



RESEARCH ARTICLE

Profiles of inflammatory markers and their association with cardiometabolic parameters in rural and urban Uganda

[version 1; peer review: 1 not approved]

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Abstract

Background: Inflammation may be one of the pathways explaining differences in cardiometabolic risk between urban and rural residents. We investigated associations of inflammatory markers with rural versus urban residence, and with selected cardiometabolic parameters previously observed to differ between rural and urban residents: homeostatic model assessment of insulin resistance (HOMA-IR), fasting blood glucose (FBG), blood pressure (BP) and body mass index (BMI).

Methods: From two community surveys conducted in Uganda, 313 healthy individuals aged ≥ 10 years were selected by age- and sex-stratified random sampling (rural Lake Victoria island communities, 212; urban Entebbe municipality, 101). Fluorescence intensities of plasma cytokines and chemokines were measured using a bead-based multiplex immunoassay. We used linear regression to examine associations between the analytes and rural-urban residence and principal component analysis (PCA) to further investigate patterns in the relationships. Correlations between analytes and metabolic parameters were assessed using Pearson's correlation coefficient.

Results: The urban setting had higher mean levels of IL-5 (3.27 vs 3.14, adjusted mean difference [95% confidence interval] 0.12[0.01,0.23] $p=0.04$), IFN- γ (26.80 vs 20.52, 6.30[2.18,10.41] $p=0.003$), EGF (5.67 vs 5.07, 0.60[0.32,0.98] $p<0.00001$), VEGF (3.68 vs 3.28, 0.40[0.25,0.56] $p<0.00001$), CD40 Ligand (4.82 vs 4.51, 0.31[0.12,

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report

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0.50] $p=0.001$) and Serpin-E1 (9.57 vs 9.46, 0.11[0.05,0.17] $p<0.00001$), but lower levels of GMCSF (2.94 vs 3.05, -0.10[-0.19,-0.02] $p=0.02$), CCL2 (2.82 vs 3.10, -0.45[-0.70,-0.21] $p<0.00001$) and CXCL10 (5.48 vs 5.96, -0.49[-0.71,-0.27] $p<0.00001$), compared to the rural setting. In PCA, the urban setting had lower representation of some classical inflammatory mediators but higher representation of various chemoattractants and vasoactive peptides. HOMA-IR, FBG, BP and BMI were positively correlated with several principal components characterised by pro-inflammatory analytes.

Conclusions: In developing countries, immunological profiles differ between rural and urban environments. Differential expression of certain pro-inflammatory mediators may have important health consequences including contributing to increased cardiometabolic risk observed in the urban environment.

Keywords

Rural, Urban, Inflammation, Cytokines, Immunoassay, Hypertension, Diabetes, Cardiometabolic, Africa

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Introduction

Inflammation is a necessary component of the immune system response to injury and infection. However, chronic, low-grade inflammation has been implicated in the development and progression of cardiometabolic disorders such as type 2 diabetes¹ and atherosclerosis². The burden of these diseases is increasing globally with cardiovascular diseases being the top contributors to global mortality and morbidity^{3,4}. This increase has been attributed to changes in lifestyle and diet as a result of urbanisation and the burden is heavier among individuals in urban areas. Currently, more than 50% of the world's population lives in urban areas and this is expected to increase⁵. Environmental exposures such as microbial exposure, helminth infections, diet and pollution influence the inflammatory status of individuals. This may lead to differences in immune status between individuals in rural and urban settings⁶. Therefore, inflammation may be one of the pathways explaining the disparity in cardiometabolic disease burden and risk between rural and urban areas.

Few studies have investigated, in the general population, rural-urban differences in inflammatory markers. A study in Senegal showed that urban inhabitants had lower pro-inflammatory and T-cell activation profiles than rural inhabitants⁶. Another study in Morocco investigating gene expression in blood leukocyte samples showed differences in the leukocyte transcriptome among rural, urban and nomadic populations⁷. Other studies have focused on particular disease states and the results are mixed and inconsistent. For example, compared with rural upbringing, urban upbringing has been associated with higher concentrations of the pro-inflammatory cytokine interleukin (IL)-6 and suppressed concentrations of the anti-inflammatory cytokine IL-10 following acute social stress⁸. A study investigating the effect of exposure to a forest environment on stress levels and cytokines showed that, compared with two hours of forest exposure, urban exposure for two hours resulted in higher levels of the chemokine IL-8 and the pro-inflammatory cytokine tumour necrosis factor (TNF)-alpha⁹. In the investigation of diabetes risk in India, urban dwellers had higher levels of TNF-alpha, IL-6 and the adipokine leptin than rural dwellers¹⁰. A study conducted in Ghana among individuals with type 2 diabetes showed higher levels of TNF-alpha but lower levels of IL-6 among urban participants than in rural participants¹¹. Conversely, plasma levels of the chemoattractant C-C motif chemokine (CCL) 11 have been shown to be lower in urban, compared to rural dwellers in a study investigating the effect of CCL on cognitive function¹². Among individuals with atopy, urban inhabitants had higher levels of gene expression for IL-4R, a type 2 cytokine receptor, compared with rural inhabitants¹³. In another study in Ghana, urban children had increased gene expression of IL-10 compared with rural children, irrespective of helminth infection status¹⁴.

Different environments exert different influences on the immune status of the inhabitants. To our knowledge, no study has investigated rural-urban differences in inflammatory markers in Uganda or explored the implications of rural-urban differences for cardiometabolic risk. We, therefore, exploited a unique

opportunity to measure inflammatory markers from individuals living in a rural and urban setting. We investigated rural-urban differences in inflammatory cytokines and chemokines and examined correlations between the inflammatory markers and the metabolic parameters HOMA-IR, fasting blood glucose, blood pressure and body mass index (BMI), which we had found to be different between the urban and rural settings¹⁵.

Methods

Study design and setting

This was a cross-sectional comparative study. We measured pro- and anti-inflammatory cytokines and chemokines in stored plasma of individuals who participated in two cross-sectional surveys conducted in rural and urban Uganda. The rural survey was conducted from April to December 2017 in 26 island fishing communities in Lake Victoria, Koome sub-county, Mukono district as part of the Lake Victoria Island Intervention Study on Worms and Allergy-related related diseases (LaVI-ISWA). These fishing villages are located on the shores of the islands and have a high burden of schistosomiasis. LaVI-ISWA was a cluster-randomised trial designed to investigate the effects of community-wide intensive (quarterly single-dose praziquantel and triple-dose albendazole) versus standard (annual single-dose praziquantel and six-monthly single-dose albendazole) anthelmintic intervention on health outcomes^{16,17}. The LaVIISWA protocol has been published¹⁸. This survey, the rural metabolic survey, was conducted in participants aged ≥ 10 years after four years of the anthelmintic intervention to investigate its effects on metabolic outcomes¹⁶. The urban survey was conducted between September 2016 and September 2017 in 23 sub-wards of Entebbe municipality, Wakiso district, Uganda. Entebbe is located on a peninsula on the northern shore of Lake Victoria. The urban survey was a cross-sectional study that collected data on allergy-related and metabolic outcomes¹⁹. In both surveys, data were collected on sociodemographic characteristics (obtained using a questionnaire), fasting plasma glucose, fasting lipid profile (triglycerides, total cholesterol, high-density lipoprotein [HDL]-cholesterol and low-density lipoprotein [LDL]-cholesterol), blood pressure, body mass index, waist and hip circumference. The field tools and procedures in both surveys were aligned to enable a comparison of the outcomes between the two surveys.

Participants and samples

We used plasma samples from 313 healthy participants, 212 from the rural setting (104 from the intensive arm and 108 from the standard arm of the LaVIISWA trial) and 101 from the urban setting. These were randomly selected from each survey using *STATA* software (*STATA* v13, College Station, Texas, USA). No formal sample size calculation was performed because this work was exploratory. Participants with incomplete data on helminth infection status, blood pressure, glucose, insulin, lipid profile, BMI, waist and hip circumference or did not have stored plasma samples were excluded from the sampling frame. The selection was stratified by age (10–19 years, 20–29 years, 30–39 years and 40 years) and sex. This was to ensure that the age and sex distribution of participants whose samples were used was similar between the two settings because

age and sex might be predictive of inflammatory marker levels. Stored plasma samples were retrieved and cytokines/chemokines measured using a bead-based multiplex immunoassay (*Luminex*).

Measurement of inflammatory markers

We measured the fluorescence intensity and concentrations of 30 analytes in plasma. Commercially available kits which included chemokines, Th1 and Th2 cytokines, and adipokines were selected. The analytes measured were adiponectin/Acrp30, serpin E1/PAI1, chemokine (C-C motif) ligand (CCL)2/JE/monocyte chemoattractant protein (MCP)1, CCL3/MIP1-alpha, CCL4/MIP1-beta, CCL11/Eotaxin, CCL22/MDC, CD40 Ligand/TNFSF5, chemokine (C-X-C motif) ligand (CXCL)1/GRO-alpha/KC, CXCL2/Gro-beta/MIP2/CINC3, CXCL10/interferon-gamma induced protein (IP)10/CRG2, epidermal growth factor (EGF), Fas, granulocyte-macrophage colony-stimulating factor (GM-CSF), interferon (IFN)-alpha, IFN-gamma, Interleukin (IL) 1-alpha, IL-1beta/IL-1F2, IL-2, IL-4, IL-5, IL-6, IL-8/CXCL8, IL-10, IL-12 p70, IL-13, IL-17/IL-17A, leptin, tumour necrosis factor (TNF)-alpha and vascular endothelial growth factor (VEGF)-C.

Plasma samples, stored in -80 degrees Celsius refrigerators, were thawed to room temperature and tested. The multiplex assays were performed using the Human Premixed Multi-analyte Magnetic Luminex Assay kit (R and D Systems Inc, Minneapolis, Minnesota, USA; catalogue numbers, LXSAHM-01, LXSAHM-02, LXSAHM-27; Lot numbers L128094-6) according to the manufacturer's instructions. The thirty analytes were assessed using three panels, a one-plex assay for VEGF, a two-plex assay for adiponectin and serpin E1 and a 27-plex assay for the other analytes. In brief, all reagents were prepared as instructed by the kit manufacturer. To each well, 50ul of sample (or standard) were added followed by 50ul of diluted Microparticle Cocktail. The mixture was incubated on a shaker for two hours and washed with wash buffer three times. Diluted Biotin-Antibody Cocktail was added (50ul to each well), incubated on the shaker and washed with wash buffer three times. Thereafter, diluted Streptavidin-PE was added (50ul to each well), incubated on the shaker for 30 minutes and again washed three times. Wash buffer was then added to each well and incubated for two minutes on a shaker. All incubations were at room temperature and each time, the shaker was rotating at 800 rpm.

The results were read using the Bio-Plex 200 System (Bio-Rad Laboratories, Hercules, California, USA) and the [Bio-Plex Manager software](#) (version 6.0; Bio-Rad Laboratories, Hercules, California, USA) after imputing the assay working range (lower and upper limits of quantification) for each analyte according to the manufacturer's instructions. Raw fluorescence intensity values and observed concentrations were obtained.

Statistical methods

The analyses were performed using STATA version 13.0 (College Station, Texas). The 30 cytokines/chemokines were the outcomes of interest in this analysis. The exposures of interest were the rural-urban setting and the metabolic parameters homeostatic model assessment of insulin resistance (HOMA-IR),

fasting plasma glucose, blood pressure and body mass index (BMI). In all our analyses, we used the raw fluorescence intensity values of the cytokines. This is because the analytes were predominantly low in abundance and the observed concentrations were mostly extrapolated beyond the standard range. Using the observed concentrations would require censoring i.e. not assigning values for concentrations that are out of the standard range. This would affect most of the cytokines measured and therefore limit the statistical power of the analysis. Analysis using fluorescence intensities does not require censoring and provides more statistical power than analysis based on concentrations. It also does not require correction for background fluorescence because the automatic calibration steps produce a highly reproducible signal even across different Luminex machines. This approach for analysing Luminex data has been validated^{20,21} and used previously^{22,23}.

The fluorescence intensities of cytokines were compared between the rural and urban setting. All analyte intensities were first log-transformed. The mean log intensity of each analyte in each setting was then calculated and regression models fitted to assess for differences. Crude and adjusted mean differences (adjusted for age and sex) and their 95% confidence intervals are presented. Further analyses investigated correlation patterns between the outcomes and then used principal component analysis to transform from a set of correlated outcome variables to a set of uncorrelated variables (linear combinations of the original outcomes) aiming to capture as much variability as possible with the first few components. The newly generated uncorrelated variables were then compared between the settings. Selection of the number of principal components to retain was based on the Kayser criterion, therefore, components with an eigenvalue of ≥ 1 were retained. A scree plot was constructed to give a visual representation of the PCA eigenvalues. In the interpretation of the principal components, we computed the unrotated matrix containing the factor loadings of each variable for each component. For each variable, the principal component with the highest loading for that variable was interpreted as representative of the variable (adapted from 24). In all the analyses, data from the rural setting included all rural participants regardless of LaVIISWA trial arm. This is because no strong differences in the cytokines were found between the study arms.

To determine the relationship between cytokines/chemokines and metabolic parameters we computed Pearson correlation coefficients. This is to investigate the strength and direction of any linear relationship. We obtained coefficients for the correlation of the log-transformed intensity for each analyte with HOMA-IR, fasting plasma glucose, blood pressure (systolic and diastolic) and BMI. We also correlated the metabolic outcomes with the retained components from the PCA. Correlation coefficients and their p-values are reported. No formal adjustment was made for multiple testing.

Ethical approvals and consent

Ethical clearance for this work was obtained from the Uganda Virus Research Institute Research Ethics Committee (reference number GC/127/17/01/573), the Uganda National Council

for Science and Technology (reference number HS 2185) and the London School of Hygiene and Tropical Medicine Research Ethics Committee (reference number 9917). All participants provided written informed consent to participate in the surveys and have their samples stored for this work (see consent forms and information sheets in the extended data²⁵).

Results

Characteristics of the participants

The characteristics of the participants selected for this analysis are shown in Table 1²⁵. Samples from 313 participants (rural, 212; urban, 101) were analysed. The participants had a median age of 30 years and were selected to have comparable distributions of age and sex. Participants in the urban setting had a lower prevalence of the helminths *Schistosoma mansoni* (28.7% vs 53.6%) and *Trichuris trichiura* (1.0% vs 9.5%) than those in the rural setting.

Associations between the analytes and study setting

The associations (both unadjusted and adjusted) between rural-urban setting and level of each analyte are shown in Table 2. Somewhat higher levels of the Th2 associated cytokine IL-5 were observed in the urban setting on adjusted analysis (3.27 vs 3.14, adjusted mean difference [95% confidence interval] 0.12 [0.01, 0.23] $p=0.04$). However, no differences were observed for the other Th2 associated cytokines (IL-4 and IL-13), Th1 associated cytokines (IL-2, IFN-gamma and TNF-alpha), Th17 associated cytokines (IL-17) or the regulatory cytokines (IL-10, TGF-beta) between the rural and urban settings. Furthermore, no differences were observed for the adipokines adiponectin and leptin.

Comparison of chemokines, growth factors and other cytokines between the rural and urban setting showed interesting differences. Individuals in the urban setting had higher levels of

Table 1. Characteristics of the study participants.

	Urban setting	Rural setting		
		Overall	Standard arm	Intensive arm
Sex, male	50/101 (49.5%)	108/212 (50.9%)	55/108 (50.9%)	53/104 (51.0%)
Age (median, IQR)	30 (20.0, 39.0)	30 (20.0, 38.5)	30 (20, 40)	29 (20, 37.5)
Age group				
10–19yrs	25/101 (24.8%)	51/212 (24.1%)	26/108 (24.1%)	25/104 (24.0%)
20–29yrs	25/101 (24.8%)	54/212 (25.5%)	26/108 (24.1%)	28/104 (25.9%)
30–39yrs	26/101 (25.7%)	55/212 (25.9%)	28/108 (25.9%)	27/104 (26.0%)
40+	25/101 (24.8%)	52/212 (24.5%)	28/108 (25.9%)	24/104 (23.1%)
<i>Schistosoma mansoni</i> , stool Kato Katz*	10/101 (9.9%)	73/211 (34.6%)	35/107 (32.7%)	38/104 (36.5%)
<i>Schistosoma mansoni</i> , stool PCR*	29/101 (28.7%)	113/211 (53.6%)	61/107 (57.0%)	52/104 (50.0%)
Hookworm, stool Kato Katz*	2/101 (2.0%)	6/211 (2.8%)	3/107 (2.8%)	3/104 (2.9%)
Hookworm, stool PCR*	2/101 (2.0%)	8/194 (4.1%)	4/104 (3.9%)	4/90 (4.4%)
<i>Trichuris trichiura</i> , stool Kato Katz*	1/101 (1.0%)	20/211 (9.5%)	15/107 (14.0%)	5/104 (4.8%)
<i>Ascaris lumbricoides</i> , stool Kato Katz*	0	2/211 (1.0%)	2/107 (1.9%)	0
<i>Mansonella perstans</i> , modified Knott's*	2/101 (2.0%)	6/211 (2.8%)	2/107 (1.9%)	4/104 (3.9%)
<i>Strongyloides stercoralis</i> , stool PCR*	4/101 (4.0%)	14/211 (6.6%)	8/107 (7.5%)	6/104 (5.8%)
HOMA-IR (mean, SD)**	1.72 (2.0)	1.77 (1.4)	1.65 (1.2)	1.89 (1.6)
Fasting blood glucose mmol/L (mean, SD)**	4.64 (0.8)	4.79 (0.7)	4.75 (0.8)	4.83 (0.6)
Systolic blood pressure, mmHg (mean, SD)**	117.29 (13.0)	114.48 (14.9)	113.48 (16.3)	115.51 (13.3)
Diastolic blood pressure, mmHg (mean, SD)**	76.56 (9.9)	75.46 (10.5)	74.90 (10.6)	76.04 (10.4)
Body mass index, kg/m ² (mean, SD)**	23.81 (5.7)	22.89 (4.2)	22.43 (3.9)	23.36 (4.4)

IQR, Interquartile range; SD, Standard deviation; PCR, Polymerase chain reaction; HOMA-IR, Homeostatic model assessment of insulin resistance;

*Results of chi-squared tests for whether participant characteristics differed between urban and rural setting were: *Schistosoma mansoni* stool Kato Katz ($p<0.001$); *Schistosoma mansoni* stool PCR ($p<0.001$), Hookworm stool Kato Katz ($p=0.652$), Hookworm stool PCR ($p=0.334$), *Trichuris trichiura* ($p=0.005$), *Ascaris lumbricoides* ($p=0.326$), *Mansonella perstans* ($p=0.652$) *Strongyloides stercoralis* ($p=0.343$); **Results of two-sample t -tests for whether participant characteristics differed between urban and rural setting were: HOMA-IR ($p=0.815$), Fasting blood glucose ($p=0.084$), Systolic blood pressure ($p=0.105$), Diastolic blood pressure ($p=0.380$), Body mass index ($p=0.108$).

Table 2. Associations between inflammatory analyte intensities and study setting.

Analytes	Mean log fluorescence intensity*		Mean difference (95% CI)			
	Rural	Urban	Unadjusted	P value	Adjusted for age and sex	P value
IL1-alpha	2.91	2.88	-0.03 (-0.10, 0.03)	0.32	-0.03 (-0.10, 0.03)	0.33
IL1-beta	2.52	2.58	0.05 (-0.07, 0.18)	0.39	0.05 (-0.07, 0.18)	0.40
IL-2	2.98	2.97	-0.01 (-0.08, 0.06)	0.76	-0.01 (-0.08, 0.06)	0.76
IL-4	2.92	2.93	0.00 (-0.04, 0.05)	0.91	0.00 (-0.04, 0.05)	0.89
IL-5	3.14	3.27	0.13 (-0.01, 0.26)	0.06	0.12 (0.01, 0.23)	0.04
IL-6	3.04	3.00	-0.04 (-0.19, 0.11)	0.58	-0.04 (-0.19, 0.11)	0.57
IL-10	3.52	3.43	-0.09 (-0.19, 0.01)	0.06	-0.09 (-0.19, 0.00)	0.06
IL-12 p70	2.98	2.99	0.01 (-0.05, 0.07)	0.84	0.01 (-0.05, 0.07)	0.82
IL-13	3.15	3.17	0.02 (-0.04, 0.07)	0.51	0.02 (-0.04, 0.07)	0.50
IL-17	2.35	2.35	0.00 (-0.05, 0.04)	0.85	0.00 (-0.05, 0.04)	0.84
TNF-alpha	3.04	2.94	-0.10 (-0.23, 0.03)	0.12	-0.10 (-0.23, 0.03)	0.119
VEGF-C	3.28	3.68	0.40 (0.24, 0.56)	<0.0001	0.40 (0.25, 0.56)	<0.0001
IFN-alpha	20.52	26.80	6.28 (2.20, 10.37)	0.003	6.30 (2.18, 10.41)	0.003
IFN-gamma	2.84	2.82	-0.02 (-0.08, 0.04)	0.43	-0.02 (-0.08, 0.04)	0.43
CD40L	4.51	4.82	0.31 (0.12, 0.50)	0.001	0.31 (0.12, 0.50)	0.001
serpin E1	9.46	9.57	0.11 (0.05, 0.17)	0.0003	0.11 (0.05, 0.17)	<0.0001
EGF	5.07	5.67	0.61 (0.33, 0.89)	<0.0001	0.60 (0.32, 0.89)	<0.0001
Fas	7.52	7.39	-0.13 (-0.31, 0.05)	0.16	-0.13 (-0.31, 0.06)	0.17
GMCSF	3.05	2.94	-0.10 (-0.19, -0.02)	0.016	-0.10 (-0.19, -0.02)	0.02
CCL2	3.10	2.82	-0.28 (-0.46, -0.10)	0.002	-0.45 (-0.70, -0.21)	<0.0001
CCL3	2.72	2.82	0.10 (-0.10, 0.10)	0.32	0.10 (-0.10, 0.30)	0.31
CCL4	2.92	2.91	-0.01 (-0.14, 0.12)	0.88	-0.02 (-0.19, 0.16)	0.85
CCL11	3.16	3.14	-0.02 (-0.07, 0.03)	0.49	-0.02 (-0.07, 0.04)	0.51
CCL22	5.69	5.61	-0.08 (-0.40, 0.25)	0.64	-0.08 (-0.38, 0.22)	0.60
CXCL1	3.15	3.35	0.20 (-0.02, 0.42)	0.07	0.20 (-0.02, 0.42)	0.07
CXCL2	4.87	5.52	0.65 (0.27, 1.04)	0.001	0.56 (0.20, 0.92)	0.003
CXCL10	5.96	5.48	-0.48 (-0.70, -0.26)	<0.0001	-0.49 (-0.71, -0.27)	<0.0001
IL-8/CXCL8	3.44	3.36	-0.08 (-0.31, 0.15)	0.48	-0.08 (-0.31, 0.15)	0.48
Adiponectin	9.30	9.32	0.02 (-0.07, 0.11)	0.69	0.02 (-0.07, 0.11)	0.69
Leptin	5.19	5.53	0.35 (-0.11, 0.80)	0.14	0.31 (-0.02, 0.63)	0.06

CI, Confidence interval; * log transformed because of skewed data

IFN- α , a member of the type 1 interferon family secreted in response to viral infection (26.80 vs 20.52, 6.30 [2.18, 10.41] $p=0.003$). Participants in the urban setting also had higher levels of the growth factors EGF (5.67 vs 5.07, 0.60 [0.32, 0.98] $p<0.00001$) and VEGF (3.68 vs 3.28, 0.40 [0.25, 0.56] $p<0.00001$), and CD40L, a member of the TNF superfamily involved in B-cell activation and class-switching (4.82 vs 4.51, 0.31 [0.12, 0.50] $p=0.001$). Higher levels of serpin E1, an inhibitor of tissue plasminogen activator and urokinase, were also observed in the urban setting (9.57 vs 9.46, 0.11 [0.05, 0.17] $p<0.00001$). Conversely, participants in the urban setting had lower levels of GMCSF (2.94 vs 3.05, -0.10 [-0.19, -0.02] $p=0.02$) and the chemokines CCL2 (2.82 vs 3.10, -0.45 [-0.70, -0.21] $p<0.00001$) and CXCL10 (5.48 vs 5.96, -0.49 [-0.71, -0.27] $p<0.00001$). However, the levels of the chemokine CXCL2 were higher in the urban setting (5.52 vs 4.87, 0.56 [0.20, 0.92] $p=0.003$). No differences were observed for the other analytes.

Associations between the analytes and selected metabolic parameters

The correlations between the analytes and metabolic parameters are shown in [Table 3](#). There was a positive correlation between insulin resistance (HOMA-IR) and the pro-inflammatory analytes CCL22, CXCL1, IL-5, IL-17 and Leptin. Similarly, fasting blood glucose was positively correlated with IL-5, IL-17 and Leptin. Systolic blood pressure and diastolic blood pressure were positively correlated with VEGF-C and Leptin respectively. BMI was positively correlated with IL-5 and Leptin but negatively correlated with CCL22.

Principal component analysis

Correlations were observed among the analytes and the dataset was summarised using principal component analysis. On principal component analysis, six principal components were retained and these accounted for 73% of the variance in the original 30 outcomes. The scree plot constructed to give a visual representation of the PCA eigenvalues is shown in [Figure 1](#).

The resulting principal components were challenging to interpret and did not represent any immediately obvious standard classifications. The first component explained 38% of the total variance and was constituted by CCL3, CCL11, CD40 Ligand, Fas, IFN- α , IFN- γ , IL1- α , IL-2, IL-12 and IL-13. These are a mixed selection of chemoattractants, cytokines and chemokines generally associated with inflammation. All the analytes in this component had positive loadings (coefficients). The second principal component explained a further 13% of the variance in the 30 outcomes and was characterised by chemoattractants and vasoactive mediators Serpin E1, CXCL1, CXCL2, CXCL10, EGF, GMCSF and VEGF-C (all had positive loadings except for GMCSF and CXCL10). The third principal component was characterised by negative loadings (coefficients) of CCL4, IL-8/CXCL8, and the classical markers of inflammation, IL1- β , IL-6, and TNF- α , and explained a further 9% of the variance. The other components and variables contributing to them are shown in [Table 4](#). Associations between the principal components and study setting are shown in [Table 5](#) and [Figure 2](#) and [Figure 3](#). Compared

Table 3. Correlations between inflammatory analytes and metabolic parameters.

Analyte	HOMA-IR	FBG	SBP	DBP	BMI
IL1- α	0.06	0.02	0.10	0.04	-0.03
IL1- β	0.04	0.00	0.10	0.05	0.08
IL-2	0.11*	0.05	0.07	0.03	0.03
IL-4	0.04	0.11	0.10	0.04	0.01
IL-5	0.34***	0.15**	0.02	0.13*	0.58***
IL-6	0.06	0.03	0.02	0.02	0.04
IL-10	0.07	-0.02	-0.08	-0.13*	-0.07
IL-12	0.01	-0.02	0.10	0.08	0.02
IL-13	0.04	0.02	0.09	0.02	-0.04
IL-17	0.12*	0.18**	0.05	0.04	0.00
TNF- α	0.07	0.03	0.02	0.05	0.01
VEGF-C	0.02	-0.12*	0.21**	0.11	0.12
IFN- α	0.10	0.05	0.10	0.06	0.15*
IFN- γ	0.13*	0.09	0.07	0.03	0.00
CD40L	0.06	-0.02	0.12*	0.02	0.07
Serpin E1	0.02	-0.01	0.12*	0.09	0.10
EGF	0.05	-0.03	0.12*	0.02	0.08
Fas	0.04	0.09	0.14*	0.05	0.00
GMCSF	0.08	0.09	-0.03	0.00	-0.02
CCL2	0.00	0.07	0.07	0.13*	0.02
CCL3	0.05	-0.01	0.12*	0.05	-0.01
CCL4	0.07	0.00	0.10	0.07	0.05
CCL11	0.07	0.14*	0.08	0.07	0.04
CCL22	0.17**	0.08	-0.12*	-0.13*	-0.17**
CXCL1	0.17**	0.02	0.15*	0.06	0.10
CXCL2	0.10	-0.02	0.10	0.02	0.10
CXCL10	0.08	0.09	-0.07	-0.03	-0.06
IL-8/CXCL8	0.04	-0.05	0.07	0.03	0.04
Adiponectin	-0.03	0.01	0.03	0.01	-0.01
Leptin	0.37***	0.18**	-0.03	0.15**	0.58***

*Correlation coefficient with $0.01 \leq p < 0.05$; **Correlation coefficient with $0.001 \leq p < 0.01$; ***Correlation coefficient with $p < 0.001$; HOMAIR, Homeostatic Model Assessment of Insulin Resistance; FBG, Fasting blood glucose; SBP, Systolic blood pressure; DBP, Diastolic blood pressure; BMI, Body mass index

with the rural setting, the urban setting had higher scores for principal component 2, implying higher representation of chemoattractants and vasoactive mediators among individuals in the

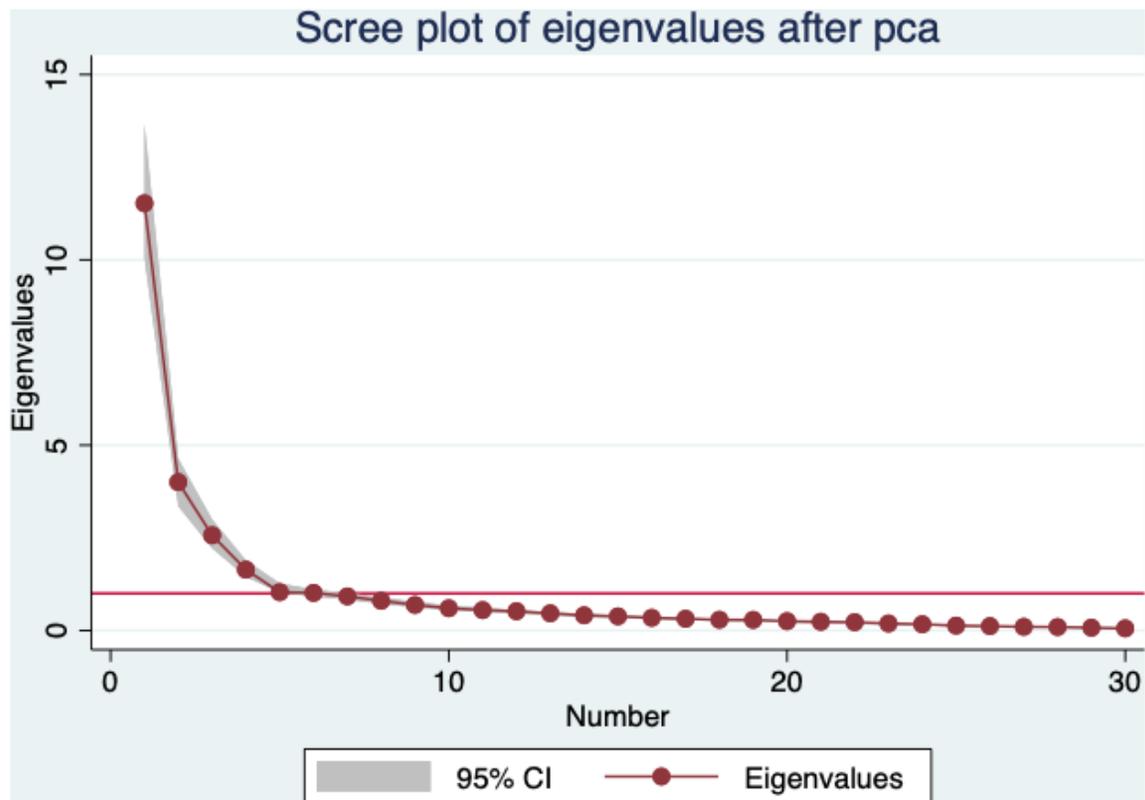


Figure 1. Scree plot of eigenvalues after principal component analysis. CI, Confidence interval.

Table 4. Principal components and variables contributing to them.

Analyte	PC1	PC2	PC3	PC4	PC5	PC6
IL1-alpha	0.22	-0.03	-0.12	-0.15	-0.04	0.08
IL1-beta	0.21	0.11	-0.31	0.09	-0.03	-0.01
IL-2	0.24	-0.07	-0.11	0.02	-0.01	0.01
IL-4	0.20	-0.11	0.08	-0.11	-0.28	0.19
IL-5	0.13	-0.07	0.22	0.60	0.02	0.07
IL-6	0.19	0.05	-0.40	0.16	0.03	-0.06
IL-10	0.19	-0.21	0.10	-0.05	0.23	-0.17
IL-12	0.20	-0.16	0.08	-0.06	-0.20	0.14
IL-13	0.22	-0.15	0.11	-0.09	-0.21	0.08
IL-17	0.18	-0.11	0.07	-0.03	-0.21	0.29
TNF-alpha	0.21	0.06	-0.33	0.10	0.02	-0.04
VEGF-C	0.12	0.34	0.21	-0.11	0.13	-0.08
IFN-alpha	0.17	0.13	0.10	0.04	-0.07	-0.10
IFN-gamma	0.21	-0.10	-0.02	-0.05	-0.09	0.14
CD40 Ligand	0.23	0.22	0.20	-0.07	0.03	-0.03

Analyte	PC1	PC2	PC3	PC4	PC5	PC6
serpin E1	0.11	0.29	0.15	-0.01	0.13	0.08
EGF	0.20	0.25	0.23	-0.04	0.03	-0.04
Fas	0.22	-0.20	0.10	-0.08	-0.10	0.04
GMCSF	0.17	-0.29	0.08	-0.05	0.25	-0.22
CCL2	0.13	-0.17	-0.05	-0.05	0.36	-0.17
CCL3	0.17	0.15	-0.16	0.00	-0.09	0.02
CCL4	0.23	0.10	-0.27	0.06	0.01	-0.02
CCL11	0.22	-0.17	0.14	-0.09	-0.12	0.04
CCL22	0.12	-0.04	0.14	-0.20	0.24	-0.10
CXCL1	0.15	0.28	0.11	-0.04	0.04	-0.03
CXCL2	0.18	0.29	0.19	-0.01	0.05	-0.11
CXCL10	0.14	-0.35	0.01	0.00	0.28	-0.20
IL-8/CXCL8	0.20	0.15	-0.30	0.07	0.05	-0.07
Adiponectin	0.01	0.02	-0.05	-0.05	0.56	0.78
Leptin	0.07	-0.06	0.21	0.67	0.04	0.04

PC, Principal component; The numbers in the table show, for each PC, the coefficients corresponding to each analyte. For each analyte, the PC with the highest loading (in bold) was interpreted as representative of that analyte.

Table 5. Associations between the principal components and study setting.

	Setting		Unadjusted		Adjusted for age and sex	
	Rural	Urban	Mean difference (95% CI)	P value	Mean difference 95% CI	P value
PC1	-0.07	0.15	0.23 (-0.58, 1.04)	0.58	0.23 (-0.59, 1.04)	0.59
PC2	-0.37	0.77	1.14 (0.68, 1.60)	<0.00001	1.14 (0.68, 1.60)	<0.00001
PC3	-0.21	0.43	0.64 (0.26, 1.02)	0.001	0.64 (0.26, 1.01)	0.001
PC4	-0.07	0.14	0.21 (-0.10, 0.51)	0.18	0.18 (-0.04, 0.40)	0.10
PC5	0.07	-0.15	-0.22 (-0.47, 0.02)	0.07	-0.23 (-0.47, 0.01)	0.06
PC6	-0.06	0.13	0.19 (-0.05, 0.43)	0.13	0.19 (-0.05, 0.43)	0.12

CI, Confidence interval; PC, Principal component

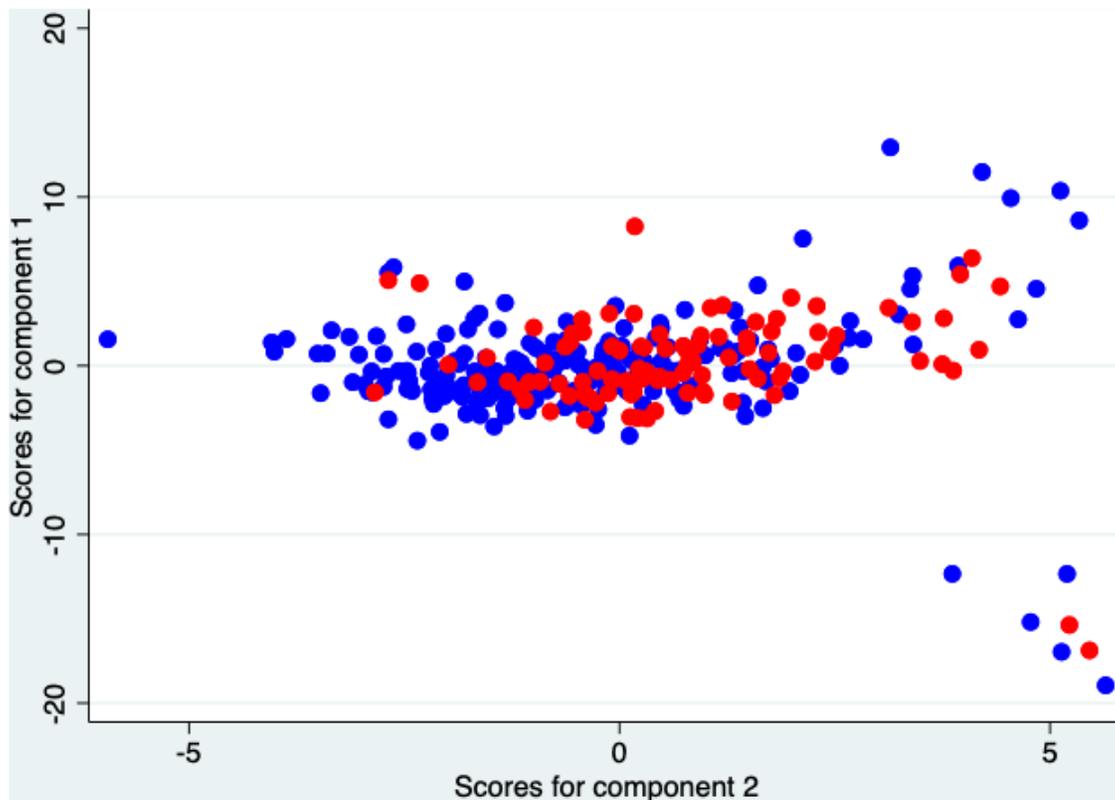


Figure 2. Principal component analysis biplot of components 1 and 2. Blue, Rural residents; Red, Urban residents.

urban setting. The urban setting also had higher scores for principal component 3. This implies that there is lower representation of the classical inflammatory mediators IL1-beta, IL-6 and TNF-alpha in the urban setting compared to the rural setting (considering the negative coefficients shown in Table 4).

Associations between the principal components and metabolic parameters are shown in Table 6. Principal component 4 (characterised by IL-5 and Leptin) was positively correlated with HOMA-IR, fasting blood glucose, diastolic blood pressure and BMI. The strongest correlations were observed with BMI (correlation coefficient 0.61, $p < 0.0001$) and HOMA-IR (0.31,

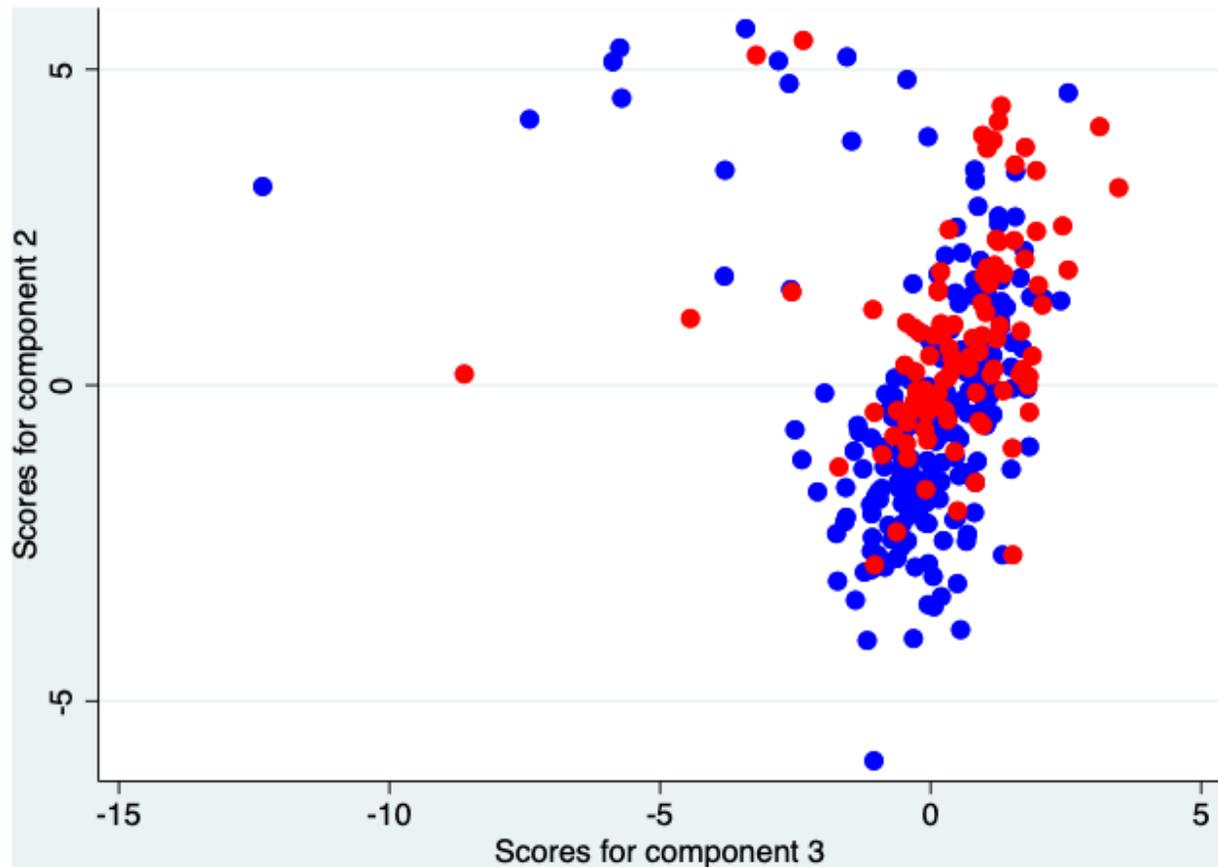


Figure 3. Principal component analysis biplot of components 2 and 3. Blue, Rural residents; Red, Urban residents.

Table 6. Associations between the principal components and metabolic parameters.

Component	HOMA-IR		FBG		SBP		DBP		BMI	
	coeff.	P-value	coeff.	P-value	coeff.	P-value	coeff.	P-value	coeff.	P-value
PC1	0.13	0.02	0.05	0.34	0.11	0.05	0.06	0.33	0.07	0.20
PC2	-0.02	0.67	-0.11	0.05	0.12	0.04	0.06	0.33	0.08	0.18
PC3	0.11	0.06	0.06	0.33	0.03	0.57	0.03	0.63	0.17	0.003
PC4	0.31	<0.0001	0.13	0.02	-0.05	0.40	0.14	0.01	0.61	<0.0001
PC5	0.05	0.41	-0.03	0.63	-0.09	0.11	-0.04	0.44	-0.01	0.89
PC6	-0.02	0.74	0.07	0.21	0.10	0.09	0.07	0.21	0.06	0.31

PC, Principal component; HOMA-IR, Homeostatic Model Assessment of Insulin Resistance; FBG, Fasting blood glucose; SBP, Systolic blood pressure; DBP, Diastolic blood pressure; BMI, Body mass index

$p < 0.0001$). Other significant correlations were found between principal component 1 and HOMA-IR (0.13, $p = 0.02$), principal component 2 and systolic blood pressure (0.12, $p = 0.04$), and principal component 3 and BMI (0.17, $p = 0.003$).

Discussion

In this paper, we have shown that, compared with the rural environment, the urban environment was associated with higher levels of pro-inflammatory chemokines: EGF, VEGF-C, CD40L,

serpin E1 but lower levels of GMCSF, CCL2 and CXCL10. Furthermore, the rural environment was associated with lower levels of the Th2 cytokine IL-5 but there was no difference in the other Th2, Th1, Th17 and regulatory cytokines between the settings. On principal component analysis, the urban environment had higher representation of chemoattractants and vasoactive mediators (PC2) but lower representation of the classical inflammatory mediators (PC3). The metabolic parameters HOMA-IR, fasting blood glucose, blood pressure and BMI correlated positively with a largely distinct set of principal components characterised by pro-inflammatory analytes.

It was interesting to find that the levels of the growth factors EGF and VEGF-C were higher in the urban setting. EGF is a pro-inflammatory peptide that stimulates cell growth and proliferation. VEGF-C promotes lymphangiogenesis by directing the proliferation and migration of lymphatic epithelial cells and is also involved in the stimulation of growth of vascular tissue²⁶. We also found lower levels of soluble CD40L in the rural compared with the urban population. CD40L is a pro-inflammatory molecule found both in a soluble form and expressed on activated T lymphocytes. It is pro-inflammatory to endothelial cells, has been suggested to be a marker of atherogenesis²⁷ and is elevated in individuals with diabetes, metabolic syndrome and insulin resistance²⁷. The higher levels of these cytokines in the urban setting is consistent with the higher burden of these conditions in urban settings.

We found higher levels of serpin E1 in the urban setting compared to the rural setting and this is consistent with findings elsewhere²⁸. Serpins have a role in inflammation and immune function. Serpin E1, also known as plasminogen inhibitor 1, is a serine protease inhibitor that inhibits tissue plasminogen inhibitor and urokinase, which are activators of plasminogen and fibrinolysis. Increased levels of serpin E1 are a risk factor for atherosclerosis and thrombosis³⁰ and evidence suggests that increased levels accelerate vascular disease in patients with diabetes^{31,32}. The lower levels of serpin E1, therefore, accord with evidence that the rural environment has lower cardiovascular risk than the urban environment. Serpin E1 levels have been found to positively correlate with insulin resistance³³ but we did not find that in our study. On data reduction by principal component analysis, the individual findings were supported by higher levels of PC2 (characterised by various chemoattractants and vasoactive peptides) in the urban, compared to the rural environment.

The rural environment is associated with a higher microbial and parasite load and burden of chronic and recurrent infections³⁴. It was therefore surprising to find no differences in the Th1, Th2, Th17 cytokines except for lower levels of IL-5 in the rural population compared to the urban population. Despite the higher burden of helminth infections in this rural setting¹⁵, there was no difference in the regulatory cytokines IL-10 and TGF-beta between the settings. It is possible that there is substantial exposure of individuals in this particular urban

setting to infectious agents (both current and previous) sufficient to achieve a similar regulatory profile to individuals in the rural setting. Owing to the complexity of the distribution of antigens which trigger the immune response, investigating cellular profiles and responses to stimulation, to complement studies of plasma levels of cytokines, would be necessary to obtain a fuller understanding of immunological profiles in different settings. For example, contrary to our findings, a study in Senegal found that rural individuals had significantly higher expression of Th1, Th2, and Th22 cells compared to the urban population in response to stimulation⁶.

In contrast, we found lower levels of the pro-inflammatory analytes GMCSF, CCL2 and CXCL10 in the urban population compared to the rural population, underscoring the complexity of immune interactions. After data reduction with principal component analysis, the urban environment had higher values for PC3, implying lower representation of the classical inflammatory mediators IL1-beta, IL-6 and TNF-alpha. This may suggest that, in our study, the urban setting is associated with less classical inflammation than the rural environment. Despite the difficulty in interpreting the principal components, it is clear that the inflammatory profiles of individuals in the rural and urban settings are different.

Our study had a number of limitations, which we have considered. The analytes were measured in plasma and generally, levels of most cytokines are low in plasma. This could have contributed to the inability to find any differences in the Th1, Th2 and Th17 cytokines. Stimulation of peripheral blood mononuclear cells may have resulted in higher measurable levels of the analytes. Genetic factors influencing the immune system were not studied, however, the ethnic groups in the two settings are similar¹⁵. Also, many tests were performed with no formal adjustment for multiple testing, therefore, chance findings cannot be ruled out. However, of the 30 analytes tested, 10 showed evidence of differing between the rural and urban settings at $p < 0.05$ (more than the $30 \times 0.05 = 1.5$ that we would expect to see by chance alone).

Taken together, our exploratory work on inflammatory analytes suggests that investigating differences in inflammatory profiles between rural and urban populations is complex but differences in these two populations exist. This could have broad implications for health such as susceptibility to infections and response to vaccines. However, the focus for this study was on the implications for cardiometabolic risk. Considering that inflammation correlates with some metabolic diseases, inflammation may be one of the pathways to explain the difference in cardiometabolic diseases between the urban and rural environments.

Data availability

Underlying data

LSHTM Data Compass: Data supporting "Profiles of inflammation in rural and urban Uganda". <https://doi.org/10.17037/DATA.0000245925>.

This project contains the following underlying data:

- Inflammation_data.csv
- Inflammation_data_codebook.html

Extended data

LSHTM Data Compass: Data supporting “Profiles of inflammation in rural and urban Uganda”. <https://doi.org/10.17037/DATA.00002459>²⁵.

This project contains the following extended data:

- Consent_forms.zip (consent forms and information sheets)

Data are available under the terms of the [Creative Commons Attribution 4.0 International license \(CC-BY 4.0\)](https://creativecommons.org/licenses/by/4.0/).

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References

1. Donath MY, Shoelson SE: **Type 2 diabetes as an inflammatory disease.** *Nat Rev Immunol.* 2011; **11**(2): 98–107. [PubMed Abstract](#) | [Publisher Full Text](#)
2. De Paoli F, Staels B, Chinetti-Gbaguidi G: **Macrophage phenotypes and their modulation in atherosclerosis.** *Circ J.* 2014; **78**(8): 1775–81. [PubMed Abstract](#) | [Publisher Full Text](#)
3. GBD 2016 Causes of Death Collaborators: **Global, regional, and national age-sex specific mortality for 264 causes of death, 1980–2016: a systematic analysis for the Global Burden of Disease Study 2016.** *Lancet.* 2017; **390**(10100): 1151–210. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
4. GBD 2016 DALYs and HALE Collaborators: **Global, regional, and national disability-adjusted life-years (DALYs) for 333 diseases and injuries and healthy life expectancy (HALE) for 195 countries and territories, 1990–2016: a systematic analysis for the Global Burden of Disease Study 2016.** *Lancet.* 2017; **390**(10100): 1260–344. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
5. United Nations Department of Economic and Social Affairs Population Division: **World Urbanization Prospects: The 2018 revision.** (ST/ESA/SER/A/420). New York: United Nations; 2019. [Reference Source](#)
6. Mbow M, de Jong SE, Meurs L, et al.: **Changes in immunological profile as a function of urbanization and lifestyle.** *Immunology.* 2014; **143**(4): 569–77. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
7. Idaghdour Y, Storey JD, Jadallah SJ, et al.: **A genome-wide gene expression signature of environmental geography in leukocytes of Moroccan Amazighs.** *PLoS Genet.* 2008; **4**(4): e1000052. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
8. Böbel TS, Hackl SB, Langgartner D, et al.: **Less immune activation following social stress in rural vs. urban participants raised with regular or no animal contact, respectively.** *Proc Natl Acad Sci U S A.* 2018; **115**(20): 5259–64. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
9. Im SG, Choi H, Jeon YH, et al.: **Comparison of Effect of Two-Hour Exposure to Forest and Urban Environments on Cytokine, Anti-Oxidant, and Stress Levels in Young Adults.** *Int J Environ Res Public Health.* 2016; **13**(7): 625. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
10. Yudkin JS, Yajnik CS, Mohamed-Ali V, et al.: **High levels of circulating proinflammatory cytokines and leptin in urban, but not rural, Indians. A potential explanation for increased risk of diabetes and coronary heart disease.** *Diabetes Care.* 1999; **22**(2): 363–4. [PubMed Abstract](#) | [Publisher Full Text](#)
11. Darko SN, Yar DD, Owusu-Dabo E, et al.: **Variations in levels of IL-6 and TNF- α in type 2 diabetes mellitus between rural and urban Ashanti Region of Ghana.** *BMC Endocr Disord.* 2015; **15**: 50. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
12. Butcher L, Pérès K, André P, et al.: **Association between plasma CCL11 (eotaxin-1) and cognitive status in older adults: Differences between rural and urban dwellers.** *Exp Gerontol.* 2018; **113**: 173–9. [PubMed Abstract](#) | [Publisher Full Text](#)
13. Danielewicz H, Dębińska A, Drabik-Chamerska A, et al.: **IL-4RA gene expression in PBMC with regard to place of living and atopy status.** *Adv Clin Exp Med.* 2018; **27**(2): 173–7. [PubMed Abstract](#) | [Publisher Full Text](#)
14. Amoah AS, Obeng BB, May L, et al.: **Urban-rural differences in the gene expression profiles of Ghanaian children.** *Genes Immun.* 2014; **15**(5): 313–9. [PubMed Abstract](#) | [Publisher Full Text](#)
15. Sanya RE, Biraro IA, Nampijja M, et al.: **Contrasting impact of rural, versus urban, living on glucose metabolism and blood pressure in Uganda [version 1; peer review: awaiting peer review].** *Wellcome Open Res.* 2020; **5**: 39. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
16. Sanya RE, Webb EL, Zziwa C, et al.: **The effect of helminth infections and their treatment on metabolic outcomes: results of a cluster-randomized trial.** *Clin Infect Dis.* 2020; **71**(3): 601–613. [PubMed Abstract](#) | [Publisher Full Text](#)
17. Sanya RE, Nkurunungi G, Hoek Spaans R, et al.: **The Impact of Intensive Versus Standard Anthelmintic Treatment on Allergy-related Outcomes, Helminth Infection Intensity, and Helminth-related Morbidity in Lake Victoria Fishing Communities, Uganda: Results From the LaVIISWA Cluster-randomized Trial.** *Clin Infect Dis.* 2019; **68**(10): 1665–74. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
18. Nampijja M, Webb EL, Kaweesa J, et al.: **The Lake Victoria Island Intervention Study on Worms and Allergy-related diseases (LaVIISWA): study protocol for a randomised controlled trial.** *Trials.* 2015; **16**: 187. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
19. Nkurunungi G, Lubyayi L, Versteeg SA, et al.: **Do helminth infections underpin urban-rural differences in risk factors for allergy-related outcomes? Clin Exp Allergy. 2019; **49**(5): 663–76. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)**
20. Breen EJ, Polaskova V, Khan A: **Bead-based multiplex immuno-assays for cytokines, chemokines, growth factors and other analytes: median fluorescence intensities versus their derived absolute concentration values for statistical analysis.** *Cytokine.* 2015; **71**(2): 188–98. [PubMed Abstract](#) | [Publisher Full Text](#)
21. Breen EJ, Tan W, Khan A: **The Statistical Value of Raw Fluorescence Signal in Luminex xMAP Based Multiplex Immunoassays.** *Sci Rep.* 2016; **6**: 26996. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
22. Schutz C, Barr D, Andrade BB, et al.: **Clinical, microbiologic, and immunologic determinants of mortality in hospitalized patients with HIV-associated tuberculosis: A prospective cohort study.** *PLoS Med.* 2019; **16**(7): e1002840. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
23. Turi KN, Shankar J, Anderson LJ, et al.: **Infant Viral Respiratory Infection Nasal Immune-Response Patterns and Their Association with Subsequent Childhood Recurrent Wheeze.** *Am J Respir Crit Care Med.* 2018; **198**(8): 1064–73. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
24. Hair JF, Black WC, Babin BJ, et al.: **Multivariate data analysis: Pearson new international edition.** Pearson Higher Ed; 2013. [Reference Source](#)
25. Sanya R, Webb E: **Data supporting “Profiles of inflammation in rural and urban Uganda”.** [Data Collection]. London School of Hygiene & Tropical Medicine, London, United Kingdom, 2021. <https://www.doi.org/10.17037/DATA.00002459>
26. Rauniyar K, Jha SK, Jeltsch M: **Biology of Vascular Endothelial Growth Factor C in the Morphogenesis of Lymphatic Vessels.** *Front Bioeng Biotechnol.* 2018; **6**: 7. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)

27. Szmítko PE, Wang CH, Weisel RD, *et al.*: **New markers of inflammation and endothelial cell activation: Part I.** *Circulation*. 2003; **108**(16): 1917–23.
[PubMed Abstract](#) | [Publisher Full Text](#)
28. Gokulakrishnan K, Deepa R, Mohan V, *et al.*: **Soluble P-selectin and CD40L levels in subjects with prediabetes, diabetes mellitus, and metabolic syndrome--the Chennai Urban Rural Epidemiology Study.** *Metabolism*. 2006; **55**(2): 237–42.
[PubMed Abstract](#) | [Publisher Full Text](#)
29. Kodaman N, Aldrich MC, Sobota R, *et al.*: **Cardiovascular Disease Risk Factors in Ghana during the Rural-to-Urban Transition: A Cross-Sectional Study.** *PLoS One*. 2016; **11**(10): e0162753.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
30. Vaughan DE: **PAI-1 and atherothrombosis.** *J Thromb Haemost*. 2005; **3**(8): 1879–83.
[PubMed Abstract](#) | [Publisher Full Text](#)
31. Hoekstra T, Geleijnse JM, Schouten EG, *et al.*: **Plasminogen activator inhibitor-type 1: its plasma determinants and relation with cardiovascular risk.** *Thromb Haemost*. 2004; **91**(5): 861–72.
[PubMed Abstract](#) | [Publisher Full Text](#)
32. Sobel BE, Woodcock-Mitchell J, Schneider DJ, *et al.*: **Increased plasminogen activator inhibitor type 1 in coronary artery atherectomy specimens from type 2 diabetic compared with nondiabetic patients: a potential factor predisposing to thrombosis and its persistence.** *Circulation*. 1998; **97**(22): 2213–21.
[PubMed Abstract](#) | [Publisher Full Text](#)
33. Juhan-Vague I, Alessi MC, Vague P: **Increased plasma plasminogen activator inhibitor 1 levels. A possible link between insulin resistance and atherothrombosis.** *Diabetologia*. 1991; **34**(7): 457–62.
[PubMed Abstract](#) | [Publisher Full Text](#)
34. Bickler SW, Wang A, Amin S, *et al.*: **Urbanization in Sub-Saharan Africa: Declining Rates of Chronic and Recurrent Infection and Their Possible Role in the Origins of Non-communicable Diseases.** *World J Surg*. 2018; **42**(6): 1617–28.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)

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The authors investigated the association of several inflammatory markers with selected cardiovascular risk markers in apparently healthy subjects aged ≥ 10 years. Data obtained from rural and urban residents were evaluated employing different statistical approaches. Differences in presentation of some classical inflammatory mediators, chemoattractant, and vasoactive peptides have been observed; and correlations between pro-inflammatory analytes and cardiometabolic risk markers were reported. It has been concluded that different expressions of certain pro-inflammatory mediators may have important health consequences and may contribute to an increased cardiometabolic risk in the urban environment.

This is a potentially interesting study with some novelty. However, in my opinion, several weaknesses should be addressed before the study can proceed to indexing.

Major comments:

Introduction -

Last paragraph: Motivation to analyze 30 different inflammatory markers in apparently healthy subjects should be explained. A hypothesis is missing – it should be clearly formulated.

Methods, Results -

I have concerns about the clinical interpretation of data obtained using the raw fluorescence intensity values of the analytes instead of using the returned concentrations. As follows from the advocacy of this attitude, several measured markers were out of the standard range, e.g. below the sensitivity of the assay or even below the blank. The authors state that analyses using fluorescence intensities do not require censoring and provide more statistical power than analysis based on concentrations. I suppose that this attitude is feasible from the mathematical (statistical) point of view but irrelevant clinically. The physician evaluates concentrations or activities of analytes with the knowledge of their reference ranges - not the physical units used for

quantification as these are clinically uninterpretable. Even if the calibration curves would be linear. As the calibration curves for 30 different analytes are not identical, the same fluorescence returns different concentrations, even by order(s) of magnitude. In biomedical research, statistics is an auxiliary method, helping in the interpretation of the results. An interpretation should be strictly done in a clinical context on clinically relevant data, and only these clinically relevant data (not physical units) should be statistically evaluated.

I suppose that the problem lies in the fact that several markers were under the detection limit of the assay (what could be anticipated as healthy subjects were studied). The statistician did not perceive the fact as a problem since the fluorescent units were available. The physicians did not comment on the fact that they evaluate concentrations and not fluorescent units. A physicist or biochemist who would get alert learning that fluorescence is to be evaluated and serve as a basis for clinical interpretation was probably not in the team. This, unfortunately, resulted in an extended statistical exercise that has no clinical relevance. If the analyte is under the detection limit of the assay it should be acknowledged. These data should be handled as unavailable instead of looking for a way how to overcome this reality eventually leading to misinterpretation.

I suppose that the authors should give the data as concentrations (not fluorescence units); for each analyte, the number of subjects in whom valid concentrations were obtained should be indicated; numbers of should be given also in the case of simple correlations performance; and multivariate analyses should be conducted using selected data sets – those which yielded reliable concentrations in all, or a vast majority of the probands. (the PCA is robust, among others, in the handling of missing data, allowing for a reasonable arbitrary selection of what percentage of data in the set must be available to include the variable into the analysis. And probably the missing data should be random, not always missing in the same subjects).

Even though in the dot plots the 95% Hotelling's ellipse is not depicted, it is obvious that subjects (e.g., in the right lower corner of Figure 1) are major outliers. In the case that such outliers are retained in the analysis, their characteristics should be given and it should be explained why they were retained in the analysis, and what changed after their exclusion.

It is not stated whether the median or the mean of the sum of fluorescence signal intensities has been used, but it is rather marginal in the context.

It is not indicated how the sample size was calculated thus, what was the adequate expected power.

Minor comments:

Methods -

The authors should indicate whether standard laboratory methods were used to quantify blood chemistry variables, and the brand and the manufacturer of the employed analyzer(s).

Results -

Table 1: Regarding decimals, for clearness, FPG, BP, and BMI data should be reported in a manner generally used in clinics. Indication of decimals or more decimals than necessary does not add to

the scientific value of the paper and is rather confusing for the reader.

Is the work clearly and accurately presented and does it cite the current literature?

No

Is the study design appropriate and is the work technically sound?

No

Are sufficient details of methods and analysis provided to allow replication by others?

Yes

If applicable, is the statistical analysis and its interpretation appropriate?

No

Are all the source data underlying the results available to ensure full reproducibility?

Yes

Are the conclusions drawn adequately supported by the results?

No

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Physiology, metabolic syndrome

I confirm that I have read this submission and believe that I have an appropriate level of expertise to state that I do not consider it to be of an acceptable scientific standard, for reasons outlined above.
