

9. Dellacà RL, Gobbi A, Pastena M, Pedotti A, Celli B. Home monitoring of within-breath respiratory mechanics by a simple and automatic forced oscillation technique device. *Physiol Meas* 2010;31:N11–N24.
10. Gobbi A, Dellacà RL, King G, Thamrin C. Toward predicting individual risk in asthma using daily home monitoring of resistance. *Am J Respir Crit Care Med* 2017;195:265–267.

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Two Randomized Trials of the Effect of Live Attenuated Influenza Vaccine on Pneumococcal Colonization

To the Editor:

The human nasopharynx is frequently colonized by *Streptococcus pneumoniae* (the pneumococcus), serving as the reservoir for transmission, a state that necessarily precedes invasive pneumococcal infection. Influenza infection increases pneumococcal colonization density and dysregulates host immune responses, increasing the risk of secondary bacterial pneumonia and death (1–3).

Live attenuated influenza vaccine (LAIV) nasal spray has been used in the United States since 2003, and it has reduced severe influenza disease in the United Kingdom since its introduction in 2013 into the national pediatric immunization program. In mice, LAIV vaccination increases the density and duration of pneumococcal colonization (2) and rates of otitis media. In children, LAIV is associated with increased rates and density of bacterial colonization (4). Although LAIV is safe and not associated with increases in pneumococcal disease, these data suggest that it could increase pneumococcal transmission to susceptible individuals (5).

We therefore undertook two trials (EudraCT 2014-004634-26) using an established human challenge model to evaluate the effects of LAIV on the dynamics of pneumococcal colonization. Some of the results of these studies have been previously reported in the form of a preprint (<https://doi.org/10.1101/343319>). An extensive immunological investigation to accompany these clinical data has been published (6). Healthy nonsmoking volunteers, 18–50 years old, consented to participate in double-blinded, randomized, placebo-controlled trials reflecting alternative scenarios: 1) immunization first (LAIV precedes nasopharyngeal inoculation with pneumococcus by 3 days) and 2) colonization first (LAIV is administered 3 days after colonization with pneumococcus). The participants, who were uncolonized at baseline, randomly received

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either intervention (nasal LAIV paired with intramuscular placebo of normal saline; AstraZeneca) or control (nasal placebo of normal saline paired with intramuscular influenza vaccination [Fluarix Tetra; GlaxoSmithKline]) with concealment by blindfolding. All of the participants gave written informed consent, with approval from the North West NHS Research Ethics Committee (14/NW/1460).

All of the participants were inoculated with *S. pneumoniae* serotype 6B strain BHN418 (80,000 cfu per nostril) in 0.1 ml solution (7). (The *S. pneumoniae* BHN418 sequence [GI:557376079] is available from <https://www.ncbi.nlm.nih.gov/nucleotide/557376079>). “Colonization positivity” was determined by serial nasal washes and defined by detection of serotype 6B by culture at a programmed time point from 2 to 29 days (7, 8). In parallel, PCR detection of pneumococcal *lytA* was performed. In the “immunization first” study, LAIV vaccination preceded pneumococcal inoculation by 3 days (primary endpoint: colonization rate). This order was reversed for the “colonization first” study (primary endpoint: area under the curve [AUC] of bacterial density between Days 2 and 14). Results are presented as modified intention to treat, excluding those who did not receive immunization or inoculation per protocol, or did not complete follow-up. Generalized linear models were used to compare colonization positivity, duration of colonization, and AUC bacterial density, with generalized estimating equations used for comparison at multiple time points. Full methodological and other details are available online in the form of a preprint (<https://doi.org/10.1101/343319>).

In the “immunization first” study (Figure 1), we enrolled 202 participants; 130 of these subjects were inoculated and 117 were analyzed ($n = 55$ LAIV, $n = 62$ control; overall mean age, 20 yr [range, 18–48 yr]; 58% female). Pneumococcal colonization rates were similar in LAIV participants and control subjects (25/55 [45.5%] vs. 24/62 [38.7%]; odds ratio [OR], 1.32; $P = 0.46$), although the LAIV-treated group had consistently yet nonsignificantly higher rates at each time point. PCR detection rates were significantly higher in the LAIV group than in the control group at Day 2 (33/55 [60.0%] vs. 25/62 [40.3%]; OR, 2.22; $P = 0.03$). The median duration of colonization was not different between the groups by conventional microbiology (22 d [interquartile range (IQR), 22–29] and 22 d [IQR, 14–29] in the LAIV and control groups, respectively; $P = 0.09$) or PCR (median, 22 d [IQR, 7–29] LAIV vs. 14 d [IQR, 7–22] control; $P = 0.45$). Mean colonization densities were consistently increased in the LAIV group, with statistical significance at Day 9 representing a 10-fold (1 \log_{10}) increase in colonization density in the LAIV group (2.82 ± 1.78 vs. 1.81 ± 1.39 \log_{10} titers, $P = 0.03$; Figure 1). PCR results showed the same pattern, with significantly higher densities in the LAIV group at Day 2 ($P = 0.03$).

Four participants with laboratory-confirmed other viral infections (three influenza B in the control arm, one rhinovirus in the LAIV arm) had among the highest bacterial densities of their cohorts. Among pneumococcal-colonized individuals, the AUC of colonization density was higher in the LAIV group than in the control group, with borderline statistical significance at Days 2–14 ($P = 0.05$), and reached statistical significance after exclusion of participants who had nasal-swab PCR evidence of concurrent wild-type viral illness (three influenza B in the control arm, one rhinovirus in the LAIV arm; data not shown; $P = 0.03$) after presenting with symptoms of illness.

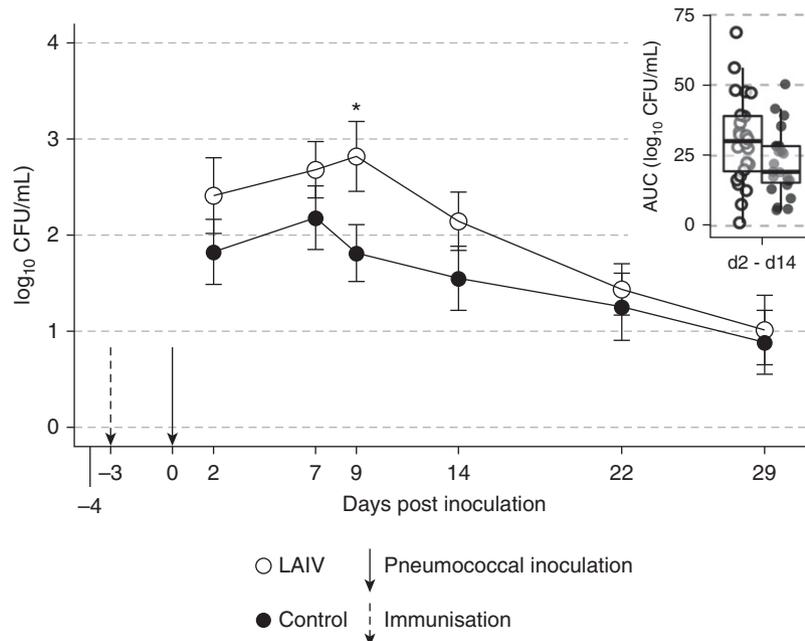


Figure 1. Immunization first: live attenuated influenza vaccine (LAIV) precedes nasopharyngeal inoculation with pneumococcus—effect on colonization dynamics of LAIV vaccination given at Day -3 . Density dynamics after pneumococcal inoculation (on Day 0) are calculated from classical microbiology [$\log_{10}(\text{cfu/ml} + 1)$]. Mean density of *Streptococcus pneumoniae* for each nasal wash time point among participants in whom serotype 6B was detectable at any point. Bars represent SE. Inset: area under the curve (AUC) of density–time from Day 2 to Day 14 (box plot of median with interquartile range, with whiskers at $1.5\times$ the interquartile range). *Statistically significant difference ($P < 0.05$).

In the “colonization first” study (Figure 2), 316 participants consented, 206 were screened, and 163 participants were included in the modified intention-to-treat analysis ($n = 73$ LAIV, $n = 90$ control; overall mean age, 20 yr [range, 18–46 yr]; 55% female).

Data from 17 participants (10%) were excluded owing to non-study-serotype *S. pneumoniae* colonization. AUC colonization densities for each time period were consistently lower in the LAIV group, although the difference was not statistically significant

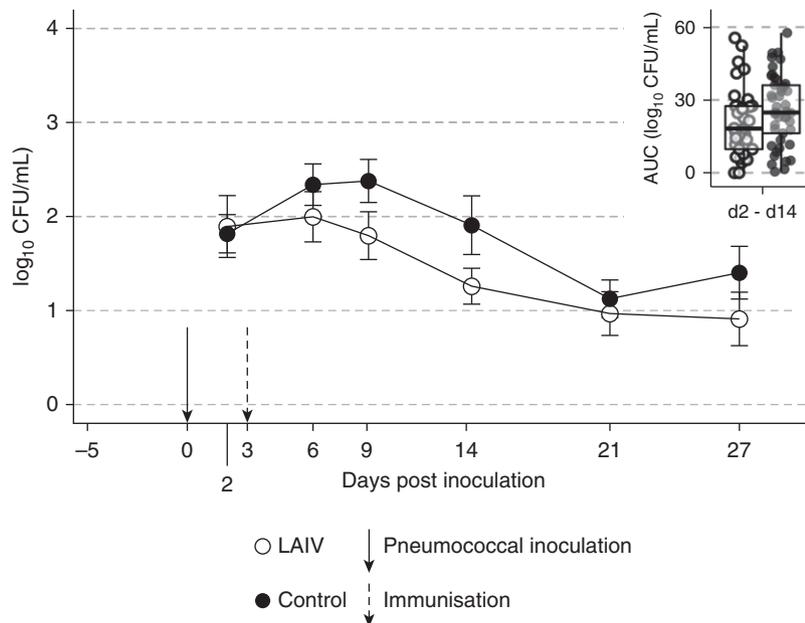


Figure 2. Colonization first: live attenuated influenza vaccine (LAIV) is administered to subjects already inoculated with pneumococcus—effect on colonization dynamics. Experimental inoculation to pneumococcus is performed on Day -3 . Density dynamics after LAIV vaccination or control (on Day 0) are calculated from classical microbiology [$\log_{10}(\text{cfu/ml} + 1)$]. Mean density of *Streptococcus pneumoniae* for each nasal wash time point among participants in whom serotype 6B was detectable at any point. Bars represent SE. Inset: area under the curve (AUC) of density–time from Day 2 to Day 14 (the primary endpoint, box plot of median with interquartile range, with whiskers at $1.5\times$ the interquartile range).

($P = 0.11$ for Days 2–14 primary endpoint; Figure 2). By PCR, a significantly lower AUC was evident in the LAIV group compared with the control group on Days 2–27 ($P = 0.03$).

Rates of colonization did not differ between the LAIV and control groups by conventional microbiology (36/73 [49.3%] vs. 45/90 [50.0%] respectively; OR, 0.97; $P = 0.93$). The median colonization duration did not differ between the two groups (21 vs. 27 d, $P = 0.17$) by conventional microbiology, although it was lower in the LAIV group by PCR (14 vs. 27 d, $P = 0.001$).

There were no serious adverse events related to the intervention in either study.

In the largest trial to date involving a controlled human coinfection model, we have studied for the first time the impact of coinfection of a live viral vaccine and a bacterial pathogen. Immunological parameters have been reported separately (6).

Antecedent LAIV administration caused modest but significant transient effects on pneumococcal colonization, in keeping with a pediatric randomized controlled trial that showed an increased pneumococcal density after LAIV (2). In our study, the inverse scenario (LAIV after pneumococcal colonization) was associated with reduced colonization density and colonization rates at Day 27, decreased AUC, and earlier bacterial clearance.

Our model, consistent with murine coinfection disease models, reinforces the notion that the precedence of pathogen exposure might determine disease outcome: pneumococcal infection after influenza might exacerbate disease, whereas pneumococcus infection preceding influenza might reduce mortality (9). We used complementary methods for bacterial detection: although PCR is more sensitive and could detect DNA in the absence of viable pathogen, the persistence beyond 2 days suggests lower-density colonization, which is unmeasurable by culture.

These studies were limited by size and the evaluation of a single pneumococcal serotype in healthy adults likely to have neutralizing influenza antibodies. Any effect of LAIV in children may therefore be more pronounced owing to lower antibody titers, increased viral shedding, and higher natural rates of pneumococcal colonization acquisition. Future vaccine studies should evaluate the effect on pathogens not directly targeted by the vaccine, including their onward transmission. ■

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References

1. Wadowsky RM, Mietzner SM, Skoner DP, Doyle WJ, Fireman P. Effect of experimental influenza A virus infection on isolation of *Streptococcus pneumoniae* and other aerobic bacteria from the oropharynx of allergic and nonallergic adult subjects. *Infect Immun* 1995;63:1153–1157.
2. Mina MJ, McCullers JA, Klugman KP. Live attenuated influenza vaccine enhances colonization of *Streptococcus pneumoniae* and *Staphylococcus aureus* in mice. *MBio* 2014;5:e01040-13.
3. Jochems SP, Marcon F, Carniel BF, Holloway M, Mitsi E, Smith E, et al. Inflammation induced by influenza virus impairs human innate immune control of pneumococcus. *Nat Immunol* 2018;19:1299–1308. Available from: <https://www.biorxiv.org/content/10.1101/347161v1>.
4. Thors V, Christensen H, Morales-Aza B, Vipond I, Muir P, Finn A. The effects of live attenuated influenza vaccine on nasopharyngeal bacteria in healthy 2 to 4 year olds: a randomized controlled trial. *Am J Respir Crit Care Med* 2016;193:1401–1409.
5. Wolter N, Tempia S, Cohen C, Madhi SA, Venter M, Moyes J, et al. High nasopharyngeal pneumococcal density, increased by viral coinfection, is associated with invasive pneumococcal pneumonia. *J Infect Dis* 2014;210:1649–1657.
6. Jochems SP, Marcon F, Carniel BF, Holloway M, Mitsi E, Smith E, et al. Inflammation induced by influenza virus impairs human innate immune control of pneumococcus. *Nat Immunol* 2018;19:1299–1308.
7. Gritzfeld JF, Wright AD, Collins AM, Pennington SH, Wright AK, Kadioglu A, et al. Experimental human pneumococcal carriage. *J Vis Exp* 2013;(72):50115.
8. Ferreira DM, Neill DR, Bangert M, Gritzfeld JF, Green N, Wright AK, et al. Controlled human infection and rechallenge with *Streptococcus pneumoniae* reveals the protective efficacy of carriage in healthy adults. *Am J Respir Crit Care Med* 2013;187:855–864. [Published erratum appears in *Am J Respir Crit Care Med* 187:1153.]

9. McCullers JA, Rehg JE. Lethal synergism between influenza virus and *Streptococcus pneumoniae*: characterization of a mouse model and the role of platelet-activating factor receptor. *J Infect Dis* 2002;186: 341–350.

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Right Ventricular Load and Function in Chronic Thromboembolic Pulmonary Hypertension: Differences between Proximal and Distal Chronic Thromboembolic Pulmonary Hypertension

To the Editor:

The clinical presentation of chronic thromboembolic pulmonary hypertension (CTEPH) ranges from central pulmonary obstruction to more peripheral obstruction and small vessel vasculopathy. Location of CTEPH lesions does not seem to influence pulmonary artery pressures (1, 2), but its effect on right ventricular (RV) load and function is currently unknown. In this retrospective analysis, we aimed to determine the influence of proximal and distal vascular lesions on RV afterload and function, integrating static and pulsatile components of afterload and RV function parameters.

Methods

Seventy-five patients from our clinical registry were selected on the basis of the presence of high-quality computed tomography–pulmonary angiography, right heart catheterization (RHC), and cardiac magnetic resonance (CMR) imaging before initiation of treatment (maximum interval between investigations was 6 mo). According to computed tomography–pulmonary angiography, CTEPH was classified as either proximal (level I/II disease: lesions starting in the main or lobar arteries) or distal (level III–IV disease: lesions starting in the segmental and subsegmental) (3) for each side separately. Twenty-nine patients with asymmetrical lesions (proximal on one side and distal on the other side) were not included in the final analysis.

RHC and CMR were performed and analyzed as previously described (4):

- Pulmonary artery (PA) pulse pressure (mm Hg) = systolic pulmonary artery pressure (PAP) – diastolic PAP
- PA compliance (ml/mm Hg) = (Q/heart rate)/pulse pressure
- Resistance–compliance (RC) time (s) = $0.75 \cdot 10^{-3} \cdot$ pulmonary vascular resistance (PVR, $\text{dyn} \cdot \text{s} \cdot \text{cm}^{-5}$) \cdot compliance

This retrospective study did not fall within the scope of the Medical Research Involving Human Subjects Act, as confirmed by the Medical Ethics Review Committee of the VU University Medical Center (2017.025).

Table 1. Baseline Characteristics, Hemodynamic and Cardiac Magnetic Resonance Profile

Variable	Proximal CTEPH (n = 21)	Distal CTEPH (n = 25)	P Value
Age, yr	65 (50–70)	66 (52–73)	0.723
Male, n (%)	13 (62)	11 (44)	0.226
6MWD, m	339 (71), n = 19	445 (108), n = 19	0.001*
NT-proBNP, ng/L	1,745 (549–4,185)	446 (150–1,360)	0.032*
DL _{CO} , % predicted	61.9 (13.8), n = 18	75.7 (13.5), n = 19	0.004*
Right heart catheterization			
mPAP, mm Hg	49.1 (12.3)	45.5 (10.2)	0.286
PAWP, mm Hg	9.7 (4.3)	11.8 (2.7)	0.058
PVR, $\text{dyn} \cdot \text{s} \cdot \text{cm}^{-5}$	740 (544–1,011)	555 (419–775), n = 24	0.162
CI, L/min/m ²	2.1 (0.4)	2.4 (0.6), n = 24	0.077
mRAP, mm Hg	11.6 (5.3), n = 18	8.2 (2.9)	0.010*
PA pulse pressure, mm Hg	52.8 (15.8)	47.9 (11.6)	0.234
Stroke volume, ml	54.8 (19.6)	64.3 (22.9), n = 22	0.150
Heart rate, beats/min	80 (71–88)	73 (63–81), n = 22	0.088
Compliance, ml/mm Hg	1.11 (0.64–1.31)	1.34 (0.80–1.89), n = 22	0.098
Cardiac magnetic resonance			
RVEF, %	34.1 (12.9)	44.7 (15.2)	0.015*
RVEDVI, ml/m ²	95.3 (26.2)	80.5 (21.4)	0.041*
LVEF, %	59.8 (9.3)	66.9 (10.1)	0.018*
LVEDVI, ml/m ²	54.2 (14.4)	53.5 (13.6)	0.873
SVI, ml/m ²	29.9 (6.1)	33.6 (9.1), n = 24	0.120

Definition of abbreviations: 6MWD = 6-minute-walk distance; CI = cardiac index; CTEPH = chronic thromboembolic pulmonary hypertension; LVEDVI = left ventricular end-diastolic volume index; LVEF = left ventricular ejection fraction; mPAP = mean pulmonary artery pressure; mRAP = mean right atrial pressure; NT-proBNP = N-terminal pro-brain natriuretic peptide; PA = pulmonary artery; PAWP = pulmonary artery wedge pressure; PVR = pulmonary vascular resistance; RVEDVI = right ventricular end-diastolic volume index; RVEF = right ventricular ejection fraction; SVI = stroke volume index.

Data are presented as mean (SD), median (interquartile range), or number of patients (%). Data apply to all 21 and 25 patients per group unless otherwise stated. Statistical tests: unpaired *t* test, chi-square test, Fisher's exact test, and Mann-Whitney test.

*Statistical significance.

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