# **Trends in Microbiology** | Microbe of the Month

# Shigella sonnei

Vincenzo Torraca , \*\* Kathryn Holt ( , 1,2,\* and Serge Mostowy ( ) 1,\*

<sup>1</sup>Department of Infection Biology, Faculty of Infectious and Tropical Diseases, London School of Hygiene and Tropical Medicine, London, UK

<sup>&</sup>lt;sup>2</sup>Department of Infectious Diseases, Central Clinical School, Monash University, Melbourne, Australia

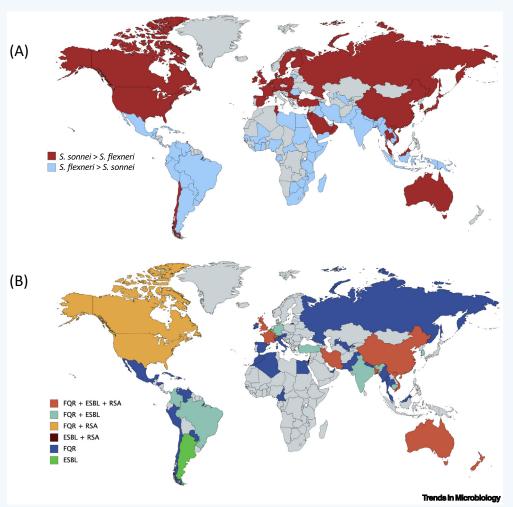


Figure 1. Epidemiology of *Shigella sonnei*. (A) Cases of *S. sonnei* versus *Shigella flexneri*. Cases of *S. sonnei* are increasing globally. Red indicates countries where *S. sonnei* is the dominant cause of shigellosis (when compared with *S. flexneri*). Blue indicates countries where a higher proportion of *S. flexneri* is still being reported (although *S. sonnei* cases may be rapidly increasing). Data presented according to Thompson *et al.* (see Literature). (B) Distribution of *S. sonnei* resistance to first- and second-line antibiotics. FQR, fluoroquinolone resistance; ESBL, extended-spectrum β-lactamase-producing (i.e., resistant to third-generation cephalosporins or carbapenems); RSA, reduced sensitivity to azithromycin. Countries are colored according to antibiotic-resistant cases reported in Table S1 in the supplemental information online.

Shigella sonnei is a rod-shaped, Gram-negative facultative intracellular pathogen. It was named 'Sonne's bacillus' after Carl Olaf Sonne who described it as a causative agent of bacillary dysentery. *S. sonnei* is distributed worldwide and represents the most common cause of shigellosis in industrialized regions in Europe, North America, and Australia. It is currently undergoing expansion in middle-income countries across Asia, Latin America, and the Middle East. *S. sonnei* evolved from *Escherichia coli* to specialize in intracellular infection of the human gut epithelium, and its genome comprises a 4.99 Mbp circular chromosome and a 216 kbp invasion plasmid (pINV) required for virulence. The chromosome is ~6% smaller than other *E. coli* and is punctuated by >300 copies of insertion sequence (IS) elements, whose expansion has degraded the genome through disruption and deletion of genes. Here we describe the key and disease facts allowing bacteria to evade host immune defences and to establish infection.

#### **KEY FACTS:**

A chromosomally encoded type VI secretion system (T6SS), involved in bacterial competition, is a key determinant for niche occupancy.

pINV-encoded virulence determinants include a type III secretion system (T3SS) involved in host cell invasion and phagosomal escape, the actin-polymerizing factor IcsA, a g4c capsule, and an unusual O-antigen encoded by genes horizontally acquired from *Plesiomonas shigelloides*.

The g4c capsule and O-antigen of S. sonnei protect the bacteria from a wide variety of host defence mechanisms, including phagocytosis, phagolysosome degradation, and complement-mediated lysis.

### **DISEASE FACTS:**

S. sonnei causes acute, self-limiting disease characterized by bloody diarrhea, fever, and abdominal pain. Infections can be life-threatening for children under 5 years and can stunt growth.

S. sonnei spreads via the fecal-oral route, as ingested bacteria can survive the gastric acidity and be released in stool.

Outbreaks are often centered around large events or care facilities. *S. sonnei* is also sexually transmitted, particularly among men who have sex with men.

The main treatment for bacillary dysentery is rehydration therapy. Fluoroquinolones, cephalosporins, and azithromycin are recommended when antimicrobials are required. However, resistance is increasingly common and new antimicrobials are needed.

There is no licensed vaccine. O-antigen-based immunization has been proposed; it holds particular promise as *S. sonnei* has only one serotype.

### \*Correspondence:

vincenzo.torraca@lshtm.ac.uk (V. Torraca), kat.holt@lshtm.ac.uk (K. Holt), and serge.mostowy@lshtm.ac.uk (S. Mostowy).



# **Trends in Microbiology** | Microbe of the Month

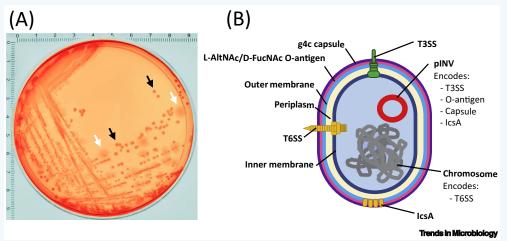


Figure 2. Shigella sonnei Virulence Determinants. (A) S. sonnei 53G plated on Congo Red agar. The virulence plasmid (pINV) of S. sonnei is essential for pathogenesis in vivo but is unstable outside the host (and is lost when bacteria are cultured in vitro). As a result of maintenance or loss of pINV, S. sonnei presents two different phenotypes on Congo Red plates. Black arrows indicate small smooth red colonies (Phase I S. sonnei). These colonies retain pINV and accumulate Congo Red dye because they express a type III secretion system (T3SS). The small smooth phenotype is due to expression of O-antigen (O-Ag). White arrows indicate large rough white colonies (Phase II S. sonnei). These colonies are unable to accumulate Congo Red and have an irregular shape because they lose pINV (which encodes both T3SS and O-Ag). In this case, bacteria were plated from a glycerol stock (originally obtained from a liquid culture of a single Phase I colony) and incubated overnight at 37°C. Grid numbers indicate the size in centimetres. (B) Schematic representing key S. sonnei virulence determinants. S. sonnei encodes a T3SS crucial for host cell invasion; lcsA which mediates actin-based motility; a T6SS crucial for bacterial competition and niche occupancy; a horizontally acquired g4c capsule and O-antigen involved in resistance to phagocytosis, complement-mediated lysis, and phagolysosomal degradation. L-AltNAc, 2-acetamido-2-deoxy-L-altruronic acid; D-FucNAc, N-acetyl-2-acetamido-4-amino-2,4-dideoxy-D-fucose. Figure adapted from Torraca et al. (see Literature).

## **Acknowledgments**

Work in the S.M. laboratory is supported by a European Research Council Consolidator Grant (772853 – ENTRAPMENT), Wellcome Trust Senior Research Fellowship (206444/Z/17/Z), and the Lister Institute of Preventive Medicine. K.E.H. is supported by a Senior Medical Research Fellowship from the Viertel Foundation of Australia.

### **Supplemental Information**

 $Supplemental\ information\ associated\ with\ this\ article\ can\ be\ found\ online\ at\ https://doi.org/10.1016/j.tim.2020.02.011.$ 

### Literature

- Anderson, M.C. et al. (2017) Shigella sonnei encodes a functional T6SS used for interbacterial competition and niche occupancy. Cell Host Microbe 21, 769–776.e3
- 2. Baker, K.S. *et al.* (2015) Intercontinental dissemination of azithromycin-resistant shigellosis through sexual transmission: A cross-sectional study. *Lancet Infect. Dis.* 15, 913–921
- Baker, K.S. et al. (2018) Horizontal antimicrobial resistance transfer drives epidemics of multiple Shigella species. Nat. Commun. 9, 1462
- 4. Caboni, M. et al. (2015) An O-antigen capsule modulates bacterial pathogenesis in Shigella sonnei. PLoS Pathog. 11,
- 5. Chung The, H. et al. (2019) Dissecting the molecular evolution of fluoroquinolone-resistant Shigella sonnei. Nat Commun
- 6. Hawkey, J. et al. (2019) Impact of insertion sequences on convergent evolution of Shigella species. bioRxiv, 680777
- Holt, K.E. et al. (2012) Shigella sonnei genome sequencing and phylogenetic analysis indicate recent global dissemination from Europe. Nat. Genet. 44, 1056–1059
- 8. Kotloff, K.L. et al. (2018) Shigellosis. Lancet 391, 801–812
- Thompson, C.N. et al. (2015) The rising dominance of Shigella sonnei: An intercontinental shift in the etiology of bacillary dysentery. PLoS Negl. Trop. Dis. 9, e0003708
- 10. Torraca, V. et al. (2019) Shigella sonnei infection of zebrafish reveals that O-antigen mediates neutrophil tolerance and dysentery incidence. PLoS Pathog. 15, e1008006

### TAXONOMY AND CLASSIFICATION:

KINGDOM: Bacteria
PHYLUM: Proteobacteria
CLASS: Gammaproteobacteria
ORDER: Enterobacteriales
FAMILY: Enterobacteriaceae
GENUS: Shigella (whole-genome
comparative analysis does not support
distinct genus designation and suggests
placement of Shigella spp. as lineages

within the species Escherichia coli)

SPECIES: Shigella sonnei

SUBSPECIES: S. sonnei is monoclonal. All circulating strains originated from a common ancestor in Europe ~1500 AD. There is only one serogroup (Shigella serogroup D) consisting of one serotype. Five distinct subtypes (lineages I–V) have been identified by whole-genome sequencing; the common laboratory strain 53G belongs to lineage II; however, most clinical isolates now belong to lineage III.

