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Dengue vaccine: reliably determining previous exposure

Dengue fever is the most prevalent and widespread mosquito-borne viral disease, and can only be countered by integrated prevention and control strategies, including sustained vector control programmes, the best evidence-based clinical care, and vaccination.

The first dengue vaccine, CYD-tetravalent dengue vaccine (CYD-TDV) or Dengvaxia (Sanofi Pasteur, Lyon, France), is licensed in 20 countries. Initial findings from two large phase 3 clinical trials have shown good but incomplete protection, in particular against severe dengue and dengue disease that requires hospital admission.1 2 In November, 2017, Sanofi Pasteur announced the results from additional studies showing that the vaccine has a differential performance in individuals who have previously been infected by dengue virus (seropositive) versus those without previous dengue virus infection (seronegative).3 Vaccine efficacy against laboratory-confirmed symptomatic dengue virus infection was high among individuals who were seropositive at baseline and aged at least 9 years (76%, 95% CI 63·9–84·0), but much lower among participants who were seronegative at baseline (38%, –0·9 to 62·9). Furthermore, vaccination of individuals who are seronegative increases their risk of severe dengue or dengue that requires hospital admission.4 Subsequently, in April, 2018, WHO’s Strategic Advisory Group of Experts recommended that in countries considering the introduction of vaccination with CYD-TDV, pre-vaccination screening should be preferred to assess dengue virus serostatus, and only people who are dengue seropositive should be vaccinated. To this end, WHO also encouraged the urgent development of rapid diagnostic tests (RDTs) to establish serostatus.

Because dengue viruses are of the genus Flavivirus, composed of genetically, structurally, and antigenically related viruses, such as Zika virus, yellow fever virus, and tick-borne encephalitis virus, antibody detection tests have high cross-reactivity and consequently poor reliability for the diagnosis of past dengue virus infection.5 With large flavivirus-exposed populations in Latin America, Asia, and Africa, a rapidly growing population of travellers from and to endemic areas,4 and increased vaccination coverage against yellow fever virus, tick-borne encephalitis virus, and Japanese encephalitis virus in travellers and in certain endemic regions, it is evident that establishing serostatus is extremely challenging. Many commercial immunoglobulin (Ig)G-containing point-of-care tests (POCTs) and dengue IgG ELISA assays are available. For the purpose of pre-vaccination screening, the assay with the highest sensitivity and specificity would be the desirable option. Low sensitivity would result in under-vaccinating individuals who are truly seropositive and who would benefit from the vaccine, whereas low specificity would lead to falsely vaccinating people who are truly seronegative, putting them at risk of severe dengue during the next natural infection with dengue virus.

The plaque reduction neutralisation test (PRNT) is still considered the gold standard for establishing serostatus, but requires specific laboratory and technical capacity, and is labour intensive, costly, and time consuming. Although dengue IgG ELISA correlates reasonably well with the results of a PRNT, the test has a lower sensitivity and specificity than PRNT, although the studies6 7 8 that showed this lower sensitivity and specificity were done before Zika virus became a widespread problem in the Americas. Dengue IgG ELISA requires about 2·5 h of laboratory time, excluding the time needed for sample transportation to the laboratory, batch analysis, and reporting to the clinician, which requires two visits for the vaccinees. Point-of-care testing that uses RDTs provides the vaccine recipient with a result within 15–30 min, and can be done in an outpatient or outreach setting, such as schools and care facilities, using a finger prick sample. Thus, a decision on vaccination eligibility can be made during the same visit, thereby ensuring a reasonable vaccine uptake. POCTs generally have lower sensitivity and specificity than dengue IgG ELISAs. However, this low sensitivity and specificity needs to be weighed up against the faster speed of testing, lower cost, and improved accessibility outside specialised laboratories compared with IgG ELISAs.

As available RDTs were mainly developed for the purpose of diagnosing acute dengue virus infection, further efforts would be justified to fine-tune such RDTs to increase sensitivity and specificity for diagnosing serostatus. To increase the sensitivity of dengue RDTs to detect previous dengue virus infection, several modifications could be contemplated, one of which would be recalibration by changing the concentration...
of the IgG capture antigen or detection reagent to lower the limit of anti-dengue IgG detection. To address cross-reactivity with other flaviviruses, particularly Zika, other modifications should be considered to improve specificity.

Diagnostic methods for flavivirus antibody detection use the entire virus particle, recombinant surface E-protein, or NS1 protein as antibody-capturing antigens. Epitopes with high aminocacid sequence homology among serotypes or flaviviruses can trigger cross-reactive responses, but type-specific antibodies directed towards unique E and NS1 epitopes are also raised.9,11 The art of responses, but type-specific antibodies directed towards among serotypes or flaviviruses can trigger cross-reactive epitopes with high aminoacid sequence homology use the entire virus particle, recombinant surface specificity. other modifications should be considered to improve cross-reactivity with other flaviviruses, particularly Zika, to flavivirus. The epitope specificity and the abundance subsets of antibodies in people who have been exposed sufficiently high sensitivity when aiming to detect specific of such subsets of antibodies might also differ between populations and individuals as a result of differences in genetic background, co-circulating pathogens, vaccination, history of natural infection, and timing of sampling. Combining multiple type-specific epitopes and antigen-antibody crosslinking could potentially help address this issue. Evaluation platforms and access to well characterised samples to accelerate test development and access to market are urgently needed.

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