophils,⁵⁶ knocking out matrix metalloproteinases,⁵⁷ 235 and reducing bioavailable zinc,⁵⁸ all have detrimental effects on placentation in mice leading 236 237 to fetal demise. The roles of neutrophils and zinc in placentation and protection against pathways leading to preterm birth are only just beginning to be understood and represent a 238 239 new frontier in reproductive biology.

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241 Infectious threats to the fetus

Microbial infections in pregnant women are major contributors to preterm birth, growth restriction, stillbirth and infection in newborns. Screening for and treating infections in pregnant women has well-established positive effects and there is a need for wider coverage for syphilis, chlamydia, gonorrhoea, HIV, and malaria. However, even in parts of the world where the prevalence of these infections is low, the majority of spontaneous preterm birth – that is, preterm birth preceded by labour or preterm pre-labour rupture of membranes – is also

248 likely to be caused by microbial infection given the high prevalence of chorioamnionitis

found in membrane and placenta tissue on histopathological examination.⁵⁹⁻⁶¹

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251 Chorioamnionitis refers to infiltration of the fetal membranes by maternal neutrophils. It is 252 usually asymptomatic during pregnancy and the diagnosis is made after the birth of the 253 infant. Whilst it is presumed to be caused by colonisation by bacteria that ascended the cervix 254 from the vagina, identification of microbes in these tissues is seldom undertaken. When 255 molecular methods are used to detect microbes in fetal membranes, the most common species identified are members of the Ureaplasma genus of bacteria.⁶¹⁻⁶³ Some species of 256 257 Ureaplasma are able to break down antimicrobial defences and exploit natural weaknesses in 258 the immune system that are unmasked by pregnancy in some women. This may explain the 259 association between spontaneous preterm birth and both periodontal disease and urinary tract infections.^{64,65} The mouth, the vagina, and the urinary tract are dependent on the same 260 261 mechanisms (antibodies, antimicrobial peptides and neutrophils) to protect against microbial 262 invasion.

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264 There are three general pathways through which infection could lead to spontaneous preterm birth. First, there are likely unique features of certain bacterial species, as opposed to viruses 265 266 or parasites, that trigger the expression of COX2 on their invasion of the placenta, fetal 267 membranes or amniotic fluid. Injecting bacteria or bacterial products into the uteri of pregnant mice is the most widely-used method of modelling preterm birth.⁶⁶ It could be that 268 269 COX2 can be upregulated by signalling through molecules, such as toll-like receptors 2 and 4, that specifically recognize certain types of bacteria and bacterial products.⁶⁷ Secondly, 270 271 microbes that are able to ascend the cervix from the vagina could simply damage the fetal 272 membranes causing rupture (figure 5). In this scenario, there may not be inflammation or the

activation of mechanisms that lead to labour. In many cases of preterm pre-labour rupture of
membranes, labour does not occur after a sufficient period of time and the infant must be
delivered by labour induction or Caesarean section due to loss of amniotic fluid and the
concerns regarding the potential for systemic spread of the infection. Finally, high levels of
inflammatory cytokines in the placenta and may be able to activate COX2 and the pathways
that culminate in labour.⁶⁸ This may be an evolutionary adaptation to delivery the infant from
an unfavourable environment where the mother's life is under threat.

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Inflammation likely suppresses fetal growth by inhibiting the growth hormone/insulin-like growth factor (GH/IGF) axis (Figure 4). In a study comparing maternal plasma, placental, and cord blood levels of IGF-1 and its inhibitory binding proteins in pregnancies with and without placental malaria, IGF-1 levels were reduced by 28% in plasma samples from women with placental malaria and by 25% in their neonates compared with samples from uninfected women.⁶⁹ The inhibitory IGF binding protein-1 was elevated in cord blood of neonates with placental malaria.⁶⁹

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289 Clues to the molecular interactions between inflammation and growth factors come from the observation of poor growth in children with systemic inflammation,⁷⁰ and elevated 290 inflammation in children with poor growth.⁷¹ A surprising result of treating children with 291 292 anti-tumour necrosis factor alpha and other anti-cytokine therapeutics for inflammatory conditions was the restoration of normal growth trajectory.⁷⁰ Studies in mice indicate that 293 294 interleukin-6, a key inflammatory cytokine that is elevated in response to infection, may have the ability to directly suppress IGF-1 and growth hormone.⁷² The slowing of growth in 295 296 response to inflammation may be an evolutionary adaptation to promote successful vaginal

birth. As the mother's body prepares for labour, the increase in systemic inflammatorycvtokines may contribute to the observed slowing of head growth at the end of pregnancy.

299 Cervical shortening and preterm birth

When a woman's cervix shortens in the course of pregnancy, there is an increased risk of 300 301 preterm birth. It is not known why this occurs in some women, but it is associated with the 302 premature expression of proteins involved in the recruitment of monocytes and neutrophils which could lead to the premature destruction of collagen and loss of integrity.⁷³ As a key 303 304 hormone responsible for maintaining pregnancy, progesterone delivered directly to the cervix 305 via vaginal pessaries, injected intramuscularly (IM) or taken as tablets has been tested in randomized controlled trials to determine its effect on preterm birth. The results of these trials 306 have been controversial and contradictory,^{74,75} leading the FDA to withdraw the indication 307 for IM progesterone for the prevention of preterm birth due to lack of efficacy.⁷⁶ However, a 308 309 recent individual patient data meta-analysis revealed that both IM and vaginal progesterone 310 supplementation are more effective at reducing preterm birth in women who have a short 311 cervix (< 25 mm) compared to women with other risk factors, with evidence of benefit in reducing preterm birth before 34 weeks more certain for vaginal progesterone.⁷⁷ Serial 312 313 ultrasound surveillance of cervical length is required to reliably detect cervical shortening, 314 which may preclude the use of cervical monitoring in resource-poor settings. Analysis of 315 soluble factors in amniotic and vaginal fluids have identified macrophage chemoattractant protein 1 as a biomarker with the strongest association with cervical shortening.^{73,78,79} 316 317 Macrophage chemoattractant protein 1 is easy to detect in mucus from the vaginal end of the cervix and holds potential to report cervical shortening with minimal equipment. 318

319 **Pre-eclampsia, fetal growth restriction and preterm birth**

320 Major problems arising during implantation and early placental development result in 321 miscarriage. However, minor issues often remain silent until around mid-gestation when the 322 fetus overtakes the placenta in size. At this time, minor inadequacies in placental size, 323 patterning or maternal blood supply can result in an inability to meet the requirements for the 324 growth and development of the fetus. For reasons that are not completely understood, one of 325 the most common signs that there are supply-and-demand issues with a pregnancy is the 326 elevation of the pregnant woman's blood pressure. The clinical definition of pre-eclampsia 327 has recently been expanded to include the development of high blood pressure during pregnancy along with any related problem, not only elevated protein in the urine.⁸⁰ Five 328 329 percent of pregnancies worldwide are affected by pre-eclampsia with 76.000 attributable 330 maternal deaths per year, second only to post-partum haemorrhage as a cause of maternal 331 death. Around 500,000 fetal and newborn deaths each year are attributed to pre-eclampsia and eclampsia.⁸⁰ Approximately 9% of all preterm birth is by induction of labour or 332 Caesarean section to treat severe pre-eclampsia and eclampsia.⁸¹ 333

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Pre-existing maternal cardiovascular vulnerability and poor cardiovascular adaptation to pregnancy are increasingly recognised as important to the development of pre-eclampsia.⁸² Pregnancy has even been described as a stress-test that reveals women who have poor cardiovascular reserve or dysfunction.⁸³ It is therefore unsurprising that well-established treatments for cardiovascular disease such as low-dose aspirin, when given during pregnancy, also reduce the risk of preterm pre-eclampsia,⁸⁴ and new treatments (statins) are under investigation.⁸⁵

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A calcium-rich diet or calcium supplementation during pregnancy are also able to reduce the
 risk of pre-eclampsia and associated morbidity and mortality in the newborn.⁸⁶ It is likely that

345 both aspirin and calcium are able to prevent the establishment of a systemic vasoconstrictive 346 environment. In chronic, sustained high blood pressure, the ratio of the vasoconstrictive 347 thromboxane to the vasodilator prostacyclin is skewed towards vasoconstriction. Both 348 molecules are synthesized by cyclooxygenases 1 and 2 (COX1/2). At low doses, aspirin 349 appears to be able selectively and irreversibly to inactivate COX1 in platelets, thus reducing thromboxane production and restoring this ratio to normotensive levels.⁸⁷ However, aspirin 350 351 has been shown to be most effective at preventing preterm pre-eclampsia when commenced early in pregnancy (< 16 weeks) suggesting a supportive effect on early placentation.⁸⁸ 352

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354 Changing social and environmental contexts

355 Some subgroups of pregnant women, such as smokers, primi- and secundigravidae, 356 teenagers, and women with low body mass index scores, tend to respond more favourably to 357 nutrient supplementation or preventive treatment of infections, reducing the risk of delivering 358 small and vulnerable newborns. However, this does not justify the exclusive use of these 359 interventions strategies to reduce the prevalence of small vulnerable newborns. Increased 360 antenatal contacts afford opportunities to address the wellbeing of pregnant women in a more 361 holistic way. Depression, anxiety, lack of agency, chronic illness, physical workload and intimate partner abuse can all be exacerbated by pregnancy. High levels of psychological and 362 363 physical stress during pregnancy are associated with growth restriction and shorter pregnancy duration.⁸⁹⁻⁹¹ Cortisol entering the placenta from the fetal circulation is an important step in 364 365 the preparation of mother and child for birth. Although increases in cortisol and corticotropin releasing hormone in the mother's circulation are normal during pregnancy, it is possible that 366 367 prolonged elevated or acute bursts of cortisol may be able to trigger preterm labour. 368 Furthermore, elevated cortisol has also been associated with higher concentrations of

proinflammatory cytokines,^{92,93} that can negatively affect fetal growth as previously
described (Figure 4).

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Creation of energy from oxygen combined with glucose and other monosaccharides is the 372 373 final step in the pathway that powers fetal growth. The pathway starts with clean air that is 374 free of pollutants that interfere with oxygen binding by maternal hemoglobin. In addition to 375 increasing the burden of oxidative stress, smoking and cooking over biomass fuels can limit oxygen delivery to the placenta (Figure 4).⁹⁴ Exposure to air pollution and living at high 376 altitude have also been linked to fetal growth restriction.^{95,96} Interventions that help women to 377 quit or reduce smoking during pregnancy reduce the risk of giving birth to a small infant.⁹⁷ 378 379 Countries that have banned smoking in indoor public spaces have experienced a dramatic reduction in the prevalence of preterm and low birth weight newborns.⁹⁸⁻¹⁰⁰ Low- and middle-380 381 income countries have higher outdoor pollution levels and indoor pollution due to a reliance on solid biomass (usually wood) fuels and chimneyless stoves for cooking and heating.¹⁰¹ 382 383 Because women are more exposed to indoor pollution from cookstoves and heating due to a 384 greater amount of time spent in the home, the World Health Organization considers indoor pollution as a "silent killer" of women in low-resource settings.¹⁰² Trials of liquid fuel 385 386 cookstoves have so far failed to significantly lower the risk of low birth weight, preterm birth 387 or small-for-gestational-age births, potentially through being unable to reduce sufficient airborne particulate matter to have an observable effect.^{103,104} 388 389 New evidence is emerging on the effect extra heat on pregnancy outcomes, with a 5% (95%) 390 CI 3% - 7%) increase in the odds of having a preterm birth every one degrees above seasonal average.^{105,106} Further epidemiological evidence suggests that conception and early first 391

trimester are particularly vulnerable to heat stress, increasing the risk of stillbirth and preterm

393 birth.¹⁰⁷ In animals, transient elevated temperatures lead to reduced feeding and overall food

intake resulting in growth restriction in the fetus.¹⁰⁸ However, the damage may run deeper
with loss of intestinal barrier function, changes to intestinal epithelial morphology.¹⁰⁹

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397 Food and water-borne pollutants are also likely to contribute to the prevalence of small 398 vulnerable newborns. Components of Aspergillus fungal spores collectively known as aflatoxins are common contaminants of food production in under-resourced settings.¹¹⁰ High 399 concentrations of aflatoxins in maternal and cord blood are associated with low birthweight, 400 likely mediated through growth restriction, although the exact mechanism is not known.¹¹¹ In 401 402 addition to known teratogenic and carcinogenic effects of aflatoxins, they may also interfere 403 with hormone secretion and signaling and thus are part of a wider group of both natural and 404 artificial toxicants known as endocrine disruptors, which include bisphenol A, phthalates, pesticides, polychlorinated biphenyls, polybrominated diethyl ethers and dioxins.¹¹² Of 405 406 particular concern is the high levels of phthalate metabolites that contaminate food and water 407 globally. In keeping with their role in modulating estrogen levels, different phthalate 408 compounds can increase or reduce gestational length and are therefore associated with both pre- and post-term birth.¹¹³ Governments have sought to ban the use of phthalates in plastics 409 production, however the toxicity of potential replacements is uncertain.¹¹² 410

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412 What can be done? The foreground and the horizon

Knowledge of the mechanisms that lead to the birth of a small vulnerable newborn continues to grow as well as our understanding of how to intervene to reduce or prevent this outcome. In the short term, increasing the quantity and quality of antenatal contacts with healthcare providers affords the opportunity to monitor and support physical (weight gain, fetal growth, prevention and treatment of pregnancy complications) and psychological (mental health, agency) wellbeing. Significant reduction in preterm birth and growth restriction can be

419 achieved with broader implementation of proven antenatal interventions, including multiple 420 micronutrient supplements, balanced protein energy supplements, aspirin, treatment of 421 syphilis, education for smoking cessation, prevention of malaria in pregnancy, treatment of 422 asymptomatic bacteriuria, and progesterone provided vaginally as presented with this 423 series.¹¹⁴ In addition, the specific vulnerability of those *in utero* to poor air quality, heat 424 waves and toxins in food and water should contribute the urgency of global efforts to reduce 425 harmful environmental exposures and the impact of climate change.

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427 In the longer term, new knowledge can be used to improve our understanding of the 428 molecular and cellular biology underlying risk factors that inform interventions for 429 populations with the greatest ability to benefit. Risk stratification tools and algorithms that 430 incorporate individual risk profiles, together with biomarkers, can identify individuals who 431 might benefit from pre-emptive care and early pathway-specific interventions. For example, a 432 test that predicts future cervical shortening would identify women who are most likely to 433 benefit from progesterone supplementation without the need for serial ultrasound monitoring. 434 Progesterone supplementation itself is also evolving with new analogues that are resistant to inhibition by the mechanisms that lead to labour.¹¹⁵ Tests that can be performed and 435 436 interpreted in the timescale of an antenatal care visit (point-of-care tests) will improve uptake 437 of treatment for infections; treatment can be issued on the same day removing the need to 438 return to clinic for follow-up. Point-of-care tests should fulfil the WHO ASSURED 439 (Affordable, Sensitive, Specific, User-friendly, Rapid, and Equipment-free, and Deliverable) criteria for use in low resource settings.¹¹⁶ 440

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Placental histopathology is underutilized as a means to diagnose chorioamnionitis and otherplacental conditions leading to birth of small vulnerable newborns. In cases of preterm pre-

labour rupture of membranes, the rupture site is the "scene of the crime" and should be fully
investigated. If *Ureaplasma* species are the leading cause of spontaneous preterm birth,
prevalence and virulence factors need to be resolved at the level of species. It will be
important to demonstrate a causal relationship between species and spontaneous labour and
membrane rupture so that antibiotics that can "cure" the individual and prevent these
outcomes are not overused.

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451 There are also new opportunities to understand placental health in situ. A particularly 452 promising development is the discovery of extracellular vesicles which are small particles 453 consisting of a lipid bilayer containing the proteins, metabolites, RNA, and DNA that have 454 budded off from a parent cell. In pregnancy, extracellular vesicles in the maternal circulation mainly come from fetal trophoblasts of the placenta.¹¹⁷ Extracellular vesicles in a peripheral 455 456 blood may reveal key aspects of the placental environment including oxygen tension, glucose 457 concentration, inflammation, and vascular dysfunction. In abnormal states such as gestational 458 diabetes and pre-eclampsia, numbers of extracellular vesicles are elevated and contain molecular signatures of these conditions.¹¹⁸ 459

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461 Every woman's journey through pregnancy and childbirth is unique and the ultimate goal 462 should be individually tailored care for all with an eye towards optimizing both mother and 463 infant health and wellbeing. Personalized antenatal care does not need to be complex or 464 expensive but the barriers may be higher in low- and middle income settings in comparison 465 with a pragmatic public health approach. Interventions can span from the bedside (e.g., better 466 gestational age assessment) to the clinic (e.g. pre-eclampsia screening) to the operating room 467 (e.g. safer anaesthesia for Caesarean sections) and to society generally (e.g. limiting tobacco 468 or pollution exposure). A more precise deployment of the existing toolkit of interventions is

- 469 likely to be more cost effective. However, many aspects of even healthy pregnancy remain
- 470 poorly understood, and it is only with continuous discovery that we move forward.

Authors' contributions

PA and NK, in collaboration with other members of the Lancet SVN steering committee, designed the study. NK and PJH verified the underlying data and PJH conducted the analyses. All authors participated in the conceptualisation and drafting of the original manuscript, reviewed and edited subsequent drafts, and approved the final version of the manuscript.

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References

- 1. Lawn JE, Ohuma EO, Bradley E et al. Small babies, big risks: Global estimates of prevalence and mortality for vulnerable newborns to accelerate change and improve counting. Lancet SVN series paper 2.
- Bryce E, Gurung S, Tong H, et al. Population attributable fractions for risk factors for spontaneous preterm births in 81 low- and middle-income countries: A systematic analysis. J Glob Health. 2022 Mar 26;12:04013. doi: 10.7189/jogh.12.04013. PMID: 35356651; PMCID: PMC8959104.
- Gurung S, Tong HH, Bryce E, et al. A systematic review on estimating population attributable fraction for risk factors for small-for-gestational-age births in 81 low- and middle-income countries. J Glob Health. 2022 Mar 26;12:04024. doi: 10.7189/jogh.12.04024. PMID: 35356650; PMCID: PMC8942297.
- Peña-Rosas JP, De-Regil LM, Garcia-Casal MN, Dowswell T. Daily oral iron supplementation during pregnancy. Cochrane Database Syst Rev. 2015 Jul 22;2015(7):CD004736. doi: 10.1002/14651858.CD004736.pub5. PMID: 26198451; PMCID: PMC8918165.
- Brocklehurst P, Gordon A, Heatley E, Milan SJ. Antibiotics for treating bacterial vaginosis in pregnancy. Cochrane Database Syst Rev. 2013 Jan 31;(1):CD000262. doi: 10.1002/14651858.CD000262.pub4. PMID: 23440777.
- Subtil D, Brabant G, Tilloy E, et al. Early clindamycin for bacterial vaginosis in pregnancy (PREMEVA): a multicentre, double-blind, randomised controlled trial. Lancet. 2018 Nov 17;392(10160):2171-2179. doi: 10.1016/S0140-6736(18)31617-9. Epub 2018 Oct 12. PMID: 30322724.
- Fleming TP, Watkins AJ, Velazquez MA, et al. Origins of lifetime health around the time of conception: causes and consequences. Lancet. 2018 May 5;391(10132):1842-1852. doi: 10.1016/S0140-6736(18)30312-X. Epub 2018 Apr 16. PMID: 29673874; PMCID: PMC5975952.
- Tobi EW, Goeman JJ, Monajemi R, et al. DNA methylation signatures link prenatal famine exposure to growth and metabolism. Nat Commun. 2014 Nov 26;5:5592. doi: 10.1038/ncomms6592. Erratum in: Nat Commun. 2015;6:7740. PMID: 25424739; PMCID: PMC4246417.
- Maltepe E, Fisher SJ. Placenta: the forgotten organ. Annu Rev Cell Dev Biol. 2015;31:523-52. doi: 10.1146/annurev-cellbio-100814-125620. Epub 2015 Oct 5. PMID: 26443191.

- Butte NF. Carbohydrate and lipid metabolism in pregnancy: normal compared with gestational diabetes mellitus. Am J Clin Nutr. 2000 May;71(5 Suppl):1256S-61S. doi: 10.1093/ajcn/71.5.1256s. PMID: 10799399.
- Tunster SJ, Watson ED, Fowden AL, Burton GJ. Placental glycogen stores and fetal growth: insights from genetic mouse models. Reproduction. 2020 Jun;159(6):R213-R235. doi: 10.1530/REP-20-0007. PMID: 32191912.
- Kaur H, Muhlhausler BS, Roberts CT, Gatford KL. The growth hormone-insulin like growth factor axis in pregnancy. J Endocrinol. 2021 Sep 1:JOE-21-0087.R1. doi: 10.1530/JOE-21-0087. PMID: 34479185.
- Giustina A, Mazziotti G, Canalis E. Growth hormone, insulin-like growth factors, and the skeleton. Endocr Rev. 2008 Aug;29(5):535-59. doi: 10.1210/er.2007-0036. Epub 2008 Apr 24. PMID: 18436706; PMCID: PMC2726838.
- Sferruzzi-Perri AN, Sandovici I, Constancia M, Fowden AL. Placental phenotype and the insulin-like growth factors: resource allocation to fetal growth. J Physiol. 2017 Aug 1;595(15):5057-5093. doi: 10.1113/JP273330. Epub 2017 May 23. PMID: 28337745; PMCID: PMC5538190.
- Fasshauer M, Blüher M, Stumvoll M. Adipokines in gestational diabetes. Lancet Diabetes Endocrinol. 2014 Jun;2(6):488-99. doi: 10.1016/S2213-8587(13)70176-1. Epub 2013 Dec 30. PMID: 24731659.
- Smits I, Hoftiezer L, van Dillen J, Hogeveen M. Neonatal hypoglycaemia and body proportionality in small for gestational age newborns: a retrospective cohort study. Eur J Pediatr. 2022 Oct;181(10):3655-3662. doi: 10.1007/s00431-022-04592-8. Epub 2022 Aug 18. PMID: 35980543; PMCID: PMC9508048.
- 17. Crane JP, Kopta MM. Comparative newborn anthropometric data in symmetric versus asymmetric intrauterine growth retardation. Am J Obstet Gynecol. 1980 Nov 1;138(5):518-22. doi: 10.1016/0002-9378(80)90279-3. PMID: 7191639.
- Mesiano S, Chan EC, Fitter JT, Kwek K, Yeo G, Smith R. Progesterone withdrawal and estrogen activation in human parturition are coordinated by progesterone receptor A expression in the myometrium. J Clin Endocrinol Metab. 2002 Jun;87(6):2924-30. doi: 10.1210/jcem.87.6.8609. PMID: 12050275.
- Myers KM, Socrate S, Paskaleva A, House M. A study of the anisotropy and tension/compression behavior of human cervical tissue. J Biomech Eng. 2010 Feb;132(2):021003. doi: 10.1115/1.3197847. PMID: 20370240.

- Amabebe E, Anumba DOC. Mechanistic Insights into Immune Suppression and Evasion in Bacterial Vaginosis. Curr Microbiol. 2022 Feb 7;79(3):84. doi: 10.1007/s00284-022-02771-2. PMID: 35128579; PMCID: PMC8818625.
- 21. Cole AM. Innate host defense of human vaginal and cervical mucosae. Curr Top Microbiol Immunol. 2006;306:199-230. PMID: 16909923.
- Hunter PJ, Sheikh S, David AL, Peebles DM, Klein N. Cervical leukocytes and spontaneous preterm birth. J Reprod Immunol. 2016 Feb;113:42-9. doi: 10.1016/j.jri.2015.11.002. Epub 2015 Nov 21. PMID: 26637953; PMCID: PMC4764650.
- 23. Condon JC, Jeyasuria P, Faust JM, Mendelson CR. Surfactant protein secreted by the maturing mouse fetal lung acts as a hormone that signals the initiation of parturition. Proc Natl Acad Sci U S A. 2004 Apr 6;101(14):4978-83. doi: 10.1073/pnas.0401124101. Epub 2004 Mar 25. PMID: 15044702; PMCID: PMC387359.
- Garcia-Verdugo I, Leiber D, Robin P, Billon-Denis E, Chaby R, Tanfin Z. Direct interaction of surfactant protein A with myometrial binding sites: signaling and modulation by bacterial lipopolysaccharide. Biol Reprod. 2007 Apr;76(4):681-91. doi: 10.1095/biolreprod.106.058131. Epub 2007 Jan 3. PMID: 17202387.
- Vannuccini S, Bocchi C, Severi FM, Challis JR, Petraglia F. Endocrinology of human parturition. Ann Endocrinol (Paris). 2016 Jun;77(2):105-13. doi: 10.1016/j.ando.2016.04.025. Epub 2016 May 5. PMID: 27155774.
- 26. Challis JR, Lye SJ, Gibb W. Prostaglandins and parturition. Ann N Y Acad Sci. 1997 Sep 26;828:254-67. doi: 10.1111/j.1749-6632.1997.tb48546.x. PMID: 9329846.
- Sennström MB, Brauner A, Byström B, Malmström A, Ekman G. Matrix metalloproteinase-8 correlates with the cervical ripening process in humans. Acta Obstet Gynecol Scand. 2003 Oct;82(10):904-11. doi: 10.1080/j.1600-0412.2003.00249.x. PMID: 12956839.
- Winkler M, Rath W. Changes in the cervical extracellular matrix during pregnancy and parturition. J Perinat Med. 1999;27(1):45-60. doi: 10.1515/JPM.1999.006. PMID: 10343934.
- Saccone G, Saccone I, Berghella V. Omega-3 long-chain polyunsaturated fatty acids and fish oil supplementation during pregnancy: which evidence? J Matern Fetal Neonatal Med. 2016;29(15):2389-97. doi: 10.3109/14767058.2015.1086742. PMID: 26382010
- 30. Simmonds LA, Sullivan TR, Skubisz M, Middleton PF, Best KP, Yelland LN, Quinlivan J, Zhou SJ, Liu G, McPhee AJ, Gibson RA, Makrides M. Omega-3 fatty acid supplementation in pregnancy-baseline omega-3 status and early preterm birth: exploratory analysis of a randomised controlled trial. BJOG. 2020

Jul;127(8):975-981. doi: 10.1111/1471-0528.16168. Epub 2020 Mar 3. PMID: 32034969.

- Middleton P, Gomersall JC, Gould JF, Shepherd E, Olsen SF, Makrides M. Omega-3 fatty acid addition during pregnancy. Cochrane Database Syst Rev. 2018 Nov 15;11(11):CD003402. doi: 10.1002/14651858.CD003402.pub3. PMID: 30480773; PMCID: PMC6516961.
- 32. Hanson MA, Bardsley A, De-Regil LM, et al. The International Federation of Gynecology and Obstetrics (FIGO) recommendations on adolescent, preconception, and maternal nutrition: "Think Nutrition First". Int J Gynaecol Obstet. 2015 Oct;131 Suppl 4:S213-53. doi: 10.1016/S0020-7292(15)30034-5. PMID: 26433230.
- Sanghavi M, Rutherford JD. Cardiovascular physiology of pregnancy. Circulation. 2014 Sep 16;130(12):1003-8. doi: 10.1161/CIRCULATIONAHA.114.009029. PMID: 25223771.
- McAuliffe F, Kametas N, Costello J, Rafferty GF, Greenough A, Nicolaides K. Respiratory function in singleton and twin pregnancy. BJOG. 2002 Jul;109(7):765-9. doi: 10.1111/j.1471-0528.2002.01515.x. PMID: 12135212.
- 35. Retief FP, Brink AJ. A study of pregnancy anaemia: blood volume changes correlated with other parameters of haemopoietic efficiency. J Obstet Gynaecol Br Commonw. 1967 Oct;74(5):683-93. doi: 10.1111/j.1471-0528.1967.tb03781.x. PMID: 6058531.
- Dockree S, Shine B, Pavord S, Impey L, Vatish M. White blood cells in pregnancy: reference intervals for before and after delivery. EBioMedicine. 2021 Dec;74:103715. doi: 10.1016/j.ebiom.2021.103715. Epub 2021 Nov 23. PMID: 34826802; PMCID: PMC8626574.
- Nakamura N, Miyazaki K, Kitano Y, Fujisaki S, Okamura H. Suppression of cytotoxic T-lymphocyte activity during human pregnancy. J Reprod Immunol. 1993 Mar;23(2):119-30. doi: 10.1016/0165-0378(93)90002-y. PMID: 8389871.
- Wald A, Van Thiel DH, Hoechstetter L, et al. Effect of pregnancy on gastrointestinal transit. Dig Dis Sci. 1982 Nov;27(11):1015-8. doi: 10.1007/BF01391748. PMID: 7140485.
- Kac G, Arnold CD, Matias SL, Mridha MK, Dewey KG. Gestational weight gain and newborn anthropometric outcomes in rural Bangladesh. Matern Child Nutr. 2019 Oct;15(4):e12816. doi: 10.1111/mcn.12816. Epub 2019 Apr 24. PMID: 30903801; PMCID: PMC6859981.
- 40. Shi H, Chen L, Wang Y, et al. Severity of Anemia During Pregnancy and Adverse Maternal and Fetal Outcomes. JAMA Netw Open. 2022 Feb 1;5(2):e2147046. doi: 10.1001/jamanetworkopen.2021.47046. PMID: 35113162; PMCID: PMC8814908.

- Keats EC, Haider BA, Tam E, Bhutta ZA. Multiple-micronutrient supplementation for women during pregnancy. Cochrane Database Syst Rev. 2019 Mar 14;3(3):CD004905. doi: 10.1002/14651858.CD004905.pub6. PMID: 30873598; PMCID: PMC6418471.
- 42. Smith ER, Shankar AH, Wu LS, et al. Modifiers of the effect of maternal multiple micronutrient supplementation on stillbirth, birth outcomes, and infant mortality: a meta-analysis of individual patient data from 17 randomised trials in low-income and middle-income countries. Lancet Glob Health. 2017 Nov;5(11):e1090-e1100. doi: 10.1016/S2214-109X(17)30371-6. PMID: 29025632.
- 43. Lassi ZS, Padhani ZA, Rabbani A, et al. Impact of Dietary Interventions during Pregnancy on Maternal, Neonatal, and Child Outcomes in Low- and Middle-Income Countries. Nutrients. 2020 Feb 19;12(2):531. doi: 10.3390/nu12020531. PMID: 32092933; PMCID: PMC7071393.
- 44. Fakhraei R, Denize K, Simon A, et al. Predictors of Adverse Pregnancy Outcomes in Pregnant Women Living with Obesity: A Systematic Review. Int J Environ Res Public Health. 2022 Feb 12;19(4):2063. doi: 10.3390/ijerph19042063. PMID: 35206265; PMCID: PMC8872310.
- 45. Han Z, Lutsiv O, Mulla S, McDonald SD; Knowledge Synthesis Group. Maternal height and the risk of preterm birth and low birth weight: a systematic review and meta-analyses. J Obstet Gynaecol Can. 2012 Aug;34(8):721-746. doi: 10.1016/S1701-2163(16)35337-3. PMID: 22947405.
- 46. Colleoni F, Lattuada D, Garretto A, et al. Maternal blood mitochondrial DNA content during normal and intrauterine growth restricted (IUGR) pregnancy. Am J Obstet Gynecol. 2010 Oct;203(4):365.e1-6. doi: 10.1016/j.ajog.2010.05.027. Epub 2010 Jul 8. PMID: 20619387.
- Burton GJ, Jauniaux E. Oxidative stress. Best Pract Res Clin Obstet Gynaecol. 2011 Jun;25(3):287-99. doi: 10.1016/j.bpobgyn.2010.10.016. PMID: 21130690; PMCID: PMC3101336.
- Moore TA, Ahmad IM, Zimmerman MC. Oxidative Stress and Preterm Birth: An Integrative Review. Biol Res Nurs. 2018 Oct;20(5):497-512. doi: 10.1177/1099800418791028. PMID: 30068228; PMCID: PMC6346316.
- 49. Stone WL, Bailey B, Khraisha N. The pathophysiology of smoking during pregnancy: a systems biology approach. Front Biosci (Elite Ed). 2014 Jun 1;6(2):318-28. doi: 10.2741/e708. PMID: 24896208.
- 50. Pawlosky RJ, Hibbeln JR, Salem N Jr. Compartmental analyses of plasma n-3 essential fatty acids among male and female smokers and nonsmokers. J Lipid

Res. 2007 Apr;48(4):935-43. doi: 10.1194/jlr.M600310-JLR200. PMID: 17234605.

- 51. Kuper SG, Abramovici AR, Jauk VC, Harper LM, Biggio JR, Tita AT. The effect of omega-3 supplementation on pregnancy outcomes by smoking status. Am J Obstet Gynecol. 2017 Oct;217(4):476.e1-476.e6. doi: 10.1016/j.ajog.2017.05.033. Epub 2017 May 23. PMID: 28549983; PMCID: PMC5614814.
- Wessels I, Maywald M, Rink L. Zinc as a Gatekeeper of Immune Function. Nutrients. 2017 Nov 25;9(12):1286. doi: 10.3390/nu9121286. PMID: 29186856; PMCID: PMC5748737.
- 53. Naber TH, van den Hamer CJ, van den Broek WJ, Roelofs H. Zinc exchange by blood cells in nearly physiologic standard conditions. Biol Trace Elem Res. 1994 Oct-Nov;46(1-2):29-50. doi: 10.1007/BF02790066. PMID: 7888283.
- 54. Turner TC, Sok MCP, Hymel LA, et al. Harnessing lipid signaling pathways to target specialized pro-angiogenic neutrophil subsets for regenerative immunotherapy. Sci Adv. 2020 Oct 30;6(44):eaba7702. doi: 10.1126/sciadv.aba7702. PMID: 33127670; PMCID: PMC7608810.
- Nadkarni S, Smith J, Sferruzzi-Perri AN, et al. Neutrophils induce proangiogenic T cells with a regulatory phenotype in pregnancy. Proc Natl Acad Sci U S A. 2016 Dec 27;113(52):E8415-E8424. doi: 10.1073/pnas.1611944114. Epub 2016 Dec 12. PMID: 27956610; PMCID: PMC5206541.
- Yamamoto H, Flannery ML, Kupriyanov S, et al. Defective trophoblast function in mice with a targeted mutation of Ets2. Genes Dev. 1998 May 1;12(9):1315-26. doi: 10.1101/gad.12.9.1315. PMID: 9573048; PMCID: PMC316781.
- Wilson RL, Leemaqz SY, Goh Z, et al. Zinc is a critical regulator of placental morphogenesis and maternal hemodynamics during pregnancy in mice. Sci Rep. 2017 Nov 9;7(1):15137. doi: 10.1038/s41598-017-15085-2. PMID: 29123159; PMCID: PMC5680205.
- Chellam VG, Rushton DI. Chorioamnionitis and funiculitis in the placentas of 200 births weighing less than 2.5 kg. Br J Obstet Gynaecol. 1985 Aug;92(8):808-14. doi: 10.1111/j.1471-0528.1985.tb03050.x. PMID: 4027203.
- Doyle RM, Harris K, Kamiza S, et al. Bacterial communities found in placental tissues are associated with severe chorioamnionitis and adverse birth outcomes. PLoS One. 2017 Jul 12;12(7):e0180167. doi: 10.1371/journal.pone.0180167. PMID: 28700642; PMCID: PMC5507499.
- Al-Adnani M, Sebire NJ. The role of perinatal pathological examination in subclinical infection in obstetrics. Best Pract Res Clin Obstet Gynaecol. 2007 Jun;21(3):505-21. doi: 10.1016/j.bpobgyn.2007.02.001. Epub 2007 Apr 19. PMID: 17448728.

- Sprong KE, Mabenge M, Wright CA, Govender S. Ureaplasma species and preterm birth: current perspectives. Crit Rev Microbiol. 2020 Mar;46(2):169-181. doi: 10.1080/1040841X.2020.1736986. Epub 2020 Mar 6. PMID: 32141797.
- Sweeney EL, Dando SJ, Kallapur SG, Knox CL. The Human Ureaplasma Species as Causative Agents of Chorioamnionitis. Clin Microbiol Rev. 2016 Dec 14;30(1):349-379. doi: 10.1128/CMR.00091-16. PMID: 27974410; PMCID: PMC5217797.
- de Goffau MC, Lager S, Sovio U, et al. Human placenta has no microbiome but can contain potential pathogens. Nature. 2019 Aug;572(7769):329-334. doi: 10.1038/s41586-019-1451-5. Epub 2019 Jul 31. Erratum in: Nature. 2019 Oct;574(7778):E15. PMID: 31367035; PMCID: PMC6697540.
- 64. Kranz A, Feierabend N, Sliwka D, Wiesegart A, Abele H, Graf J. Assessment of the Association of Periodontal Diseases in Pregnant Women and the Efficacy of Periodontal Treatment in the Context of Premature Births and Pregnancy Complications - a Narrative Review. Geburtshilfe Frauenheilkd. 2022 Aug 10;82(8):831-841. doi: 10.1055/a-1868-4693. PMID: 35967744; PMCID: PMC9365472.
- 65. Ansaldi Y, Martinez de Tejada Weber B. Urinary tract infections in pregnancy. Clin Microbiol Infect. 2022 Aug 27:S1198-743X(22)00431-1. doi: 10.1016/j.cmi.2022.08.015. Epub ahead of print. PMID: 36031053.
- 66. Gilman-Sachs A, Dambaeva S, Salazar Garcia MD, Hussein Y, Kwak-Kim J, Beaman K. Inflammation induced preterm labor and birth. J Reprod Immunol. 2018 Sep;129:53-58. doi: 10.1016/j.jri.2018.06.029. Epub 2018 Jun 30. PMID: 30025845.
- Salvi V, Vaira X, Gianello V, et al.. TLR Signalling Pathways Diverge in Their Ability to Induce PGE2. Mediators Inflamm. 2016;2016:5678046. doi: 10.1155/2016/5678046. Epub 2016 Aug 18. PMID: 27630451; PMCID: PMC5007370.
- Kyathanahalli C, Snedden M, Hirsch E. Is Human Labor at Term an Inflammatory Condition?[†]. Biol Reprod. 2022 Sep 29:ioac182. doi: 10.1093/biolre/ioac182. Epub ahead of print. PMID: 36173900.
- Umbers AJ, Boeuf P, Clapham C, et al. Placental malaria-associated inflammation disturbs the insulin-like growth factor axis of fetal growth regulation. J Infect Dis. 2011 Feb 15;203(4):561-9. doi: 10.1093/infdis/jiq080. Epub 2011 Jan 7. PMID: 21216864; PMCID: PMC3071224.
- Walters TD, Griffiths AM. Mechanisms of growth impairment in pediatric Crohn's disease. Nat Rev Gastroenterol Hepatol. 2009 Sep;6(9):513-23. doi: 10.1038/nrgastro.2009.124. PMID: 19713986.

- 71. Prendergast AJ, Rukobo S, Chasekwa B, et al. Stunting is characterized by chronic inflammation in Zimbabwean infants. PLoS One. 2014 Feb 18;9(2):e86928. doi: 10.1371/journal.pone.0086928. PMID: 24558364; PMCID: PMC3928146.
- 72. Hsiao EY, Patterson PH. Activation of the maternal immune system induces endocrine changes in the placenta via IL-6. Brain Behav Immun. 2011 May;25(4):604-15. doi: 10.1016/j.bbi.2010.12.017. Epub 2010 Dec 30. PMID: 21195166; PMCID: PMC3081363.
- 73. Keeler SM, Kiefer DG, Rust OA, et al. Comprehensive amniotic fluid cytokine profile evaluation in women with a short cervix: which cytokine(s) correlates best with outcome? Am J Obstet Gynecol. 2009 Sep;201(3):276.e1-6. doi: 10.1016/j.ajog.2009.05.045. PMID: 19733278.
- 74. Eichelberger KY, Manuck TA. Progesterone has no place in the prevention of preterm delivery: AGAINST: A call for a measured response to the OPPTIMUM trial. BJOG. 2016 Aug;123(9):1511. doi: 10.1111/1471-0528.14161. PMID: 27440593.
- 75. Prior M, Hibberd R, Asemota N, Thornton JG. Inadvertent P-hacking among trials and systematic reviews of the effect of progestogens in pregnancy? A systematic review and meta-analysis. BJOG. 2017 Jun;124(7):1008-1015. doi: 10.1111/1471-0528.14506. Epub 2017 Mar 20. PMID: 28318099.
- 76. Bone, Reproductive, and Urologic Drugs Advisory Committee. NDA 021945 Hydroxyprogesterone Caproate Injection (trade name Makena) An FDA Briefing document. October 2019.
- 77. EPPPIC Group. Evaluating Progestogens for Preventing Preterm birth International Collaborative (EPPPIC): meta-analysis of individual participant data from randomised controlled trials. Lancet. 2021 Mar 27;397(10280):1183-1194. doi: 10.1016/S0140-6736(21)00217-8. Erratum in: Lancet. 2021 Apr 17;397(10283):1446. PMID: 33773630.
- Chandiramani M, Seed PT, Orsi NM, et al. Limited relationship between cervico-vaginal fluid cytokine profiles and cervical shortening in women at high risk of spontaneous preterm birth. PLoS One. 2012;7(12):e52412. doi: 10.1371/journal.pone.0052412. Epub 2012 Dec 26. PMID: 23300664; PMCID: PMC3530581.
- 79. Tarca AL, Fitzgerald W, Chaemsaithong P, et al. The cytokine network in women with an asymptomatic short cervix and the risk of preterm delivery. Am J Reprod Immunol. 2017 Sep;78(3):e12686. doi: 10.1111/aji.12686. Epub 2017 Jun 6. PMID: 28585708; PMCID: PMC5575567.
- 80. Poon LC, Shennan A, Hyett JA, et al. The International Federation of Gynecology and Obstetrics (FIGO) initiative on pre-eclampsia: A pragmatic guide for first-trimester screening and prevention. Int J Gynaecol Obstet. 2019

May;145 Suppl 1(Suppl 1):1-33. doi: 10.1002/ijgo.12802. Erratum in: Int J Gynaecol Obstet. 2019 Sep;146(3):390-391. PMID: 31111484; PMCID: PMC6944283.

- Barros FC, Papageorghiou AT, Victora CG, et al. The distribution of clinical phenotypes of preterm birth syndrome: implications for prevention. JAMA Pediatr. 2015 Mar;169(3):220-9. doi: 10.1001/jamapediatrics.2014.3040. PMID: 25561016.
- Thilaganathan B, Kalafat E. Cardiovascular System in Preeclampsia and Beyond. Hypertension. 2019 Mar;73(3):522-531. doi: 10.1161/HYPERTENSIONAHA.118.11191. PMID: 30712425; PMCID: PMC6380450.
- Craici I, Wagner S, Garovic VD. Preeclampsia and future cardiovascular risk: formal risk factor or failed stress test? Ther Adv Cardiovasc Dis. 2008 Aug;2(4):249-59. doi: 10.1177/1753944708094227. PMID: 19124425; PMCID: PMC2674507.
- Duley L, Meher S, Hunter KE, Seidler AL, Askie LM. Antiplatelet agents for preventing pre-eclampsia and its complications. Cochrane Database Syst Rev. 2019 Oct 30;2019(10):CD004659. doi: 10.1002/14651858.CD004659.pub3. PMID: 31684684; PMCID: PMC6820858
- 85. Smith DD, Costantine MM. The role of statins in the prevention of preeclampsia. Am J Obstet Gynecol. 2022 Feb;226(2S):S1171-S1181. doi: 10.1016/j.ajog.2020.08.040. Epub 2020 Aug 17. PMID: 32818477; PMCID: PMC8237152.
- Hofmeyr GJ, Lawrie TA, Atallah ÁN, Torloni MR. Calcium supplementation during pregnancy for preventing hypertensive disorders and related problems. Cochrane Database Syst Rev. 2018 Oct 1;10(10):CD001059. doi: 10.1002/14651858.CD001059.pub5. PMID: 30277579; PMCID: PMC6517256.
- Masotti G, Galanti G, Poggesi L, Abbate R, Neri Serneri GG. Differential inhibition of prostacyclin production and platelet aggregation by aspirin. Lancet. 1979 Dec 8;2(8154):1213-7. doi: 10.1016/s0140-6736(79)92334-1. PMID: 92623.
- Rolnik DL, Nicolaides KH, Poon LC. Prevention of preeclampsia with aspirin. Am J Obstet Gynecol. 2022 Feb;226(2S):S1108-S1119. doi: 10.1016/j.ajog.2020.08.045. Epub 2020 Aug 21. PMID: 32835720.
- 89. Stewart CP, Oaks BM, Laugero KD, et al. Maternal cortisol and stress are associated with birth outcomes, but are not affected by lipid-based nutrient supplements during pregnancy: an analysis of data from a randomized controlled trial in rural Malawi. BMC Pregnancy Childbirth. 2015 Dec 22;15:346. doi: 10.1186/s12884-015-0793-8. PMID: 26694646; PMCID: PMC4688934.

- 90. Shapiro GD, Fraser WD, Frasch MG, Séguin JR. Psychosocial stress in pregnancy and preterm birth: associations and mechanisms. J Perinat Med. 2013 Nov;41(6):631-45. doi: 10.1515/jpm-2012-0295. PMID: 24216160; PMCID: PMC5179252.
- Christian LM, Glaser R, Porter K, Iams JD. Stress-induced inflammatory responses in women: effects of race and pregnancy. Psychosom Med. 2013 Sep;75(7):658-69. doi: 10.1097/PSY.0b013e31829bbc89. Epub 2013 Jul 19. PMID: 23873713; PMCID: PMC3788648.
- 92. Coussons-Read ME, Okun ML, Schmitt MP, Giese S. Prenatal stress alters cytokine levels in a manner that may endanger human pregnancy. Psychosom Med. 2005 Jul-Aug;67(4):625-31. doi: 10.1097/01.psy.0000170331.74960.ad. PMID: 16046378.
- 93. Coussons-Read ME, Okun ML, Nettles CD. Psychosocial stress increases inflammatory markers and alters cytokine production across pregnancy. Brain Behav Immun. 2007 Mar;21(3):343-50. doi: 10.1016/j.bbi.2006.08.006. Epub 2006 Oct 6. PMID: 17029703.
- 94. Zdravkovic T, Genbacev O, McMaster MT, Fisher SJ. The adverse effects of maternal smoking on the human placenta: a review. Placenta. 2005 Apr;26 Suppl A:S81-6. doi: 10.1016/j.placenta.2005.02.003. PMID: 15837073.
- 95. Siddiqui AR, Gold EB, Yang X, Lee K, Brown KH, Bhutta ZA. Prenatal exposure to wood fuel smoke and low birth weight. Environ Health Perspect. 2008 Apr;116(4):543-9. doi: 10.1289/ehp.10782. PMID: 18414641; PMCID: PMC2290983.
- 96. Moore LG. Fetal growth restriction and maternal oxygen transport during high altitude pregnancy. High Alt Med Biol. 2003 Summer;4(2):141-56. doi: 10.1089/152702903322022767. PMID: 12855048.
- Chamberlain C, O'Mara-Eves A, Porter J, et al. Psychosocial interventions for supporting women to stop smoking in pregnancy. Cochrane Database Syst Rev. 2017 Feb 14;2(2):CD001055. doi: 10.1002/14651858.CD001055.pub5. PMID: 28196405; PMCID: PMC6472671.
- 98. Kabir Z, Clarke V, Conroy R, McNamee E, Daly S, Clancy L. Low birthweight and preterm birth rates 1 year before and after the Irish workplace smoking ban. BJOG. 2009 Dec;116(13):1782-7. doi: 10.1111/j.1471-0528.2009.02374.x.
- 99. Faber T, Kumar A, Mackenbach JP, Millett C, Basu S, Sheikh A, Been JV. Effect of tobacco control policies on perinatal and child health: a systematic review and meta-analysis. Lancet Public Health. 2017 Sep 5;2(9):e420-e437. doi: 10.1016/S2468-2667(17)30144-5. PMID: 28944313; PMCID: PMC5592249.
- 100. Díez-Izquierdo A, Balaguer A, Lidón-Moyano C, et al. Correlation between tobacco control policies and preterm births and low birth weight in Europe.

Environ Res. 2018 Jan;160:547-553. doi: 10.1016/j.envres.2017.10.033. Epub 2017 Oct 28. PMID: 29089104.

- 101. World Health Organization Regional Office for Europe. Air quality guidelines: global update 2005: particulate matter, ozone, nitrogen dioxide and sulfur dioxide. World Health Organization. Regional Office for Europe; 2006.
- 102. Austin KF, Mejia MT. Household air pollution as a silent killer: women's status and solid fuel use in developing nations. Popul Environ. 2017 Sep;39(1):1–25.
- 103. Katz J, Tielsch JM, Khatry SK, et al. Impact of Improved Biomass and Liquid Petroleum Gas Stoves on Birth Outcomes in Rural Nepal: Results of 2 Randomized Trials. Glob Health Sci Pract. 2020 Sep 30;8(3):372–82.
- 104. Clasen TF, Chang HH, Thompson LM, et al. Liquefied Petroleum Gas or Biomass for Cooking and Effects on Birth Weight. N Engl J Med. 2022 Nov 10;387(19):1735-1746. doi: 10.1056/NEJMoa2206734. Epub 2022 Oct 10. PMID: 36214599.
- 105. Kuehn L, McCormick S. Heat Exposure and Maternal Health in the Face of Climate Change. Int J Environ Res Public Health. 2017 Jul 29;14(8):853. doi: 10.3390/ijerph14080853. PMID: 28758917; PMCID: PMC5580557.
- 106. Chersich MF, Pham MD, Areal A, et al. Associations between high temperatures in pregnancy and risk of preterm birth, low birth weight, and stillbirths: systematic review and meta-analysis. BMJ. 2020 Nov 4;371:m3811. doi: 10.1136/bmj.m3811. PMID: 33148618; PMCID: PMC7610201.
- 107. Wang J, Tong S, Williams G, Pan X. Exposure to Heat Wave During Pregnancy and Adverse Birth Outcomes: An Exploration of Susceptible Windows. Epidemiology. 2019 Jul;30 Suppl 1:S115-S121. doi: 10.1097/EDE.000000000000995. PMID: 31181014.
- 108. Olivier K, Reinders LA, Clarke MW, Crew RC, Pereira G, Maloney SK, Wyrwoll CS. Maternal, Placental, and Fetal Responses to Intermittent Heat Exposure During Late Gestation in Mice. Reprod Sci. 2021 Feb;28(2):416-425. doi: 10.1007/s43032-020-00291-7. Epub 2020 Aug 17. PMID: 32804351.
- 109. Ghulam Mohyuddin S, Khan I, Zada A, et al. Influence of Heat Stress on Intestinal Epithelial Barrier Function, Tight Junction Protein, and Immune and Reproductive Physiology. Biomed Res Int. 2022 Sep 1;2022:8547379. doi: 10.1155/2022/8547379. PMID: 36093404; PMCID: PMC9458360.
- 110. Wan J, Chen B, Rao J. Occurrence and preventive strategies to control mycotoxins in cereal-based food. Compr Rev Food Sci Food Saf. 2020

May;19(3):928-953. doi: 10.1111/1541-4337.12546. Epub 2020 Mar 4. PMID: 33331688.

- 111. da Silva JVB, de Oliveira CAF, Ramalho LNZ. Effects of Prenatal Exposure to Aflatoxin B1: A Review. Molecules. 2021 Dec 2;26(23):7312. doi: 10.3390/molecules26237312. PMID: 34885894; PMCID: PMC8659025.
- 112. Gore AC, Chappell VA, Fenton SE, Flaws JA, Nadal A, Prins GS, Toppari J, Zoeller RT. Executive Summary to EDC-2: The Endocrine Society's Second Scientific Statement on Endocrine-Disrupting Chemicals. Endocr Rev. 2015 Dec;36(6):593-602. doi: 10.1210/er.2015-1093. Epub 2015 Sep 28. PMID: 26414233; PMCID: PMC4702495.
- 113. Wu Y, Wang J, Wei Y, et al. Maternal exposure to endocrine disrupting chemicals (EDCs) and preterm birth: A systematic review, meta-analysis, and meta-regression analysis. Environ Pollut. 2022 Jan 1;292(Pt A):118264. doi: 10.1016/j.envpol.2021.118264. Epub 2021 Oct 1. PMID: 34606968.
- 114. Hofmeyr GJ, Black RE, Rogozińska E, et al. Evidence-based antenatal interventions to reduce the incidence of small vulnerable newborns and their associated poor outcomes. Lancet SVN series paper 4.
- 115. Shynlova O, Nadeem L, Dorogin A, Mesiano S, Lye SJ. The selective progesterone receptor modulator-promegestone-delays term parturition and prevents systemic inflammation-mediated preterm birth in mice. Am J Obstet Gynecol. 2022 Feb;226(2):249.e1-249.e21. doi: 10.1016/j.ajog.2021.08.013. Epub 2021 Aug 19. PMID: 34418351; PMCID: PMC8810746.
- 116. Peeling RW, Mabey D, Herring A, Hook EW 3rd. Why do we need qualityassured diagnostic tests for sexually transmitted infections? Nat Rev Microbiol. 2006 Dec;4(12):909-21. doi: 10.1038/nrmicro1555. PMID: 17109030.
- 117. Nakahara A, Nair S, Ormazabal V, et al. Circulating Placental Extracellular Vesicles and Their Potential Roles During Pregnancy. Ochsner J. 2020 Winter;20(4):439-445. doi: 10.31486/toj.20.0049. PMID: 33408584
- 118. Awoyemi T, Tannetta D, Zhang W, et al. Glycosylated Siglec-6 expression in syncytiotrophoblast-derived extracellular vesicles from preeclampsia placentas. Biochem Biophys Res Commun. 2020 Dec 17;533(4):838-844. doi: 10.1016/j.bbrc.2020.09.081. PMID: 32998819.

Organ system	Change	
Heart	Cardiac output increases by 50%. ³³	
Lungs	Ventilation (volume/minute) increases by 50%. ³⁴	
Vasculature	Vascular resistance decreases by $30 - 50\%$. ³³	
Red blood cells	Early 10% decrease in RBC and hemoglobin per volume	
	due to increase in plasma volume. ³⁵	
White blood cells	Circulating neutrophil counts increase by 50%. ³⁶	
	T cells become less responsive to antigenic stimulation. ³⁷	
Gastro-intestinal tract	Transit time slows down, possibly to allow longer time	
	for absorption of nutrients. ³⁸	
Pancreas	Small increase in insulin production in response to mild	
	insulin resistance in maternal tissues. ¹⁰	

Table 1. Changes to organ systems in women during pregnancy.

Maternal nutritional factor	Potential mechanistic pathways	Outcomes
Nutrient supply (energy and	Energy and nutrient delivery to the	Growth
macronutrients:	placenta and fetus. ⁴³	restriction
carbohydrates, proteins,		
lipids)		
Body composition	Underweight or low GWG: low energy	Growth
(underweight, overweight);	supply. ³²	restriction
gestational weight gain	Overweight or excess GWG: metabolic	
(GWG)	and hormonal dysregulation, gestational	
	diabetes, hypertension, inflammation. ⁴⁴	
Dietary quality	Metabolic and hormonal dysregulation,	Growth
	gestational diabetes, hypertension,	restriction,
	inflammation, oxidative stress.	preterm birth
Stature	Small "container effect" on uterine and	Growth
	placental size. ⁴⁵	restriction
Micronutrients related to	Oxygen supply to placenta and fetus.	Growth
cardiac function, anaemia		restriction,
and oxygen supply (e.g.,		preterm birth
iron, riboflavin, folic acid,		
vitamin B12, vitamin C)		
Nutrients that support	Ability to fight infection and control	Fetal growth
immune function (e.g., zinc,	inflammation.	restriction,
fatty acids, vitamin D, iron)		preterm birth
Antioxidants and cofactors	Ability to reduce and repair damage	Fetal growth
of antioxidant enzymes (e.g.,	caused by oxidative stress.	restriction,
vitamins C, E, carotenoids,		preterm birth
copper, zinc, fatty acids)		
Nutrients related to cortisol	Control of inflammation, prevention of	Fetal growth
metabolism (e.g., fatty acids,	preterm COX2 activation and	restriction,
zinc, magnesium)	prostaglandin production.	preterm birth
Nutrients related to	Mitochondrial efficiency and protection	Fetal growth
mitochondrial function (e.g.,	against oxidative stress. ⁴⁶	restriction
vitamins C and E, zinc,		
copper, iodine, selenium)		
Nutrients related to	Omega-3 fatty acids: competitive	Preterm birth
production of prostaglandins	inhibition of preterm production of	
(e.g., long chain poly-	prostaglandins E2 and F2 α from	
unsaturated fatty acids)	arachidonic acid. ²⁹	

Table 2. Nutritional factors related to the small vulnerable newborn

Undernourishment	Infection	Characteristics of woman	Environmental exposures
Anaemia	HIV	and pregnancy	and psychosocial stress
Zinc deficiency	Malaria	First pregnancy	Unwanted pregnancy
Calcium deficiency	Syphilis	Adolescent pregnancy	Intimate partner abuse
hort stature	Chlamydia	Short interpregnancy interval	Lack of support or agency
ow BMI	Gonorrhoea	Extreme parity	Mental illness
Inadequate weight gain Bacterial vaginosis Trichomonas vaginalis Group B Streptococcus	Urinary tract infection	Older age	Smoking
	Bacterial vaginosis	Preeclampsia	Alcohol abuse
	Placental dysfunction	Drug abuse	
	Group B Streptococcus	Gestational diabetes	Toxins
		Hypothyroidism	Endocrine disruptors
		Cervical weakness	Indoor air pollution
		Uterine malformations	Outdoor air pollution
		Endometriosis	Heat waves
		Multiple pregnancy	High altitude

Panel 1. Risk factors for the birth of a small vulnerable newborn



Figure 1. Developing fetus and fully developed placenta. The basic body plan with rudimentary organs are in place by 5 weeks post fertilization. The umbilical artery carries deoxygenated, waste-replete, nutrient-depleted fetal blood to the placental villi where waste is exchanged for nutrients and carbon dioxide is exchanged for oxygen from maternal blood.



Figure 2. Conceptual model of key determinants of gestational length. When the fetus is ready to be born, cortisol enters the placenta and circulation and activates cyclooxygenase-2 to generate prostaglandin E2, which directs cervical and uterine remodelling. Estrogens override the suppressive effects of progesterone and oxytocins trigger uterine contractions. CRH – corticotropin releasing hormone.



Figure 3. Conceptual model of key determinants of fetal growth. Hormones, nutrients and oxygen from the mother are taken up by the placenta and transferred to the fetal circulation to support synthesis of fetal tissue.



- 3 Figure 4. Examples of exposures that are able to contribute to both preterm birth and growth restriction via different pathways and ways to
- 4 intervene toward prevention. Zinc deficiency (A), psychological and physical stress (B) and poor air quality/tobacco smoke (C) contribute to the
- 5 birth of a small vulnerable newborn. MMS multiple micronutrient supplements, COX2 cyclooxygenase 2, LCPUFA long chain
- 6 polyunsaturated fatty acids, IGF insulin-like growth factor, ROS reactive oxygen species.





8

10 Figure 5. Immune defence of the cervix. The cervix remains long and closed for the duration

11 of pregnancy. It is defended by antimicrobial chemicals including peptides, antibodies and

12 enzymes. Neutrophils are also present in the mucus and are able to destroy invading

13 microbes. In the absence of adequate immune defence, bacteria are able to colonize and

14 damage the membranes leading to rupture or chorioamnionitis.

15