

## 1 Rethinking Schistosomiasis Vaccine Development: Synthetic Vesicles

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### 14 Abstract

15 There is currently no vaccine against schistosomiasis. With few *Schistosoma* vaccine candidates in  
16 clinical trials, unexplored antigens from the vulnerable schistosomulum should be considered as possible  
17 vaccine candidates. In addition, we suggest developing synthetic vesicles as a new delivery vehicle and  
18 adjuvant for immunoprophylactic schistosomula vaccine candidates.

19 **Keywords:** *Schistosoma*, schistosomula, synthetic vesicles, vaccine candidates, delivery, adjuvant

### 20 It is Time to Think Elimination

21 Schistosomiasis is one of the most prevalent parasitic diseases worldwide. Treatment of schistosomiasis  
22 in populations at risk with a single dose of praziquantel annually has not prevented transmission of  
23 *Schistosoma* and subsequently reinfection is common in endemic areas. The World Health Organisation  
24 (WHO) reported that 219 million people worldwide needed preventative treatment against  
25 schistosomiasis in 2015. Of those who required treatment, less than one third received it through mass  
26 drug administration (MDA) programmes (1). Even more disconcerting is that modelling studies suggest  
27 that MDA will only reduce the prevalence of schistosomiasis if more than 70% of communities  
28 participate and the MDA is conducted annually (2). A drug-based strategy alone therefore may not move  
29 national schistosomiasis programs of low to middle income countries from morbidity control towards  
30 elimination (3). Other interventions, working alongside MDA, such as vaccination, could effectively

31 prevent reinfection, and thus eliminate schistosomiasis. Vaccination with radiation-attenuated cercariae  
32 protects murine and non-human primate models against challenges with schistosomes. However, using  
33 radiation-attenuated cercariae in human trials is impractical because it is difficult to produce under good  
34 manufacturing practice (GMP), and delivery of the vaccine under liquid nitrogen presents considerable  
35 logistical challenges. As a consequence, recombinant antigens that can be easily produced are being  
36 considered as potential subunit vaccine candidates (Reviewed in (4)). Some of these vaccine candidates  
37 are efficacious against challenge infection in animal models, but show low immunogenicity as purified  
38 single antigens when tested further in human preclinical tests. Therefore, we suggest two approaches to  
39 improve the immunogenicity of *Schistosoma* vaccine antigens. Firstly, multiple, and not single, antigens  
40 should be used (both EV and non-EV encoded) in the development of schistosomiasis vaccine. Secondly,  
41 we consider synthetic vesicles as a proof of concept antigen delivery and adjuvant system. *Schistosoma*-  
42 shed vesicles have been recently identified (5, 6), but whether we can design synthetic stimulatory  
43 versions of these vesicles to deliver *Schistosoma* vaccine targets is a question yet to be addressed. This  
44 forum article examines the potential of using synthetic vesicles as adjuvant and delivery vehicle  
45 containing multiple schistosomula vaccine candidates.

#### 46 **Targeting Schistosomula Antigens as Vaccine Candidates**

47 The schistosomulum is the transition phase between a free-living non-feeding cercaria in fresh water  
48 and the parasitic blood fluke in the mammalian host. When cercariae penetrate human skin, they  
49 transform into the skin-stage schistosomula (Fig. 1). The skin-stage schistosomula up regulate specific  
50 genes during transformation to facilitate invasion and to survive the hostile host immune response (7).  
51 The schistosomula also develop a new double lipid bilayer outer membrane covering the tegument that  
52 facilitates survival within the host. Just as the new coat develops, the early post-penetration  
53 schistosomulum is vulnerable to host immune-mediated attack (8). The late phase schistosomulum is  
54 less susceptible to both eosinophil and macrophage mediated cytotoxicity when it develops towards  
55 adulthood. Early schistosomulum antigens are therefore possible candidates to develop a prophylactic  
56 vaccine against human schistosome infections. However, there are few current efforts to identify and  
57 prioritise schistosomula antigens for a novel vaccine. One such initiative was TheSchistoVac consortium  
58 that targeted antigens highly expressed by the skin schistosomula for vaccine development  
59 (<http://www.theschistovac.eu/>). The work done provides a template for future targeted (stage-specific)  
60 vaccine development.

61 Schistosomes are complex multicellular organisms, and this may partly explain why current vaccines  
62 composed of a single antigen are not capable of inducing long-lived protective immunity. We propose  
63 multiple antigen preparations to target different aspects of the early stage schistosomula ranging from  
64 tegument formation and turnover to metabolite (glucose) uptake. In fact, the multivalent chimeric  
65 schistosomiasis vaccine of SmTSP-2 and Sm29 induces more robust immune responses compared to  
66 single antigen preparations in mice (9). Although identifying new antigens based on the schistosomulum  
67 is a critical step, combining new and existing antigens as a multiple vaccine preparation is, we believe, a  
68 necessary step in designing the next generation of vaccine to a complex, multicellular organism. We  
69 would suggest both non-EV (to target the schistosomula) and EV encoded (to target secreted EVs)  
70 antigens. Finally, the multiple antigen vaccine will require new tools such as synthetic vesicles to be  
71 delivered to immune cells.

## 72 **Synthetic Vesicles to Deliver *Schistosoma* Vaccine Candidates**

73 Schistosomes release excreted/secreted (E/S) products to survive the hostile host immune system.  
74 Among these products characterised to date are *Schistosoma*-shed vesicles known as extracellular  
75 vesicles (EVs) (5, 6), spherical structures encapsulated by a lipid bilayer and shown to be responsible for  
76 intercellular communication (10). The major subsets of EVs are exosomes, microvesicles and apoptotic  
77 bodies. EVs are classified based on their biogenesis, their size, and what surface markers they express.  
78 Of importance is that *Schistosoma* EVs (derived from both schistosomula and adult worms) contain  
79 potential vaccine candidates including SmTSP-2 and Sm29 (5, 6).

80 We suggest packaging schistosome vaccine antigens in synthetic vesicles because naturally occurring *S.*  
81 *mansoni* EVs may contain inhibitory biological material such as miRNAs and tsRNAs (6). Packaging  
82 parasite antigens into vesicles will improve their antigenicity compared to using the antigens directly for  
83 vaccination (11). Another advantage of utilizing synthetic vesicles is that they are free of host proteins  
84 that have been described in EVs of other parasites such as *Echinostoma caproni* and *Fasciola hepatica*  
85 (12). The proof-of-concept for manufacturing synthetic vesicles already exists with other lipid molecules  
86 such as virus like particles (VLPs) and outer membrane vesicles (OMVs). The pros and cons of antigen  
87 delivery using synthetic vesicles are summarised in Table 1. We suggest it is now time to take this  
88 technology forward and target the schistosome.

89 Vaccine candidates within vesicles are also effectively protected from degradation as they move through  
90 body fluids, improving their stability within host. For synthetic vesicles to work as adjuvants, additional  
91 ligands that target receptors on antigen presenting cells such as pathogen recognition receptors on

92 dendritic cells could be added on the vesicle surface using glycosylphosphatidylinositol (GPI) anchors for  
93 robust cellular responses. With appropriate thought given to the incorporation of membrane-embedded  
94 glycoprotein ligands or receptors, targeting specific immunological cells could be engineered and  
95 achieved. In addition to targeting the actual schistosomula, immune responses induced by synthetic  
96 vesicles (to EV encoded antigens) will also target and neutralise *Schistosoma* EVs, decreasing the ability  
97 of the schistosomula to dampen immune responses and make the environment less suitable for survival.  
98 All in all, immune responses to multiple antigen preparations from the early phase of the  
99 schistosomulum packaged in synthetic vesicles may prevent development of the adult schistosomes and  
100 subsequently the laying of eggs that cause pathology associated with schistosomiasis.

### 101 **Conclusion**

102 Although schistosomiasis is treatable, reinfections are common in endemic areas. It is widely  
103 acknowledged that a vaccine used alongside chemotherapy would control and possibly eliminate  
104 schistosomiasis. We have suggested using synthetic vesicles that are preloaded with multiple  
105 schistosomula antigens to elicit protective, skin-stage host responses as a next-generation anti-  
106 schistosomal vaccine. As we move towards 2025, the year WHO set to eliminate schistosomiasis  
107 globally, these and other novel approaches are required to develop vaccines.

### 108 **Conflict of Interests**

109 The authors declare that there is no conflict of interest.

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121 **References**

- 122 1. Schistosomiasis and soil-transmitted helminthiasis: number of people treated in 2015. *Wkly*  
123 *Epidemiol Rec.* 2016;91(49-50):585-95.
- 124 2. Gurarie D, Yoon N, Li E, Ndeffo-Mbah M, Durham D, Phillips AE, et al. Modelling control of  
125 *Schistosoma haematobium* infection: predictions of the long-term impact of mass drug administration in  
126 Africa. *Parasit Vectors.* 2015;8:529.
- 127 3. Ross AG, Olveda RM, Chy D, Olveda DU, Li Y, Harn DA, et al. Can mass drug administration lead  
128 to the sustainable control of schistosomiasis? *J Infect Dis.* 2015;211(2):283-9.
- 129 4. Hewitson JP, Maizels RM. Vaccination against helminth parasite infections. *Expert Rev Vaccines.*  
130 2014;13(4):473-87.
- 131 5. Sotillo J, Pearson M, Potriquet J, Becker L, Pickering D, Mulvenna J, et al. Extracellular vesicles  
132 secreted by *Schistosoma mansoni* contain protein vaccine candidates. *Int J Parasitol.* 2016; 46(1):1-5.  
133 Nowacki FC, Swain MT, Klychnikov OI, Niazi U, Ivens A, Quintana JF, et al. Protein and small non-  
134 coding RNA-enriched extracellular vesicles are released by the pathogenic blood fluke *Schistosoma*  
135 *mansoni*. *J Extracell Vesicles.* 2015;4:28665.
- 136 7. Fitzpatrick JM, Peak E, Perally S, Chalmers IW, Barrett J, Yoshino TP, et al. Anti-schistosomal  
137 intervention targets identified by lifecycle transcriptomic analyses. *PLoS Negl Trop Dis.* 2009;3(11):e543.
- 138 8. Butterworth AE, Sturrock RF, Houba V, Rees PH. Antibody-dependent cell-mediated damage to  
139 schistosomula in vitro. *Nature.* 1974;252(5483):503-5.
- 140 9. Pinheiro CS, Ribeiro AP, Cardoso FC, Martins VP, Figueiredo BC, Assis NR, et al. A multivalent  
141 chimeric vaccine composed of *Schistosoma mansoni* SmTSP-2 and Sm29 was able to induce protection  
142 against infection in mice. *Parasite Immunol.* 2014;36(7):303-12.
- 143 10. Coakley G, Buck AH, Maizels RM. Host parasite communications—Messages from helminths for  
144 the immune system: Parasite communication and cell-cell interactions. *Molecular and biochemical*  
145 *parasitology.* 2016;208(1):33-40.
- 146 11. Beauvillain C, Juste MO, Dion S, Pierre J, Dimier-Poisson I. Exosomes are an effective vaccine  
147 against congenital toxoplasmosis in mice. *Vaccine.* 2009;27(11):1750-7.
- 148 12. Marcilla A, Trelis M, Cortes A, Sotillo J, Cantalapiedra F, Minguez MT, et al. Extracellular vesicles  
149 from parasitic helminths contain specific excretory/secretory proteins and are internalized in intestinal  
150 host cells. *PLoS One.* 2012;7(9):e45974.

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153 **Figure 1. Migration of schistosomula through the host skin.** 1. Cercaria is attracted to human skin. 2  
154 Cercaria burrows through the skin and detaches the bifurcated tail to form a schistosomulum. 3  
155 Schistosomulum release excretion/secretion (ES) products including extracellular vesicles (EVs) that  
156 interact with resident Langerhans cells, which migrate to skin draining lymph nodes to initiate adaptive  
157 immune responses. 4 The schistosomulum moves towards the basement membrane that temporarily  
158 halts their migration. 5 Once in the dermis, the schistosomulum is vulnerable to antibody-mediated  
159 killing by granulocytes. 6 Schistosome-induced cytokines activate more phagocytes and polarize the  
160 immune response towards inflammatory responses. 7 Schistosomulum penetrates dermal veins and

161 migrates to the lungs. 8 Schistosomulum is coated with host proteins as an immune evasion mechanism.

162 The figures were adapted and modified from Servier Medical Art (<http://smart.servier.com/>).

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166 Table 1. Pros and cons of antigen delivery via synthetic vesicles

|               |  |
|---------------|--|
| Advantages    | EV and non-EV antigens target both schistosomula and EVs increasing immunogenicity   |
|               | Antigen presenting cells can be targeted by displaying specific ligands on outer surface of synthetic vesicles using GPI anchors                           |
|               | Adding molecules that activate antigen presenting cells means that synthetic vesicles are not just antigen delivery vehicles, but also an adjuvant as well |
|               | Synthetic vesicles exhibit natural EV properties such as stability and resistance to enzymatic degradation in body fluids                                  |
|               | Naturally occurring EV cargo, such as miRNA, with inhibitory properties are avoided in synthetic vesicles  |
|               | Synthetic vesicles lack host proteins, a potential mechanism for decreasing immunogenicity   |
|               |  |
| Disadvantages | Expensive to manufacture   |
|               | Risk of reactogenicity associated with synthetic materials may lead to adverse effects in humans   |
|               | Extensive regulatory requirements are expected for human use license   |

167

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Figure 1

