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# Unexpectedly High Prevalence of Cytomegalovirus DNAemia in Older Children and Adolescents With Perinatally Acquired Human Immunodeficiency Virus Infection

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**Background.** Older children and adolescents with perinatally acquired human immunodeficiency virus (PHIV) infection in Africa experience multiple comorbidities that are not typical of HIV-associated opportunistic infections, including growth impairment and chronic lung disease. We examined associations between plasma cytomegalovirus (CMV) DNA and lung function and growth.

**Methods.** Plasma CMV DNA loads were measured children aged 6–16 years with PHIV (n = 402) and HIV-uninfected controls (n = 224). The HIV-infected children were either newly diagnosed or known HIV infected and stable on antiretroviral therapy (ART) for >6 months. CMV DNA loads were measured using quantitative polymerase chain reaction. CMV DNAemia was modeled as a time-varying outcome using longitudinal mixed-effects logistic regression.

**Results.** At enrollment, CMV DNAemia  $\geq 1000$  copies/mL (defined as “clinically significant”) was detected in 5.8% of uninfected children, 14.7% of HIV-infected participants stable on ART, and 22.6% of HIV-infected ART-naive children ( $\chi^2 = 23.8$ ,  $P < .001$ ). The prevalence of CMV DNAemia  $\geq 1000$  copies/mL was associated with CD4 counts  $< 350$  cells/ $\mu$ L. Among HIV-infected ART-naive children, the presence of CMV DNAemia of  $\geq 1000$  copies/mL was independently associated with reduced lung function (adjusted odds ratio [aOR] = 3.23; 95% confidence interval [CI], 1.23–8.46;  $P = .017$ ). Among ART-treated children, stunting was associated with CMV DNAemia of  $\geq 1000$  copies/mL (aOR = 2.79; 95% CI, 0.97–8.02;  $P = .057$ ).

**Conclusions.** Clinically significant levels of CMV DNAemia were common in older children with PHIV, even those on ART, suggesting a role for inadequately controlled CMV infection in the pathogenesis of PHIV comorbidities in Africa.

**Keywords.** CMV; HIV; adolescent; chronic lung disease; stunting.

Before the widespread availability of antiretroviral therapy (ART), pediatric human immunodeficiency virus (HIV) in Africa was characterized by rapid disease progression, with <50% of children with perinatally acquired HIV infection (PHIV) surviving beyond 2 years [1]. At that time, the survival of HIV-infected children into adolescence seemed unlikely. However, it is now thought that one third of children with PHIV progress more slowly and can survive into their teens, even without ART [2]. The ART scale-up for PHIV in Africa has led to dramatic falls in mortality, leading to increasing numbers of ART-treated children reaching adolescence [3].

Long-term survival with PHIV, even for those on ART, is associated with significant health problems that are not typical of HIV-associated opportunistic infections or AIDS-defining illnesses [4]. Reports from sub-Saharan Africa have described significant morbidity in older children with PHIV [4], including chronic lung disease (CLD), cardiac abnormalities [5], growth retardation, pubertal delay, and neurocognitive disorders. Many of these PHIV comorbidities have not previously been reported outside Africa, and their pathogenesis is poorly understood.

The most serious comorbidity described in older children with PHIV is CLD, reported in more than 30% of HIV-infected children aged >10 years in HIV care but frequently misdiagnosed and treated as pulmonary tuberculosis [6]. The severity of CLD was only recently recognized, and little is known about its etiology [6, 7]. Lung function and radiological abnormalities in older children with PHIV show no associations with either treatment status or ART duration, suggesting that once established, this form of CLD does not respond to ART. Indeed, a recent study showed that 25% of older children with PHIV on ART (median treatment duration, 4.7 years; viral suppression

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below detection in 79%) reported symptoms of CLD, predominantly cough and dyspnea [8].

We hypothesized that cytomegalovirus (CMV) might contribute to the poor health of older children with PHIV, in view of the distinct epidemiology of CMV infection in sub-Saharan Africa. Most African infants acquire CMV infection in the first year of life, in contrast to developed countries where CMV infection occurs significantly later (mean age of 28.6 years in 1 large US study [9]). In the Gambia, 85% of infants acquired CMV infection by 1 year of age, increasing to 100% by 18 months [10]. HIV-uninfected infants generate potent cellular responses to CMV that are associated with viral control, although urinary CMV excretion may continue intermittently to age 5 years [11]. It is therefore highly likely that most African children with PHIV will acquire CMV coinfection in infancy.

CMV coinfection in ART-untreated infants with PHIV can lead to rapid disease progression [12, 13] and often infects the lungs, causing pneumonitis [14]. No studies have examined the long-term impact of infant CMV pneumonitis on subsequent lung function. However, it is plausible that repeated episodes of CMV pneumonitis or inflammation driven by subclinical CMV replication in HIV-infected children could lead to CLD.

We hypothesized that ART-untreated infants with PHIV who acquire CMV coinfection in early life fail to establish adequate immune control of CMV, leading to episodes of CMV reactivation and/or persistent CMV replication that may increase immune activation and drive some of the comorbidities described in adolescent survivors of PHIV.

## METHODS

### Study Setting and Participants

We studied 626 participants within 2 prospective cohort studies investigating the natural history of comorbidities that affect children with PHIV, including lung function and growth. The ZENITH cohort recruited children aged 6–16 years who were newly diagnosed with HIV following provider-initiated HIV testing in primary healthcare clinics in Harare, Zimbabwe [15]. The INHALE cohort recruited children in the same age group who had been taking ART for >6 months from an HIV clinic at Harare Children's Hospital, which served the same catchment population as the primary healthcare clinics. Inclusion/exclusion criteria are described elsewhere [16, 17]. PHIV diagnosis was made using criteria previously established in Zimbabwe [4].

Spirometry was performed at baseline using EasyOne World spirometers (ndd Medical Technologies, Zurich, Switzerland) according to American Thoracic Society standard procedures [7]. The highest forced expiratory volume in the first second of expiration (FEV1) and forced vital capacity (FVC) measurements, that is, the volume of air exhaled with maximum effort after a full inspiration, were used, with other indices recorded from the best trace (largest total of FEV1 and FVC). Using the

Global Lung Function Initiative 2012 reference ranges (accounting for height, sex, age, and ethnicity), lung function was considered abnormal if either FVC or FEV1:FVC ratio *z* scores were less than  $-1.64$  (below the 10th centile). Height was recorded at baseline for all participants, and HIV-infected participants were followed for 18 months, with clinical and laboratory examinations every 3 months. Blood samples were collected and plasma was separated on the same day at the Biomedical Research and Training Institute (BRTI) in Harare, then stored at  $-80^{\circ}\text{C}$  until required.

Children who tested HIV negative in the same age group as cohort participants joined the study as population controls and provided a single blood sample without further follow-up. Recruitment started in January 2013, and follow-up lasted until April 2015 (Figure 1).

### Extraction of CMV DNA From Plasma

Total viral nucleic acids were extracted at the BRTI laboratories in Harare from 200  $\mu\text{L}$  plasma using the QIAamp MinElute Virus Spin Kit (Qiagen, Hilden, Germany). Then, 60  $\mu\text{L}$  of total viral nucleic acids including CMV DNA were eluted and immediately stored at  $-80^{\circ}\text{C}$  for subsequent testing.

### CMV Detection and Quantification

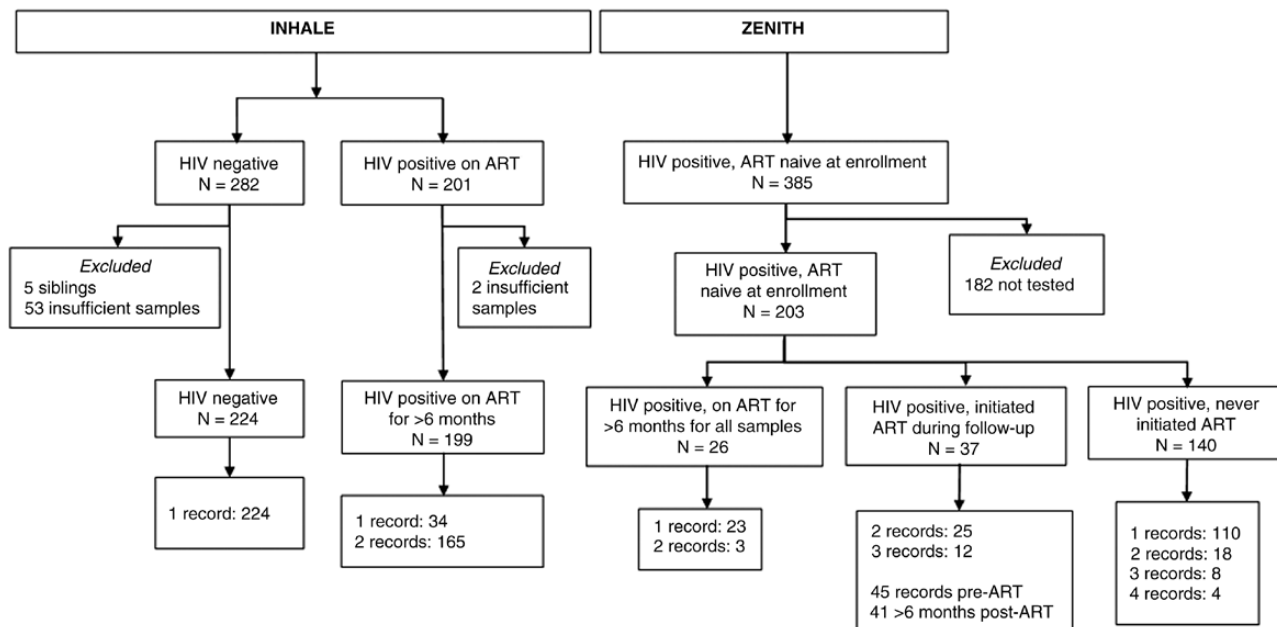
CMV DNAemia detection and quantification was performed by quantitative polymerase chain reaction (PCR) using the RealStar CMV PCR kit v1.0 (Altona Diagnostics, Hamburg, Germany). CMV DNA was quantified with the QuantStudio 3 Real-Time PCR System (Applied Biosystems, CA) (see Supplementary Methods for details).

### Statistical Analyses

“Clinically relevant” CMV viral load (VL) was defined as plasma CMV VL  $\geq 1000$  copies/mL. Stunting was defined as a height-for-age more than 2 standard deviations below the mean, using World Health Organization reference standards.  $\chi^2$  tests were used for univariate analysis. For multivariate analysis of time-varying outcomes, we used longitudinal mixed-methods logistic regression. ART-naive participants and those stable on ART were modeled separately. Data were analyzed using Stata v14.1 (StataCorp, College Station, TX). As a sensitivity analysis, we also examined associations of risk factors with prevalence of any detectable CMV DNA (see Supplementary Methods for details).

### Ethical Considerations

Informed consent (parent or guardian) and assent (participant) were obtained from all participants prior to enrollment. The Medical Research Council of Zimbabwe, the BRTI Institutional Review Board, and the Ethics Committees of Harare City Health Department, Harare Central Hospital, and the London School of Hygiene and Tropical Medicine approved the study.



**Figure 1.** Flow chart of study participants. Abbreviations: ART, antiretroviral therapy; HIV, human immunodeficiency virus; INHALE, INvestigation of Heart And Lung disEase among older children in Harare; ZENITH, Zimbabwe study for ENhancing Testing and Improving Treatment of HIV in children.

## RESULTS

### Baseline Characteristics of Study Participants

A total of 626 children and adolescents age 6–16 years were included. When the first sample was taken, 177 were HIV infected and ART naive, 225 were HIV infected on ART for >6 months, and 224 were uninfected controls (Figure 1). Baseline characteristics are provided in Table 1. During follow-up, 37 of the 177 originally ART-naive participants initiated ART and had a sample taken at least 6 months after starting therapy (Figure 1). Their pre-ART samples were included in the ART-naive analysis, and post-ART samples were included in the analysis of ART-experienced participants. There were 889 observations: 626 at baseline, 33 at 6 months, 37 at 12 months, 159 at 18 months, and 34 at 24 months. The median age at recruitment was similar between groups (11 years), and 50% were female. Stunting was more prevalent among HIV-positive children than their uninfected counterparts (32.5% vs 9.6%). Stunting was also associated with age; prevalence was 13.0% for children aged 6–9 years, 23.3% for children aged 10–12 years, and 39.6% for children aged >13 years ( $\chi^2 = 37.5$ ,  $P < .001$ ). Abnormal lung function (defined as described above) was common, ranging from 15.5% in the uninfected group to 24.1% in the group stable on ART and 29.4% in the ART-naive HIV-positive group.

CMV DNAemia was detected in 8.9% of uninfected children, 24.0% of HIV-positive participants established on ART, and 38.4% of the HIV-positive ART-naive children. At baseline, the prevalence of clinically relevant CMV DNAemia (CMV VL

$\geq 1000$  copies/mL) in the uninfected, HIV-positive on ART, and HIV-positive ART-naive groups was 5.8%, 14.7%, and 22.6%, respectively ( $\chi^2 = 23.8$ ,  $P < .001$ ; Table 1).

### Association Between CD4 Count and CMV Viral Load

Of the 55 HIV-positive participants with clinically significant CMV DNAemia at enrollment, 52% were on ART for >2 years before joining the cohort. The proportion of children with clinically relevant CMV viremia was inversely associated with CD4 count at recruitment in the ART-stable group (Table 1). In a multivariate analysis controlling for repeated measures and adjusting for ART status, age, and sex, CD4 count <350 cells/ $\mu$ L was associated with more than twice the odds of CMV VL >1000 copies/mL (Table 2). After adjusting for CD4 count, age, and sex, no longer being on ART showed significant protection against high CMV DNAemia; however, each additional year on ART was associated with 0.84 the odds of CMV VL  $\geq 1000$  copies among ART-treated participants (Table 3), even after adjusting for CD4 count. HIV RNA <1000 copies/mL was not associated with CMV DNAemia among participants on ART (Table 3).

### Stunting Is Associated With HIV and High CMV DNAemia

At baseline, 29.2% of HIV-infected ART-naive participants and 34.3% of those on ART were stunted compared to 9.6% of HIV-uninfected participants (Table 1). In ART-naive HIV-infected participants, the prevalence of CMV VL  $\geq 1000$  copies/mL was higher among stunted than nonstunted participants (40.8% vs 14.3%,  $\chi^2 = 14.2$ ,  $P < .001$ ). In participants who were on ART or

**Table 1. Baseline Characteristics of the Study Population and Prevalence of High Cytomegalovirus DNAemia**

Characteristic	Category	HIV Negative <sup>a</sup>		HIV Positive on ART <sup>a</sup>		HIV Positive ART-Naive <sup>a</sup>	
		N (%)	CMV ≥1000 n/N (%)	N (%)	CMV ≥1000 n/N (%)	N (%)	CMV ≥1000 n/N (%)
Gender	Male	108 (48.2)	5/108 (4.6)	114 (50.7)	17/114 (14.9)	90 (50.9)	19/90 (21.1)
	Female	116 (51.8)	8/116 (6.9)	111 (49.3)	16/111 (14.4)	87 (49.1)	21/87 (24.1)
Age (years)	6–9	83 (37.1)	9/83 (10.8)	72 (32.0)	10/72 (13.9)	73 (41.2)	16/73 (23.3)
	10–12	72 (32.1)	3/72 (4.2)	92 (40.9)	13/92 (14.1)	58 (32.8)	11/58 (19.0)
	13–16	69 (30.8)	1/69 (1.5)	61 (27.1)	10/61 (16.4)	46 (26.0)	12/46 (26.1)
Height for age <sup>b</sup>	Stunted	21 (9.6)	1/21 (4.8)	77 (34.3)	12/77 (15.6)	49 (29.2)	20/49 (40.8)
	Not stunted	198 (90.4)	12/198 (6.1)	147 (65.7)	21/147 (14.3)	119 (70.8)	17/119 (14.3)
Lung function <sup>c</sup>	Normal	158 (84.5)	10/158 (6.3)	148 (75.9)	23/148 (15.5)	108 (63.3)	22/108 (20.4)
	Reduced forced vital capacity	22 (11.8)	1/22 (4.6)	38 (19.5)	4/38 (10.5)	29 (17.5)	9/29 (31.0)
	Obstruction	7 (3.8)	0 (0.0)	9 (4.6)	1/9 (11.1)	16 (9.6)	5/16 (31.3)
CD4 count (cells/μL) <sup>d</sup>	≤349	...	...	23 (10.5)	6/23 (26.1)	48 (35.1)	11/48 (22.9)
	350–499	...	...	39 (17.7)	5/39 (12.8)	22 (14.9)	3/22 (13.6)
	≥500	...	...	158 (71.8)	20/158 (12.7)	67 (50.0)	13/67 (19.4)
CMV VL (copies/mL of plasma)	Undetectable	204 (91.1)	...	171 (76.0)	...	109 (61.6)	...
	1–999	7 (3.1)	...	21 (9.3)	...	28 (15.8)	...
	1000–9999	13 (5.8)	...	27 (12.0)	...	34 (19.2)	...
	≥10000	0 (0.0)	...	6 (2.7)	...	6 (3.4)	...
HIV RNA (copies/mL of plasma)	≤999	...	...	180 (84.1)	25/178 (14.0)	...	...
	1000–9999	...	...	6 (2.8)	0/6 (0.0)	...	...
	≥10000	...	...	28 (13.1)	5/28 (17.9)	...	...

Abbreviations: ART, antiretroviral therapy; CMV, cytomegalovirus; HIV, human immunodeficiency virus; VL, viral load.

<sup>a</sup>Of the total number of participants, 224 were HIV negative, 225 HIV positive on ART, and 177 HIV positive/ART-naive.

<sup>b</sup>Data on stunting were missing for 5 HIV-uninfected and 9 HIV-positive/ART-naive participants.

<sup>c</sup>The numbers of children with successful and interpretable lung function test data (spirometry test) at recruitment were 187 (HIV negative), 192 (HIV positive on ART), and 153 (HIV positive/ART-naive).

<sup>d</sup>CD4 count was not measured for the uninfected group, and the data were available for 357/402 HIV-positive participants.

HIV negative, the prevalence of CMV VL ≥1000 copies/mL was similar among stunted and nonstunted participants (Table 1).

#### Association of Lung Function With High CMV DNAemia

A total of 89 participants did not have spirometry measurements of sufficient quality to allow lung function assessment. Both reduced FVC and spirometric features of obstruction were more common in HIV-infected than HIV-uninfected participants, particularly obstruction, which was more prevalent among ART-naive participants (Table 1). At baseline, 29.1%

of stunted children compared to 20.8% of nonstunted children had abnormal lung function (reduced FVC or obstruction) in all 3 groups combined ( $\chi^2 = 3.85$ ,  $P = .05$ ). Only in HIV-positive ART-naive participants was high CMV DNAemia prevalence greater among those with abnormal lung function (Table 1).

In a longitudinal analysis of ART-naive HIV-positive participants, reduced FVC was independently associated with high CMV DNAemia after adjusting for gender, age, CD4 count, and individual-level clustering (Table 4). Stunting and obstructive spirometry were not associated with CMV VL. Similar analysis

**Table 2. Multivariate Model of Risk Factors for High Cytomegalovirus DNAemia in All Human Immunodeficiency Virus–positive Participants**

Characteristic	Category	Univariate <sup>a</sup>		Multivariate <sup>a</sup>	
		OR (95% CI)	P Value	OR (95% CI)	P Value
CD4 count (cells/μL)	≥350	1	...	1	...
	<350	2.70 (1.32, 5.51)	.006	2.64 (1.25, 5.59)	.011
Gender	Male	1	...	1	...
	Female	0.85 (0.48, 1.49)	.563	0.86 (0.46, 1.58)	.618
Age (years; ref: 6 years)		0.96 (0.87, 1.06)	.425	0.94 (0.83, 1.06)	.313
ART status	Naive	1	...	1	...
	>6 months on ART	0.43 (0.24, 0.78)	.005	0.61 (0.32, 1.16)	.132

Abbreviations: ART, antiretroviral therapy; CI, confidence interval; OR, odds ratio; ref, reference group, controlling for clustering by individual.

<sup>a</sup>A total of 402 participants contributing 665 records were used in univariate analysis, while 367 participants contributing 591 records were used in model 1.

**Table 3. Multivariate Model of Risk Factors for High Cytomegalovirus DNAemia in Participants on Antiretroviral Therapy**

Characteristic	Category	Univariate <sup>a</sup>		Multivariate: <i>A Priori</i> Only <sup>a</sup>		Multivariate: <i>A Priori</i> or Associated With Outcome <sup>a</sup>	
		OR (95% CI)	PValue	OR (95% CI)	PValue	OR (95% CI)	PValue
Years since antiretroviral therapy initiation (ref: 0)	...	0.81 (0.69, 0.95)	.010	0.83 (0.69, 0.99)	.035	0.84 (0.70, 1.00)	.050
CD4 count (cells/μL)	≥350	1	...	1	...	1	...
	<350	3.92 (1.20, 12.77)	.023	3.75 (1.09, 12.94)	.036	4.39 (1.20, 16.04)	.025
Gender	Male	1	...	1	...	1	...
	Female	0.64 (0.27, 1.51)	.311	0.69 (0.27, 1.74)	.430	0.80 (0.31, 2.10)	.655
Age (y; ref: 6 y)	...	0.99 (0.85, 1.15)	.884	0.94 (0.79, 1.12)	.469	0.91 (0.75, 1.09)	.305
Height for age	Normal	1	...	...	...	1	...
	Stunted	3.01 (1.17, 7.77)	.022	...	...	2.79 (0.97, 8.02)	.057
Lung function	Normal	1	...	...	...	...	...
	Reduced forced vital capacity	1.22 (0.39, 3.80)	.727	...	...	...	...
	Obstructed	0.41 (0.03, 6.52)	.527	...	...	...	...
HIV RNA (337 records)	≥1000	1.01 (0.25, 4.03)	.988	...	...	...	...
	<1000	1	...	...	...	...	...

Abbreviations: CI, confidence interval; OR, odds ratio; ref, reference group, controlling for clustering by individual.

<sup>a</sup>A total of 262 participants contributing 434 records were used in univariate analysis; 256 participants contributing 411 records were used in model 1, and 251 participants contributing 405 records were used in model 2.

in HIV-positive participants on ART revealed that CD4 count <350 cells/μL and stunting (albeit weakly) were independently associated with high CMV DNAemia, while longer time on ART was associated with lower odds of high CMV DNAemia (Table 3). Abnormal lung function was not included in the model as there was no evidence it was associated with outcomes of participants on ART in univariate analysis.

When the outcome was defined as any detectable CMV, lung function and stunting were not associated with CMV DNAemia in multivariate analysis among ART-naive or ART-treated participants. Being on ART for at least 6 months was associated with lower odds

of detectable CMV DNA in plasma (adjusted odds ratio, 0.54; 95% confidence interval, 0.31–0.93;  $P = .026$ ; Supplementary Table 1).

Analysis of longitudinal samples taken 18 months apart from a subset of 159 children and adolescents with PHIV showed that 15.7% (20/127) of participants with undetectable CMV DNA at baseline had detectable levels of CMV DNA in their 18 month samples and vice versa (Table 5).

## DISCUSSION

We investigated the hypothesis that poorly controlled CMV coinfection, acquired in infancy in most African children, could

**Table 4. Multivariate Model of Risk Factors for High Cytomegalovirus DNAemia in Antiretroviral Therapy-naive Human Immunodeficiency Virus-positive Participants**

Characteristic	Category	Univariate <sup>a</sup>		Multivariate: <i>A priori</i> Only <sup>a</sup>		Multivariate: <i>A priori</i> or Associated With Outcome <sup>a</sup>	
		OR (95% CI)	PValue	OR (95% CI)	PValue	OR (95% CI)	PValue
CD4 count (cells/μL)	≥350	1	...	1	...	1	...
	<350	1.47 (0.67, 3.24)	.340	1.55 (0.68, 3.54)	.297	1.05 (0.40, 2.78)	.916
Gender	Male	1	...	1	...	1	...
	Female	1.10 (0.56, 2.16)	.793	1.15 (0.54, 2.45)	.714	1.17 (0.49, 2.79)	.723
Age (y; ref: 6 y)	...	0.97 (0.85, 1.10)	.619	0.95 (0.81, 1.11)	.514	0.94 (0.78, 1.14)	.552
Height for age	Normal	1	...	...	...	1	...
	Stunted	4.23 (1.47, 12.16)	.008	...	...	2.02 (0.78, 5.27)	.150
Lung function	Normal	1	...	...	...	1	...
	Reduced forced vital capacity	2.11 (0.89, 4.98)	.088	...	...	3.23 (1.23, 8.46)	.017
	Obstructed	1.47 (0.49, 4.41)	.485	...	...	2.04 (0.48, 8.70)	.336

Abbreviations: CI, confidence interval; OR, odds ratio; ref, reference group, controlling for clustering by individual.

<sup>a</sup>A total of 177 participants contributing 231 records were used in univariate analysis, 143 participants contributing 180 records were used in model 1, and 138 participants contributing 169 records were used in model 2.

**Table 5. Frequency of Cytomegalovirus DNAemia in a Subset of Longitudinal Samples From the Human Immunodeficiency Virus–positive Group**

		CMV DNA at 18 Months		
		Detected	Not Detected	Total <sup>a</sup>
CMV DNA at baseline	Detected	11	21	32
	Not detected	20	107	127
Total		31	128	159

Abbreviation: CMV, cytomegalovirus.

<sup>a</sup>Number of participants with 2 samples taken 18 months apart. The presence and quantity of CMV DNA was measured in both samples (baseline and follow-up) at the same time in a single plate to minimize intra- and interassay variability.

contribute to the comorbidities of older children and adolescents with PHIV. We found a surprisingly high prevalence of CMV DNAemia in older children with PHIV, both those with a new HIV diagnosis who had not yet started ART and those regarded as stable on ART. CMV DNAemia >1000 copies/mL was significantly associated with 2 major PHIV comorbidities, stunting and reduced lung function.

CLD in older children with PHIV was first described in Zimbabwe and subsequently reported in Malawi [18] and Kenya [19]. Radiological features strongly suggest obliterative bronchiolitis (OB) as the main cause [7], in which inflammation of the small airway leads ultimately to partial or complete bronchiolar obstruction. In high-income settings, OB is most often associated with allograft immune responses following lung or hematopoietic stem cell transplantation [20]; subclinical CMV infection has been linked with increased OB risk after lung transplantation [21]. Reports from the southern hemisphere describe OB following severe respiratory infections in early childhood [22]. As the epidemiology of CMV infection is distinct between sub-Saharan Africa and developed countries, we postulated that early CMV infection could contribute to the striking comorbidities associated with PHIV in Africa [4].

The trajectory of CMV viremia and shedding after primary infection remains poorly understood. Children infected in infancy were reported to secrete CMV in urine until the age of 5 years. However, a large cross-sectional survey in the United States showed that 9.7% of the CMV immunoglobulin (Ig) G-positive population aged 6–49 years were excreting CMV in urine [23]. Urinary CMV shedding was most common in the youngest participants (26.6% in children aged 6–11 years) but was also detected in adolescents (6.5% in those aged 12–19 years). Even less is known about the dynamics of CMV viremia. In one study of hospitalized children aged 3 weeks to 2 years in Lusaka, Zambia, CMV was detected in the blood of 41% and was associated with being HIV infected, underweight, and presenting with meningitis [24]. Two age peaks of CMV DNAemia were noted, particularly in HIV-infected children, one in the first year of life and a second around 18–20 months of age. This was interpreted as either CMV reactivation or potentially reinfection. CMV

DNAemia is rare in healthy adults (CMV DNA was found in only 2/416 CMV IgG-positive blood donors in the United States [25]). However, CMV is frequently reactivated during pregnancy and lactation, particularly in HIV-infected women, leading to viral shedding in the cervix and breastmilk. In HIV-uninfected pregnant women, CMV DNA is rarely detected in blood but may be found in the cervix of 37% CMV IgG-positive women, particularly during the third trimester [26]. In HIV-infected Kenyan women taking ART to prevent mother-to-child HIV transmission, CMV was detected in the cervix and breastmilk of 66% and 99% of participants, respectively, although CMV was only found in blood in 4.8% [27]. Increased CMV shedding in the cervix and breastmilk of HIV-infected women who are close to delivery increases the risk of both congenital and early CMV infection in their infants [27]. Although ART used to prevent mother-to-child HIV transmission during pregnancy reduces the risk of congenital CMV infection [13], the reduction in early postnatal CMV transmission is only modest [28].

The burden of severe opportunistic CMV infection, causing common AIDS-defining conditions such as retinitis, colitis, and esophagitis, has decreased substantially with the availability of ART. Nevertheless, studies of HIV-infected adults in the developed world have shown that subclinical CMV DNAemia (ie, CMV DNA detected in blood without clinical evidence of CMV disease) strongly predicts accelerated disease progression and early mortality, even in patients taking ART [29]. Increased levels of CMV antibodies or T-cell responses (potentially driven by CMV reactivation) are associated with both elevated immune activation [30] and the non-AIDS-defining comorbidities increasingly seen in the aging population of HIV-infected people on long-term ART [31]. There are fewer studies in Africa, but one study of South African miners with relatively early HIV disease in the pre-ART era reported that CMV DNAemia (5.2% prevalence) was associated with a 3-fold increase in mortality [32]. In that study, higher CMV VLs ( $\geq 1000$  copies/mL, our cut-off for clinically significant CMV DNAemia) were linked with significantly higher mortality (adjusted hazard ratio, 3.65) [32].

A recent metaanalysis showed that early CMV coinfection led to accelerated disease progression in ART-untreated infants with PHIV [13]. However, there is little information about the impact of CMV coinfection on HIV-infected children taking cART, particularly in Africa. Maternal CMV DNAemia close to delivery was associated with increased mortality of both mother and baby in a Kenyan cohort with limited ART access [33]. HIV and CMV plasma VLs closely correlated in the HIV-CMV-coinfected infants in this study, thought to reflect CMV-driven immune activation that fueled HIV replication in activated T cells [34, 35]. As CMV replication (like HIV) is enhanced by inflammation, the virus has developed strategies to promote immune activation, including direct upregulation of proinflammatory cytokines such as interleukin (IL)-1 $\beta$ , tumor necrosis factor- $\alpha$ , and IL-6 [36]. The association of CMV infection with inflammation is more

marked in HIV-infected patients [37]. CMV was recently shown to be replicating in the gut mucosa of HIV-infected adults (even those on ART); this was associated with disruption of mucosal integrity, translocation of gut microbial products, and increased inflammation [38]. A trial of the anti-CMV drug valganciclovir in HIV-infected adults with incomplete immune reconstitution on ART showed that CMV suppression was significantly associated with reduced T-cell activation [39].

Our study has strengths and some limitations. We studied a large well-characterized group of HIV-infected older children with PHIV, both treated and untreated, as well as age- and sex-matched controls from the same population. An important limitation is the lack of information about HIV RNAs in the ART-naive participants at enrollment (not routinely measured at the time in Zimbabwe.) The INHALE participants described as stable on ART may have elevated HIV RNAs because of poor adherence, which is characteristic of HIV-infected adolescents [40]. However, HIV RNA data, available from 337 of 434 samples from participants on ART, showed no association between HIV RNA and CMV DNAemia in univariate analysis. Last, our data do not allow us to determine the direction of associations, so we cannot conclude that CMV is a cause of stunting or CLD. An alternative interpretation would be that all 3 are independent but causally unrelated markers of progressive PHIV disease.

Additional studies are needed to establish the potential role of CMV reactivation in the pathogenesis of the serious complications of PHIV in Africa and to determine whether or not specific antiviral drugs such as valganciclovir might be useful in these conditions.

### Supplementary Data

Supplementary materials are available at *Clinical Infectious Diseases* online. Consisting of data provided by the authors to benefit the reader, the posted materials are not copyedited and are the sole responsibility of the authors, so questions or comments should be addressed to the corresponding author.

### Notes

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### References

- Newell ML, Coovadia H, Cortina-Borja M, Rollins N, Gaillard P, Dabis F; Ghent International AIDS Society Working Group on HIV Infection in Women and Children. Mortality of infected and uninfected infants born to HIV-infected mothers in Africa: a pooled analysis. *Lancet* **2004**; 364:1236–43.
- Ferrand RA, Corbett EL, Wood R, et al. AIDS among older children and adolescents in Southern Africa: projecting the time course and magnitude of the epidemic. *AIDS* **2009**; 23:2039–46.
- Slogrove AL, Sohn AH. The global epidemiology of adolescents living with HIV: time for more granular data to improve adolescent health outcomes. *Curr Opin HIV AIDS* **2018**; 13:170–8.
- Ferrand RA, Bandason T, Musvaire P, et al. Causes of acute hospitalization in adolescence: burden and spectrum of HIV-related morbidity in a country with an early-onset and severe HIV epidemic: a prospective survey. *PLoS Med* **2010**; 7:e1000178.
- Miller RF, Kaski JP, Hakim J, et al. Cardiac disease in adolescents with delayed diagnosis of vertically acquired HIV infection. *Clin Infect Dis* **2013**; 56:576–82.
- Ferrand RA, Desai SR, Hopkins C, et al. Chronic lung disease in adolescents with delayed diagnosis of vertically acquired HIV infection. *Clin Infect Dis* **2012**; 55:145–52.
- Desai SR, Nair A, Rylance J, et al. Human immunodeficiency virus-associated chronic lung disease in children and adolescents in Zimbabwe: chest radiographic and high-resolution computed tomographic findings. *Clin Infect Dis* **2018**; 66:274–81.
- Rylance J, Mchugh G, Metcalfe J, et al. Chronic lung disease in HIV-infected children established on antiretroviral therapy. *AIDS* **2016**; 30:2795–803.
- Colugnati FA, Staras SA, Dollard SC, Cannon MJ. Incidence of cytomegalovirus infection among the general population and pregnant women in the United States. *BMC Infect Dis* **2007**; 7:71.
- Kaye S, Miles D, Antoine P, et al. Virological and immunological correlates of mother-to-child transmission of cytomegalovirus in the Gambia. *J Infect Dis* **2008**; 197:1307–14.
- Miles DJ, van der Sande M, Jeffries D, et al. Cytomegalovirus infection in Gambian infants leads to profound CD8 T-cell differentiation. *J Virol* **2007**; 81:5766–76.
- Kovacs A, Schluchter M, Easley K, et al. Cytomegalovirus infection and HIV-1 disease progression in infants born to HIV-1-infected women. *Pediatric Pulmonary and Cardiovascular Complications of Vertically Transmitted HIV Infection Study Group*. *N Engl J Med* **1999**; 341:77–84.
- Ellington SR, Clarke KE, Kourtis AP. Cytomegalovirus infection in human immunodeficiency virus (HIV)-exposed and HIV-infected infants: a systematic review. *J Infect Dis* **2016**; 213:891–900.
- Zampoli M, Morrow B, Hsiao NY, Whitelaw A, Zar HJ. Prevalence and outcome of cytomegalovirus-associated pneumonia in relation to human immunodeficiency virus infection. *Pediatr Infect Dis J* **2011**; 30:413–7.
- Ferrand RA, Simms V, Dauya E, et al. The effect of community-based support for caregivers on the risk of virological failure in children and adolescents with HIV in Harare, Zimbabwe (ZENITH): an open-label, randomised controlled trial. *Lancet Child Adolesc Health* **2017**; 1:175–83.
- Ferrand RA, Meghji J, Kidia K, et al. The effectiveness of routine opt-out HIV testing for children in Harare, Zimbabwe. *J Acquir Immune Defic Syndr* **2015**; 17:e24–29.
- Majonga ED, Rehman AM, McHugh G, et al. Echocardiographic reference ranges in older children and adolescents in sub-Saharan Africa. *Int J Cardiol* **2017**; 248:409–13.
- Mwalukomo T, Rylance SJ, Webb EL, et al. Clinical characteristics and lung function in older children vertically infected with human immunodeficiency virus in Malawi. *J Pediatric Infect Dis Soc* **2016**; 5:161–9.
- Attia EF, Miller RF, Ferrand RA. Bronchiectasis and other chronic lung diseases in adolescents living with HIV. *Curr Opin Infect Dis* **2017**; 30:21–30.
- Jaramillo A, Fernández FG, Kuo EY, Trulock EP, Patterson GA, Mohanakumar T. Immune mechanisms in the pathogenesis of bronchiolitis obliterans syndrome after lung transplantation. *Pediatr Transplant* **2005**; 9:84–93.
- Paraskeva M, Bailey M, Levvey BJ, et al. Cytomegalovirus replication within the lung allograft is associated with bronchiolitis obliterans syndrome. *Am J Transplant* **2011**; 11:2190–6.
- Fischer GB, Sarria EE, Mattiello R, Mocelin HT, Castro-Rodriguez JA. Post infectious bronchiolitis obliterans in children. *Paediatr Respir Rev* **2010**; 11:233–9.



23. Amin MM, Bialek SR, Dollard SC, Wang C. Urinary cytomegalovirus shedding in the United States: the National Health and Nutrition Examination Surveys, 1999–2004. *Clin Infect Dis* **2018**;67:587–92.
24. Tembo J, Kabwe M, Chilukutu L, et al. Prevalence and risk factors for betaherpesvirus DNAemia in children >3 weeks and <2 years of age admitted to a large referral hospital in sub-Saharan Africa. *Clin Infect Dis* **2015**; 60:423–31.
25. Roback JD, Drew WL, Laycock ME, Todd D, Hillyer CD, Busch MP. CMV DNA is rarely detected in healthy blood donors using validated PCR assays. *Transfusion* **2003**; 43:314–21.
26. Shen CY, Chang SF, Chao MF, et al. Cytomegalovirus recurrence in seropositive pregnant women attending obstetric clinics. *J Med Virol* **1993**; 41:24–9.
27. Slyker J, Farquhar C, Atkinson C, et al. Compartmentalized cytomegalovirus replication and transmission in the setting of maternal HIV-1 infection. *Clin Infect Dis* **2014**; 58:564–72.
28. Richardson BA, John-Stewart G, Atkinson C, et al. Vertical cytomegalovirus transmission from HIV-infected women randomized to formula-feed or breast-feed their infants. *J Infect Dis* **2016**; 213:992–8.
29. Deayton JR, Prof Sabin CA, Johnson MA, Emery VC, Wilson P, Griffiths PD. Importance of cytomegalovirus viraemia in risk of disease progression and death in HIV-infected patients receiving highly active antiretroviral therapy. *Lancet* **2004**; 363:2116–21.
30. Patel EU, Gianella S, Newell K, et al. Elevated cytomegalovirus IgG antibody levels are associated with HIV-1 disease progression and immune activation. *AIDS* **2017**; 31:807–13.
31. Parrinello CM, Sinclair E, Landay AL, et al. Cytomegalovirus immunoglobulin G antibody is associated with subclinical carotid artery disease among HIV-infected women. *J Infect Dis* **2012**; 205:1788–96.
32. Fielding K, Koba A, Grant AD, et al. Cytomegalovirus viremia as a risk factor for mortality prior to antiretroviral therapy among HIV-infected gold miners in South Africa. *PLoS One* **2011**; 6:e25571.
33. Slyker JA, Lohman-Payne BL, Rowland-Jones SL, et al. The detection of cytomegalovirus DNA in maternal plasma is associated with mortality in HIV-1-infected women and their infants. *AIDS* **2009**; 23:117–24.
34. Slyker JA, Lohman-Payne BL, John-Stewart GC, et al. Acute cytomegalovirus infection in Kenyan HIV-infected infants. *AIDS* **2009**; 23:2173–81.
35. Slyker JA, Rowland-Jones SL, Dong T, et al. Acute cytomegalovirus infection is associated with increased frequencies of activated and apoptosis-vulnerable T cells in HIV-1-infected infants. *J Virol* **2012**; 86:11373–9.
36. Almeida GD, Porada CD, St Jeor S, Ascensao JL. Human cytomegalovirus alters interleukin-6 production by endothelial cells. *Blood* **1994**; 83:370–6.
37. Lurain NS, Hanson BA, Hotton AL, Weber KM, Cohen MH, Landay AL. The association of human cytomegalovirus with biomarkers of inflammation and immune activation in HIV-1-infected women. *AIDS Res Hum Retroviruses* **2016**; 32:134–43.
38. Maidji E, Somsouk M, Rivera JM, Hunt PW, Stoddart CA. Replication of CMV in the gut of HIV-infected individuals and epithelial barrier dysfunction. *PLoS Pathog* **2017**; 13:e1006202.
39. Hunt PW, Martin JN, Sinclair E, et al. Valganciclovir reduces T cell activation in HIV-infected individuals with incomplete CD4+ T cell recovery on antiretroviral therapy. *J Infect Dis* **2011**; 203:1474–83.
40. Hudelson C, Cluver L. Factors associated with adherence to antiretroviral therapy among adolescents living with HIV/AIDS in low- and middle-income countries: a systematic review. *AIDS Care* **2015**; 27:805–16.