Interventions to improve oral vaccine performance: a systematic review and meta-analysis

James A Church*, Edward P Parker*, Beth D Kirkpatrick, Nicholas C Grassly, Andrew J Prendergast

Summary

Background Oral vaccines underperform in low-income and middle-income countries compared with in high-income countries. Whether interventions can improve oral vaccine performance is uncertain.

Methods We did a systematic review and meta-analysis of interventions designed to increase oral vaccine efficacy or immunogenicity. We searched Ovid-MEDLINE and Embase for trials published until Oct 23, 2017. Inclusion criteria for meta-analysis were two or more studies per intervention category and available seroconversion data. We did random-effects meta-analyses to produce summary relative risk (RR) estimates. This study is registered with PROSPERO (CRD42017060608).

Findings Of 2843 studies identified, 87 were eligible for qualitative synthesis and 66 for meta-analysis. 22 different interventions were assessed for oral poliovirus vaccine (OPV), oral rotavirus vaccine (RVV), oral cholera vaccine (OCV), and oral typhoid vaccines. There was generally high heterogeneity. Seroconversion to RVV was significantly increased by delaying the first RVV dose by 4 weeks (RR 1·37, 95% CI 1·16–1·62) and OPV seroconversion was increased with monovalent or bivalent OPV compared with trivalent OPV (RR 1·51, 95% CI 1·20–1·91). There was some evidence that separating RVV and OPV increased RVV seroconversion (RR 1·21, 95% CI 1·00–1·47) and that higher vaccine inoculum improved OCV seroconversion (RR 1·12, 95% CI 1·00–1·26). There was no evidence of effect for anthelmintics, antibiotics, probiotics, zinc, vitamin A, withholding breastfeeding, extra doses, or vaccine buffering.

Interpretation Most strategies did not improve oral vaccine performance. Delaying RVV and reducing OPV valence should be considered within immunisation programmes to reduce global enteric disease. New strategies to address the gap in oral vaccine efficacy are urgently required.

Funding Wellcome Trust, Bill & Melinda Gates Foundation, UK Medical Research Council, and WHO Polio Research Committee.

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Introduction

Despite global declines in enteric disease, approximately 650000 children die from diarrhoea annually,1 with rotavirus causing a third of deaths.² A major obstacle to sustained progress is the reduced efficacy of oral vaccines in low-income and middle-income countries (LMICs).3 This phenomenon was first observed for oral poliovirus vaccine (OPV) in the 1950s and has since been described for several oral vaccines in many countries. For example, rotavirus vaccine (RVV) efficacy against severe rotavirus gastroenteritis is only 39% in sub-Saharan Africa4 and 48% in south Asia,5 compared with 85-98% in Europe and the USA.67 The reasons for oral vaccine underperformance are unclear.8 Potential explanations include concurrent enteric infections,^{9,10} microbiota composition,¹¹ environmental enteric dysfunction,12 interference from maternal antibodies,13,14 histoblood group antigens,15 and micronutrient deficiencies.^{16,17} The relative contributions of these factors can vary depending on oral vaccine target, recipient age, and setting.

Multiple studies have assessed interventions to improve oral vaccine performance, broadly categorised as adjunctive interventions given before or with vaccination (such as micronutrient supplements or antimicrobials) or adjustments to the vaccine formulation and delivery schedule (such as increased vaccine inoculum or altered timing). Reviews have assessed specific interventions¹⁸ or individual vaccines,^{19,20} but none have assessed the full range of approaches used across all oral vaccines. We therefore did a systematic review and meta-analysis of interventions to increase oral vaccine efficacy or immunogenicity in LMICs.

Methods

Search strategy and selection criteria

We followed PRISMA guidelines throughout our review. We searched Ovid-MEDLINE and Embase for English language articles published any time up to Oct 23, 2017, describing studies assessing interventions to improve oral vaccine performance (appendix p 2). We examined the reference lists of articles to identify additional studies, searched the grey literature, and contacted experts for unpublished data.

We screened full-text papers for inclusion in the qualitative synthesis. We included randomised trials, cluster-randomised trials, non-randomised trials, and



Lancet Infect Dis 2019;

19: 203–14 See **Comment** page 122

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See Online for appendix

Research in context

Evidence before this study

Oral vaccines consistently underperform when given to children in low-income and middle-income countries (LMICs) but the underlying causes and potential intervention approaches are unclear. We searched Ovid-MEDLINE and Embase for systematic reviews and meta-analyses of interventions to improve oral vaccine performance published up to May 31, 2018, using the same search strategy detailed in the appendix (p 5). We identified three systematic reviews, one including a meta-analysis. The meta-analysis, published in 1998, found that regimens containing a single dose of typhoid vaccine were less effective than were regimens with two or more doses. Of the remaining two systematic reviews, both published in 2017, one examined differences in rotavirus vaccine scheduling across eight trials, concluding that seroconversion was lower among children given the vaccine earlier in infancy (age 6 and 10 weeks) versus later in infancy (age 10 and 14 weeks). The other, restricted to probiotics, found a beneficial effect of probiotics on vaccine responses (parenteral and oral vaccines) in half of the studies. However, there have been no systematic reviews assessing all intervention strategies (both adjunctive and vaccine design or delivery adjustments) across oral poliovirus, rotavirus, cholera, and typhoid vaccines. We assessed the full range of intervention approaches and oral vaccines investigated to date, to identify whether any strategies could be adopted by immunisation programmes, and to identify research gaps to inform future trials.

meta-analyses assessing one or more interventions. We did not include case-control studies, controlled beforeafter studies, or observational data from cross-sectional studies and case series. Studies were excluded if they included fewer than ten participants, did not include a control group, or did not measure vaccine efficacy or immunogenicity. We excluded strategies bypassing the oral route (eg, use of inactivated poliovirus vaccine), and pre-licensure dose-finding trials. We did not exclude studies on the basis of participant age or setting, because studies from high-income countries or in older age groups might provide insights into oral vaccine failure among infants in LMICs. For meta-analysis, inclusion criteria required two or more studies per intervention category and available seroconversion data. The protocol is available at www.crd.york.ac.uk/PROSPERO, CRD42017060608.

For the **protocol** see www.crd.york.ac.uk/PROSPERO

Outcome definitions

The prespecified primary outcome was oral vaccine performance, defined as either vaccine efficacy or immunogenicity, depending on study design. Vaccine efficacy was defined as percentage disease reduction in the vaccinated group compared with the unvaccinated group. Our prespecified preferred measure of vaccine immunogenicity was the proportion of children with seroconversion, as defined by each study, after the last

Added value of this study

This is the first systematic review and meta-analysis of approaches to improve oral vaccine performance among children. We assessed the evidence for 22 interventions targeting four oral vaccines. Overall, we found few interventions had a substantial benefit on the basis of the available evidence, highlighting the challenge in overcoming oral vaccine underperformance. However, we found that delaying the first dose of rotavirus vaccine and reducing oral poliovirus vaccine valence can improve oral vaccine immunogenicity.

Implications of all the available evidence

Existing oral vaccines and their schedules are poorly effective among children in LMICs. Most adjunctive interventions to date have not improved oral vaccine performance. Untested interventions such as water, sanitation and hygiene, the effect of booster doses given later in infancy, and increasing vaccine inoculum for rotavirus vaccine warrant further study. Costbenefit and modelling analyses that consider the full effect of delaying the first dose of rotavirus vaccine should be undertaken. However, the global research community should also strongly consider new and innovative ways to address this efficacy gap, including a decreased reliance on oral vaccines, to reduce the global burden of enteric disease.

scheduled vaccine dose. Alternative measures included geometric mean titres and fold-rise if seroconversion was not reported (appendix p 8). Studies without seroconversion data were included in the qualitative synthesis but were not eligible for meta-analysis. Timing of vaccine immunogenicity measurement was not considered in the eligibility criteria. The chosen correlates of protection were serum neutralising antibodies for OPV, serum IgA for RVV, and vibriocidal antibodies for oral cholera vaccine (OCV).

Data analysis

Two independent reviewers (JAC and EPP) assessed eligibility of each full-text article; a third (AJP) arbitrated for cases without consensus. One reviewer (JAC) extracted data, and a second (EPP) validated data extraction for 10% of studies. Descriptive and quantitative data were entered into a spreadsheet based on the Cochrane data extraction tool.²¹ If more than one intervention or oral vaccine were reported in the same study, data were extracted separately for each. For studies with a factorial design, data were only extracted from the combined group if there was no interaction between interventions, otherwise data from individual groups were used. Where numerical data were not reported, we requested data from authors or used GetData Graph Digitiser to extract results from figures. We assessed

2843 articles yielded from initial search 1445 MEDLINE (last updated Oct 19, 2017) 1398 Embase (last updated Oct 23, 2017) 1196 articles excluded after removal of duplicates 1647 records screened 1384 articles excluded after scanning titles 263 abstracts screened 6 articles included after search of grey literature and correspondence with experts in field 165 articles excluded after scanning abstracts 7 articles included after scanning reference lists from selected articles 111 articles included for full-text screening 24 articles excluded after full-text screening 5 duplicate material 6 no intervention 6 no appropriate control group 4 no measure of vaccine response 2 unable to retrieve data 1 ineligible study type 87 articles included in qualitative synthesis* 15 micronutrients 1 antibiotic 6 probiotic 3 anthelmintic 6 withholding breastfeeding 44 dosing 10 OPV valence 11 buffer 4 miscellaneous 21 articles did not report seroconversion as the primary outcome 66 articles included in meta-analysis

Figure 1: Study selection

OPV=oral polio vaccine. *Studies exploring multiple interventions are duplicated within this breakdown.

Of the 87 studies, 66 (from 13 intervention categories) were eligible for inclusion in the meta-analysis (table). Eight studies (9.3%) included in the qualitative synthesis reported vaccine efficacy as the primary outcome. Most studies (n=66, 75.9%) reported vaccine seroconversion as the primary outcome. Additional immunogenicity characteristics are described in the appendix (p 13).

GRADE scoring is reported in the full dataset. For the full dataset see 11 studies (12.6%) had a low risk of bias across all https://osf.io/bemw6/

quality of evidence for each study using Grading of Recommendations, Assessment, Development and Evaluation (GRADE) criteria²² (appendix p 9).

To assess the effect of each intervention on seroconversion, we did random-effects meta-analyses in the R package metafor.²³ We calculated summary relative risk (RR) estimates and 95% CIs for each vaccine evaluated in two or more studies. If more than one vaccine was assessed for a given intervention, we did a pooled analysis combining across vaccines, and a mixed-effects analysis with vaccine type as a moderator. For mixed-effects models, we identified whether heterogeneity associated with vaccine type or residual heterogeneity was significant via χ^2 tests.²⁴ If residual heterogeneity was significant (p<0.05), we assessed age, setting, and background immunogenicity (seroconversion rate in the control group) as secondary moderators. We used funnel plots to check for publication bias and tested for asymmetry using Egger's test. Further details of the analysis are provided in the appendix (p 10). All analyses were done in R (version 3.4.1).

Role of the funding source

The funder had no role in study design, data collection, data analysis, data interpretation, or writing of the report. The corresponding author had full access to all data in the study and had final responsibility for the decision to submit for publication.

Results

Of 2843 articles, 87 were eligible for qualitative synthesis (figure 1). The studies are summarised in the table and the full dataset is available online. 74 (85.1%) were randomised trials and 13 (14.9%) non-randomised trials. The studies were done in 38 countries between 1972 and 2017 (appendix p 11) and enrolled between 30 and 225 998 participants (median 301, IQR 128-718). 11 studies $(12 \cdot 6\%)$ were done in adults.

22 interventions were assessed (14 adjunctive interventions and eight vaccine design or delivery interventions), grouped into 17 categories: anthelmintic therapy (n=3), antibiotic therapy (n=1), probiotic supplementation (n=6), zinc supplementation (n=6), vitamin A supplementation (n=6), other micronutrient supplementation (n=3), withholding breastfeeding (n=6), extra dose or doses (n=10), extra dose given at birth (n=6), early first dose (n=1), delayed first dose (n=4), shortened interval between doses (n=5), RVV given with versus without OPV (n=7), increased vaccine inoculum (n=11), different OPV valences (n=10), inclusion of buffer (n=11), and miscellaneous (n=4, appendix p 4). No studies of water, sanitation, or hygiene interventions were found. 12 studies included more than one intervention, of which two employed a factorial design (appendix p 12). OPV was the most common vaccine studied (46 studies, 52.9%), followed by RVV (n=24, 37.6%), OCV (n=15, $17 \cdot 2\%$), and oral typhoid vaccine (n=9, $10 \cdot 3\%$).

	Oral poliovirus vaccine	Oral rotavirus vaccine	Oral cholera vaccine	Oral typhoid vaccine
Total studies (n)*	46	24	15	9
Intervention				
Anthelmintic	0	0	2	1
Antibiotic†	1	0	0	0
Breastfeeding withheld	2	3	1	0
Buffer	1	4	3	3
Delayed first dose	0	4	0	0
Early first dose†	1	0	0	0
Extra dose or doses	1	6	1	2
Extra dose at birth	5	1	0	0
Miscellaneous†	3	0	1	0
Narrow dose interval	3	1	0	1
OPV valence	10	NA	NA	NA
Other micronutrients†	2	0	0	1
Probiotic	1	2	2	1
RVV separated from OPV	7	7	NA	NA
Vaccine inoculum	7	0	4	0
Vitamin A	4	0	1	1
Zinc	1	1	4	0
Age group				
<1 month	19	1	0	0
1–12 months	24	23	3	0
1–15 years	2	0	4	6‡
≥16 years	1	0	8	3
Mean age (SD; months)	4.2 (7.9)	1.9 (1.3)	141.6 (163.3)	187.9 (133.8)
Sex				
Men (%)	51.3	45.7	50.5	55.9
Women (%)	48.7	54.3	49.5	44·1
Location				
Africa	8	6	2	1
Asia	25	10	7	2
Europe	5	3	2	2
Americas	8	4	4	4
Oceania	0	1	0	0
Study size				
<50	7	1	2	2
50-500	37	21	13	2
>500	2	2	0	5
Total seroconversion data (n)	8838	8951	1305	252.020

RVV=oral rotavirus vaccine. OPV=oral poliovirus vaccine. *Of 87 unique studies, some studies examined two or more interventions and some reported on multiple oral vaccine targets (appendix p 12).+There were insufficient studies (fewer than two) of antibiotics, early first dose, other micronutrients, and miscellaneous interventions (maternal vitamin A, horse anti-serum, soya formula, and *Escherichia coli* K-12) for inclusion in the meta-analysis. ‡Most typhoid studies recruited children aged between 5 and 22 years.

Table: Overview of 87 intervention studies included in the systematic review

domains of study quality, 76 (87.4%) had an unclear risk of bias in at least one domain, and 42 (48.3%) had a high risk of bias in two or more domains (appendix p 14). 30 studies were downgraded during GRADE assessment for indirectness, mostly because of poor generalisability (ie, not studying children or not done in an LMIC). Potential publication bias, assessed using funnel plots, was identified in seven intervention categories (appendix p 26).

Six randomised controlled trials reported oral vaccine seroconversion following vitamin A supplementation. Among five studies eligible for meta-analysis (four on OPV, one on OCV), there was no significant effect of vitamin A supplementation on seroconversion (overall RR 1.01, 95% CI 0.99-1.03). There were six studies of zinc supplementation, of which five were included in the meta-analysis (three on OCV, one on RVV, and one on OPV). Overall, there was no significant effect of zinc supplementation on seroconversion (1.11, 95% CI 0.87-1.42; figure 2). A factorial trial in Bangladesh reported an interaction between vitamin A and zinc supplementation, whereby zinc combined with vitamin A increased seroconversion to OCV more than zinc alone.25 Single studies reported several other micronutrient interventions. There was no evidence that vitamin A given to mothers post partum,26 selenium capsules,27 or oral iodine²⁸ had an effect on OPV immunogenicity, and giving multiple micronutrients had no effect on response to oral typhoid vaccine.29

Six studies assessed withholding breastfeeding for 1–10 h (mean 3.5, SD 3.4) around the time of oral vaccination; five were eligible for meta-analysis (one on OCV, three on RVV, one on OPV). There was no evidence of benefit for seroconversion to OCV, RVV, or OPV (overall RR 0.93, 95% CI 0.75–1.14; appendix p 15).

Six studies reported probiotic interventions, of which four were eligible for meta-analysis (one on OCV, two on RVV, one on OPV). In each study, daily probiotics were started 7 days before vaccination and continued for at least 3 weeks after; three studies used lactobacillus³⁰⁻³²and one used bifidobacterium.³³ There was no evidence of an effect on seroconversion to OCV, RVV, or OPV (overall RR 1.09, 95% CI 0.84–1.41; figure 2).

32 studies reported changes in vaccine formulation (addition of a buffer, increased inoculum, or altered OPV valence); 28 were eligible for meta-analysis (eight on buffer, 11 on inoculum, nine on OPV valence). There was weak evidence that adding buffer increased seroconversion to OCV (RR 1.32, 95% CI 0.98-1.78), although there were few participants (n=219). Two typhoid studies showed increased vaccine efficacy when reconstituted in buffer compared with a capsule control.^{34,35} Overall, when combining data for all vaccines, there was weak evidence of improved seroconversion with buffer (1.03, 95% CI 0.98-1.09; appendix p 16). Increases in vaccine inoculum showed some evidence of benefit for seroconversion across four studies for OCV ($1 \cdot 12$, $1 \cdot 00 - 1 \cdot 26$), but not for OPV (1.02, 0.95–1.09; figure 2). Nine studies exploring adjustments to OPV valence were included in the metaanalysis; seven used monovalent OPV and two used bivalent OPV. Compared with trivalent OPV, monovalent and bivalent OPV significantly increased seroconversion (1.51, 95% CI 1.20-1.91), with a consistent effect across OPV1 and OPV3 (appendix p 19).

	Country	Age (months)	Vaccine	Intervention	n/N (%)	Control	n/N (%)	RR (95% CI)			
	country	Age (months)	vaccine	Intervention	11/19 (26)	Control	11/19 (76)	KK (95% CI)			
Cholera											
Ahmed (January, 2009)	Bangladesh	7.5	2×Wc-rBS	Zinc (20 mg)	11/36 (31%)	No zinc	27/47 (57%)	0.53 (0.31–0.92)			
Ahmed (January, 2009)	Bangladesh	14	2×Wc-rBS	Zinc (20 mg)	27/34 (79%)	No zinc	27/49 (55%)	1.44 (1.06–1.96)			
Ahmed (October, 2009)	Bangladesh	14	2×Wc-rBS	Zinc (20 mg)	15/18 (83%)	No zinc	11/20 (55%)	1.52 (0.97–2.37)			<u> </u>
Albert (2003)	Bangladesh	40	2× Cholerix	Zinc (20 mg)	39/63 (62%)	Placebo	29/63 (46%)	1.34 (0.97–1.87)			_
Summary								1.16 (0.75–1.79)		\Diamond	>
Rotavirus											
azarus (2017)	India	1.2	2×RV1	Zinc (5 mg)	40/143 (28%)	Placebo	37/135 (46%)	1.02 (0.70–1.49)		-+	
OPV3											
Habib (2015)	Pakistan	0.3	4×t0PV	Zinc (10 mg)	94/156 (60%)	Placebo	99/163 (61%)	0.99 (0.83–1.18)		-+-	
Overall								1.11 (0.87–1.42)		-	
leterogeneity among va	accines (Qm 0·1	5, df 2), p=0·927; r	esidual heterogenei	ty (Qe 10∙97, df 3), p=0∙	012				0.33	1.0	3
robiotic											
Cholera											
Matsuda (2011)	Bangladesh	44·1	2×Wc-rBS	Bbb01 (10 ⁹)	40/64 (63%)	Placebo	47/62 (76%)	0.82 (0.65–1.04)			
Rotavirus											
solauri (1995)	Finland	4.1	1× DxRRV	Lc ATCC	26/28 (93%)	Placebo	20/27 (74%)	1.25 (0.98–1.60)			-
azarus (2017)	India	1.2	1× RV1	LGG	42/136 (31%)	Placebo	37/135 (27%)	1.13 (0.78–1.64)		- -	-
Summary								1·21 (0·99–1·49)		\diamond	
OPV3											
DeVrese (2005)*	Germany	Adult	1× 0PV3	LGG	12/21 (57%)	Placebo	8/22 (36%)	1·57 (0·81–3·06) 1·09 (0·84–1·41)			•••
Overall											
Heterogeneity among va	accines (Qm 7·3	5, df 2), p=0∙025; re	esidual heterogenei	ty (Qe 0·22, d† 1), p=0·6	39				0.33	1.0	3
Heterogeneity among va ncreased vaccine inocul Cholera	uccines (Qm 7·3	5, df 2), p=0·025; rd	esidual heterogenei	ty (Qe 0·22, dt 1), p=0·6					0.33	1.0	3
Heterogeneity among va Increased vaccine inocul Cholera Gotuzzo (1993)	um Peru	5, df 2), p=0·025; rd Adult	esidual heterogenei 1× CVD103-	ty (Qe 0·22, dt 1), p=0·6	28/39 (72%)	Std (10 ⁸)	19/39 (49%)	1.47 (1.01-2.15)	0.33	1.0	3
Heterogeneity among va ncreased vaccine inocul Cholera Gotuzzo (1993) Sow (2017)	um Peru Mali	5, df 2), p=0·025; ro Adult Adult	esidual heterogenei 1× CVD103- 1× CVD103-	ty (Qe 0-22, df 1), p=0-6 HgR High (10 ⁹) HgR High (2x10 ⁹)	28/39 28/39 (72%) 40/48 (83%)	Std (10 ⁸) Std (2x10 ⁸)	19/39 (49%) 33/46 (72%)	1·47 (1·01-2·15) 1·16 (0·93-1·45)	0.33	1.0	3
Heterogeneity among va ncreased vaccine inocul Cholera Gotuzzo (1993) Sow (2017) Su-Arehawaratana (1992	uccines (Qm 7·3 um Peru Mali) Thailand	5, df 2), p=0-025; rr Adult Adult Adult Adult	esidual heterogenei 1× CVD103- 1× CVD103- 1× CVD103- 1× CVD103-	ty (Qe 0-22, df 1), p=0-6 HgR High (10 ⁹) HgR High (2x10 ⁹) HgR High (5x10 ⁹)	28/39 (72%) 40/48 (83%) 17/40 (43%)	Std (10 ⁸) Std (2x10 ⁸) Std (5x10 ⁸)	19/39 (49%) 33/46 (72%) 13/39 (33%)	1·47 (1·01-2·15) 1·16 (0·93-1·45) 1·28 (0·72-2·26)	0.33	1.0	3
Heterogeneity among va heterogeneity among va hetero	um Peru Mali) Thailand Indonesia	5, df 2), p=0-025; rr Adult Adult Adult Adult 60–108	esidual heterogenei 1× CVD103- 1× CVD103- 1× CVD103- 1× CVD103- 1× CVD103-	ty (Qe 0-22, df 1), p=0-6 HgR High (10 ⁹) HgR High (2x10 ⁹) HgR High (5x10 ⁹) HgR High (10 ¹⁰)	28/39 (72%) 40/48 (83%) 17/40 (43%) 30/33 (91%)	Std (10 ⁸) Std (2x10 ⁸) Std (5x10 ⁸) Std (5x10 ⁹)	19/39 (49%) 33/46 (72%) 13/39 (33%) 27/31 (87%)	1-47 (1-01-2-15) 1-16 (0-93-1-45) 1-28 (0-72-2-26) 1-04 (0-88-1-24)	0.33		-
Heterogeneity among va heterogeneity among va hetero	um Peru Mali) Thailand Indonesia Indonesia	5, df 2), p=0-025; rr Adult Adult Adult 60–108 60–108	1× CVD103- 1× CVD103- 1× CVD103- 1× CVD103- 1× CVD103- 1× CVD103- 1× CVD103-	ty (Qe 0-22, df 1), p=0-6 HgR High (10 ⁹) HgR High (2x10 ⁹) HgR High (5x10 ⁹) HgR High (10 ¹⁰) HgR High (10 ¹⁰)	28/39 (72%) 40/48 (83%) 17/40 (43%) 30/33 (91%) 26/32 (81%)	Std (10 ⁸) Std (2x10 ⁸) Std (5x10 ⁸) Std (5x10 ⁹) Std (5x10 ⁹)	19/39 (49%) 33/46 (72%) 13/39 (33%) 27/31 (87%) 21/28 (75%)	1-47 (1-01-2-15) 1-16 (0-93-1-45) 1-28 (0-72-2-26) 1-04 (0-88-1-24) 1-08 (0-83-1-42)	0.33		-
Heterogeneity among va heterogeneity among va hetero	um Peru Mali) Thailand Indonesia	5, df 2), p=0-025; rr Adult Adult Adult Adult 60–108 60–108	1× CVD103- 1× CVD103- 1× CVD103- 1× CVD103- 1× CVD103- 1× CVD103-	ty (Qe 0-22, df 1), p=0-6 HgR High (10 ⁹) HgR High (2x10 ⁹) HgR High (5x10 ⁹) HgR High (10 ¹⁰) HgR High (10 ¹⁰)	28/39 (72%) 40/48 (83%) 17/40 (43%) 30/33 (91%) 26/32 (81%)	Std (10 ⁸) Std (2x10 ⁸) Std (5x10 ⁸) Std (5x10 ⁹) Std (5x10 ⁹)	19/39 (49%) 33/46 (72%) 13/39 (33%) 27/31 (87%) 21/28 (75%)	1·47 (1·01-2·15) 1·16 (0·93-1·45) 1·28 (0·72-2·26) 1·04 (0·88-1·24) 1·08 (0·83-1·42) 1·12 (1·00-1·26)	0-33		-
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Heterogeneity among va Increased vaccine inocul Cholera Sotuzzo (1993) Sow (2017) Sow (2017) Sou-Arehawaratana (1992) Souharyono (1992) Souharyono (1992) Souharyono (1992) Souharyono (1992) Souharyono (1991) Chopra (1981) India (1981) Souharyono (1992)	eccines (Qm 7-3 um Peru Mali) Thailand Indonesia Indonesia India India	5, df 2), p=0-025; rd Adult Adult Adult 60-108 60-108 6-12 1-12 3-36	1× CVD103- 1× CVD103- 1× CVD103- 1× CVD103- 1× CVD103- 1× CVD103- 3× tOPV 5× tOPV 3× tOPV	ty (Qe 0-22, df 1), p=0-6 HgR High (10 ⁹) HgR High (2x10 ⁹) HgR High (5x10 ⁹) HgR High (10 ¹⁰) HgR High (10 ¹⁰) High (0-2 mL) High (0-4 mL) High (0-4 mL)	28/39 (72%) 40/48 (83%) 17/40 (43%) 30/33 (91%) 26/32 (81%) 21/35 (60%) 27/28 (96%) 14/25 (56%)	Std (10 ⁸) Std (2x10 ⁸) Std (5x10 ⁸) Std (5x10 ⁹) Std (5x10 ⁹) Std (0-1 mL) Std (0-2 mL) Std (0-2 mL)	19/39 (49%) 33/46 (72%) 13/39 (33%) 27/31 (87%) 21/28 (75%) 24/42 (57%) 33/34 (97%) 49/87 (55%)	1.47 (1.01-2.15) 1.16 (0.93-1.45) 1.28 (0.72-2.26) 1.04 (0.88-1.24) 1.08 (0.83-1.42) 1.12 (1.00-1.26) 1.05 (0.72-1.53) 0.99 (0.91-1.09) 0.99 (0.67-1.47)	0-33	1.0	-
Aeterogeneity among va Accreased vaccine inocul Cholera Sotuzzo (1993) Sow (2017) Su-Arehawaratana (1992) Suharyono (1992) Suharyono (1992) Summary PPV3 Agarwal (1991) Chopra (1989) hala (1981) Patriarca (1988) NUO (1905)	Peru Mali) Thailand Indonesia Indonesia India India India Brazil	5, df 2), p=0-025; rd Adult Adult Adult 60-108 60-108 60-108 6-12 1-12 3-36 9-6	1× CVD103- 1× CVD103- 1× CVD103- 1× CVD103- 1× CVD103- 1× CVD103- 3× tOPV 3× tOPV 3× tOPV 1× tOPV	ty (Qe 0-22, df 1), p=0-6 HgR High (10 ⁹) HgR High (2x10 ⁹) HgR High (5x10 ⁹) HgR High (10 ¹⁰) HgR High (10 ¹⁰) High (0-2 mL) High (0-4 mL) High (0-4 mL) High (0-4 mL)	28/39 (72%) 40/48 (83%) 17/40 (43%) 30/33 (91%) 26/32 (81%) 21/35 (60%) 27/28 (96%) 14/25 (56%) 31/73 (42%)	Std (10 ⁸) Std (2x10 ⁸) Std (5x10 ⁹) Std (5x10 ⁹) Std (0-1 mL) Std (0-2 mL) Std (0-2 mL) Std (3x10 ⁵)	19/39 (49%) 33/46 (72%) 13/39 (33%) 27/31 (87%) 21/28 (75%) 24/42 (57%) 33/34 (97%) 49/87 (55%) 13/83 (16%)	$\begin{array}{c} 1.47 (1.01-2.15) \\ 1.16 (0.93-1.45) \\ 1.28 (0.72-2.26) \\ 1.04 (0.88-1.24) \\ 1.08 (0.83-1.42) \\ 1.12 (1.00-1.26) \\ 1.12 (1.00-1.26) \\ 1.05 (0.72-1.53) \\ 0.99 (0.91-1.09) \\ 0.99 (0.67-1.47) \\ 2.71 (1.54-4.78) \\ 2.71 (1.54-4.78) \end{array}$	0-33		-
Heterogeneity among va heterogeneity among va heterogeneity among va heterogeneity among va Soutzzo (1993) Sow (2017) Sou-Archawaratana (1992) Souharyono (1992) Souharyono (1992) Souharyono (1992) Soummary DPV3 Agarwal (1991) Chopra (1989) hala (1981) Patriarca (1988) WHO (1995)	eccines (Qm 7-3 um Peru Mali Indonesia Indonesia Indonesia India India Brazil Brazil Brazil	5, df 2), p=0-025; rd Adult Adult Adult 60-108 60-108 60-108 60-12 1-12 3-36 9-6 0	1× CVD103- 1× CVD103- 1× CVD103- 1× CVD103- 1× CVD103- 1× CVD103- 3× tOPV 5× tOPV 3× tOPV 1× tOPV 1× tOPV 4× tOPV	HgR High (10 ⁹) HgR High (2x10 ⁹) HgR High (2x10 ⁹) HgR High (5x10 ⁹) HgR High (10 ¹⁰) HgR High (10 ¹⁰) High (0-2 mL) High (0-4 mL) High (0-4 mL) High (6x10 ⁵) High (6x10 ⁵)†	28/39 (72%) 40/48 (83%) 17/40 (43%) 30/33 (91%) 26/32 (81%) 21/35 (60%) 27/28 (96%) 14/25 (56%) 31/73 (42%) 125/195 (64%)	Std (10 ⁸) Std (2x10 ⁸) Std (5x10 ⁹) Std (5x10 ⁹) Std (0·1 mL) Std (0·2 mL) Std (0·2 mL) Std (3x10 ⁵) Std (3x10 ⁵)	19/39 (49%) 33/46 (72%) 13/39 (33%) 27/31 (87%) 21/28 (75%) 24/42 (57%) 33/34 (97%) 49/87 (56%) 13/83 (16%) 111/176 (63%)	$\begin{array}{c} 1\cdot47 \ (1\cdot01-2\cdot15) \\ 1\cdot16 \ (0\cdot93-1\cdot45) \\ 1\cdot28 \ (0\cdot72-2\cdot26) \\ 1\cdot04 \ (0\cdot88-1\cdot24) \\ 1\cdot08 \ (0\cdot83-1\cdot42) \\ 1\cdot12 \ (1\cdot00-1\cdot26) \\ 1\cdot05 \ (0\cdot72-1\cdot53) \\ 0\cdot99 \ (0\cdot91-1\cdot09) \\ 0\cdot99 \ (0\cdot67-1\cdot47) \\ 2\cdot71 \ (1\cdot54-4\cdot78) \\ 1\cdot02 \ (0\cdot87-1\cdot19) \end{array}$	0-33		-
Heterogeneity among va heterogeneity among va heterogeneity among va heterogeneity among va Sourzzo (1993) Sow (2017) Sou-Arehawaratana (1992) Souharyono (1992) Souharyono (1992) Soummary DPV3 Agarwal (1991) Chopra (1989) hala (1981) Patriarca (1988) WHO (1995) WHO (1995) WHO (1995)	um Peru Mali) Thailand Indonesia Indonesia India India Brazil Brazil Gambia	Adult Adult Adult Adult 60-108 60-108 60-108 60-108 60-108 60-108 60-108 60-108 60-108 60-108 60-108 60-105	1× CVD103- 1× CVD103- 1× CVD103- 1× CVD103- 1× CVD103- 1× CVD103- 3× tOPV 5× tOPV 3× tOPV 1× tOPV 4× tOPV 4× tOPV 4× tOPV	ty (Qe 0-22, df 1), p=0-6 HgR High (10 ⁹) HgR High (2x10 ⁹) HgR High (5x10 ⁹) HgR High (10 ¹⁰) HgR High (10 ¹⁰) High (0-2 mL) High (0-4 mL) High (0-4 mL) High (6x10 ⁵) High (6x10 ⁵)‡	28/39 (72%) 40/48 (83%) 17/40 (43%) 30/33 (91%) 26/32 (81%) 21/35 (60%) 27/28 (95%) 14/25 (56%) 31/73 (42%) 125/195 (64%) 151/161 (71%)	Std (10 ⁸) Std (2x10 ⁸) Std (5x10 ⁸) Std (5x10 ⁹) Std (5x10 ⁹) Std (0-1 mL) Std (0-2 mL) Std (0-2 mL) Std (0-2 mL) Std (3x10 ⁵) Std (3x10 ⁵)	19/39 (49%) 33/46 (72%) 13/39 (33%) 27/31 (87%) 21/28 (75%) 33/34 (97%) 49/87 (56%) 13/83 (16%) 111/176 (63%) 111/175 (70%)	$\begin{array}{c} 1.47 \ (1.01-2.15) \\ 1.16 \ (0.93-1.45) \\ 1.28 \ (0.72-2.26) \\ 1.04 \ (0.88-1.24) \\ 1.08 \ (0.83-1.42) \\ 1.12 \ (1.00-1.26) \\ 1.05 \ (0.72-1.53) \\ 0.99 \ (0.91-1.09) \\ 0.99 \ (0.67-1.47) \\ 2.71 \ (1.54-4.78) \\ 1.02 \ (0.87-1.19) \\ 1.02 \ (0.89-1.18) \\ \end{array}$	0-33		-
Heterogeneity among va heterogeneity among va heterogeneity among va heterogeneity among va Sourzzo (1993) Sow (2017) Su-Arehawaratana (1992) Suharyono (1992) Suharyono (1992) Suharyono (1992) Summary DPV3 Agarwal (1991) Chopra (1989) hala (1981) Patriarca (1988) WHO (1995) Summary 	veccines (Qm 7-3 um Peru Mali Indonesia Indonesia India India India Brazil Brazil Gambia	Adult Adult Adult Adult 60-108 60-108 60-108 6-12 1-12 3-36 9-6 0 0-1.5	1× CVD103- 1× CVD104- 1× CVD104-	ty (Qe 0-22, df 1), p=0-6 HgR High (10 ⁹) HgR High (2x10 ⁹) HgR High (5x10 ⁹) HgR High (10 ¹⁰) High (10 ¹⁰) High (0-2 mL) High (0-4 mL) High (0-4 mL) High (6x10 ⁵)‡ High (6x10 ⁵)‡	28/39 (72%) 40/48 (83%) 17/40 (43%) 30/33 (91%) 26/32 (81%) 21/35 (60%) 27/28 (96%) 14/25 (56%) 31/73 (42%) 125/195 (64%) 151/161 (71%)	Std (10 ⁸) Std (2x10 ⁸) Std (5x10 ⁹) Std (5x10 ⁹) Std (5x10 ⁹) Std (0-1 mL) Std (0-2 mL) Std (0-2 mL) Std (3x10 ⁵) Std (3x10 ⁵)	19/39 (49%) 33/46 (72%) 13/39 (33%) 27/31 (87%) 21/28 (75%) 33/34 (97%) 49/87 (56%) 13/83 (16%) 111/176 (63%) 111/159 (70%)	$\begin{array}{c} 1{\cdot}47(1{\cdot}01{-}2{\cdot}15)\\ 1{\cdot}16(0{\cdot}93{-}1{\cdot}45)\\ 1{\cdot}28(0{\cdot}72{-}2{\cdot}26)\\ 1{\cdot}04(0{\cdot}88{-}1{\cdot}24)\\ 1{\cdot}08(0{\cdot}83{-}1{\cdot}42)\\ 1{\cdot}12(1{\cdot}00{-}1{\cdot}26)\\ 1{\cdot}05(0{\cdot}72{-}1{\cdot}53)\\ 0{\cdot}99(0{\cdot}91{-}1{\cdot}09)\\ 0{\cdot}99(0{\cdot}67{-}1{\cdot}47)\\ 2{\cdot}71(1{\cdot}54{-}4{\cdot}78)\\ 1{\cdot}02(0{\cdot}87{-}1{\cdot}19)\\ 1{\cdot}02(0{\cdot}89{-}1{\cdot}18)\\ 1{\cdot}02(0{\cdot}95{-}1{\cdot}09)\\ \end{array}$	0-33		3
Heterogeneity among va ncreased vaccine inocul Cholera Gotuzzo (1993) 50w (2017) 50-Arehawaratana (1992) Suharyono (1992) Suharyono (1992) Summary DPV3 Agarwal (1991) Chopra (1989) hala (1981) Patriarca (1988) WHO (1995) WHO (1995) Summary Dverall Heterogeneity among va	vecines (Qm 7-3 um Peru Mali) Thailand Indonesia India India India India Brazil Gambia Accines (Qm 2-0	5, df 2), p=0-025; rd Adult Adult Adult 60-108 60-108 60-108 6-12 1-12 3-36 9-6 0 0-1.5 8, df 1), p=0-149; r	1× CVD103- 1× CVD103- 1× CVD103- 1× CVD103- 1× CVD103- 1× CVD103- 1× CVD103- 3× tOPV 5× tOPV 3× tOPV 1× tOPV 4× tOPV 4× tOPV 4× tOPV 2× tOPV	ty (Qe 0-22, df 1), p=0-6 HgR High (10 ⁹) HgR High (2x10 ⁹) HgR High (5x10 ⁹) HgR High (10 ¹⁰) HgR High (10 ¹⁰) High (0-2 mL) High (0-4 mL) High (6x10 ⁵) High (6x10 ⁵) High (6x10 ⁵) High (6x10 ⁵)	28/39 (72%) 40/48 (83%) 17/40 (43%) 30/33 (91%) 26/32 (81%) 21/35 (60%) 27/28 (96%) 14/25 (56%) 31/73 (42%) 125/195 (64%) 151/161 (71%)	Std (10 ⁸) Std (2x10 ⁸) Std (5x10 ⁸) Std (5x10 ⁹) Std (5x10 ⁹) Std (0-1 mL) Std (0-2 mL) Std (0-2 mL) Std (3x10 ⁵) Std (3x10 ⁵) Std (3x10 ⁵)	19/39 (49%) 33/46 (72%) 13/39 (33%) 27/31 (87%) 21/28 (75%) 33/34 (97%) 49/87 (56%) 13/83 (16%) 111/176 (63%) 111/159 (70%)	$\begin{array}{c} 1{\cdot}47(1{\cdot}01{-}2{\cdot}15)\\ 1{\cdot}16(0{\cdot}93{-}1{\cdot}45)\\ 1{\cdot}28(0{\cdot}72{-}2{\cdot}26)\\ 1{\cdot}04(0{\cdot}88{-}1{\cdot}24)\\ 1{\cdot}08(0{\cdot}83{-}1{\cdot}42)\\ 1{\cdot}12(1{\cdot}00{-}1{\cdot}26)\\ 1{\cdot}05(0{\cdot}72{-}1{\cdot}53)\\ 0{\cdot}99(0{\cdot}91{-}1{\cdot}09)\\ 0{\cdot}99(0{\cdot}67{-}1{\cdot}47)\\ 2{\cdot}71(1{\cdot}54{-}78)\\ 1{\cdot}02(0{\cdot}89{-}1{\cdot}19)\\ 1{\cdot}02(0{\cdot}89{-}1{\cdot}19)\\ 1{\cdot}02(0{\cdot}95{-}1{\cdot}09)\\ 1{\cdot}05(0{\cdot}99{-}1{\cdot}11)\\ \end{array}$	0-33		3
Accreased vaccine inocul cholera Sotuzzo (1993) Sow (2017) Sow	veccines (Qm 7-3 um Peru Mali) Thailand Indonesia India India India India Brazil Brazil Brazil Gambia Accines (Qm 2-0	Adult Adult Adult Adult 60-108 60-108 60-108 6-12 1-12 3-36 9-6 0 0-1.5 8, df 1), p=0.149; r	1× CVD103- 1× CVD103- 1× CVD103- 1× CVD103- 1× CVD103- 1× CVD103- 1× CVD103- 3× tOPV 5× tOPV 3× tOPV 4× tOPV 4× tOPV 4× tOPV 4× tOPV 4× tOPV	ty (Qe 0-22, df 1), p=0-6 HgR High (10 ⁹) HgR High (2x10 ⁹) HgR High (5x10 ⁹) HgR High (10 ¹⁰) HgR High (10 ¹⁰) High (0-2 mL) High (0-4 mL) High (6x10 ⁵) High (6x10 ⁵) High (6x10 ⁵) High (6x10 ⁵) ‡	28/39 (72%) 40/48 (83%) 17/40 (43%) 30/33 (91%) 26/32 (81%) 21/35 (60%) 27/28 (96%) 14/25 (56%) 31/73 (42%) 125/195 (64%) 151/161 (71%)	Std (10 ⁸) Std (2x10 ⁸) Std (5x10 ⁸) Std (5x10 ⁹) Std (5x10 ⁹) Std (0-1 mL) Std (0-2 mL) Std (0-2 mL) Std (3x10 ⁵) Std (3x10 ⁵) Std (3x10 ⁵)	19/39 (49%) 33/46 (72%) 13/39 (33%) 27/31 (87%) 21/28 (75%) 24/42 (57%) 33/34 (97%) 49/87 (56%) 13/83 (16%) 111/176 (63%) 111/159 (70%)	$\begin{array}{c} 1\text{-}47(1\text{-}01\text{-}2\text{-}15)\\ 1\text{-}16(0\text{-}93\text{-}1\text{-}45)\\ 1\text{-}28(0\text{-}72\text{-}2\text{-}26)\\ 1\text{-}04(0\text{-}88\text{-}1\text{-}24)\\ 1\text{-}08(0\text{-}83\text{-}1\text{-}42)\\ \textbf{1}\text{-}12(\textbf{1}\text{-}00\text{-}1\text{-}26)\\ \textbf{1}\text{-}05(0\text{-}72\text{-}1\text{-}53)\\ 0\text{-}99(0\text{-}91\text{-}1\text{-}09)\\ 0\text{-}99(0\text{-}71\text{-}47)\\ 2\text{-}71(1\text{-}54\text{-}78)\\ 1\text{-}02(0\text{-}87\text{-}1\text{-}19)\\ 1\text{-}02(0\text{-}85\text{-}1\text{-}19)\\ \textbf{1}\text{-}02(0\text{-}95\text{-}1\text{-}09)\\ \textbf{1}\text{-}05(0\text{-}99\text{-}1\text{-}11)\\ \end{array}$	0-33	1.0	3
Accreased vaccine inocul cholera Sotuzzo (1993) Sow (2017) Sow	veccines (Qm 7-3 um Peru Mali) Thailand Indonesia Indonesia India India India Brazil Brazil Gambia Acccines (Qm 2-0	Adult Adult Adult Adult 60-108 60-108 60-108 60-108 60-108 60-108 60-108 60-108 60-108 8, df 1), p=0-149; r	1× CVD103- 1× CVD103-	ty (Qe 0-22, df 1), p=0-6 HgR High (10 ⁹) HgR High (2x10 ⁹) HgR High (2x10 ⁹) HgR High (10 ¹⁰) High (0-2 mL) High (0-4 mL) High (0-4 mL) High (0-4 mL) High (0-4 mL) High (6x10 ⁵) High (6x10 ⁵)‡ High (6x10 ⁵)‡	28/39 (72%) 40/48 (83%) 17/40 (43%) 30/33 (91%) 26/32 (81%) 21/35 (60%) 27/28 (96%) 14/25 (56%) 31/73 (42%) 125/195 (64%) 151/161 (71%)	Std (10 ⁸) Std (2x10 ⁸) Std (5x10 ⁹) Std (5x10 ⁹) Std (5x10 ⁹) Std (0-1 mL) Std (0-2 mL) Std (0-2 mL) Std (3x10 ⁵) Std (3x10 ⁵)	19/39 (49%) 33/46 (72%) 13/39 (33%) 27/31 (87%) 21/28 (75%) 33/34 (97%) 49/87 (56%) 13/83 (16%) 111/176 (63%) 111/159 (70%)	$\begin{array}{c} 1 \cdot 47 \left(1 \cdot 01 - 2 \cdot 15 \right) \\ 1 \cdot 16 \left(0 \cdot 93 - 1 \cdot 45 \right) \\ 1 \cdot 28 \left(0.72 - 2 \cdot 26 \right) \\ 1 \cdot 04 \left(0 \cdot 88 - 1 \cdot 24 \right) \\ 1 \cdot 08 \left(0 \cdot 83 - 1 \cdot 42 \right) \\ 1 \cdot 12 \left(1 \cdot 00 - 1 \cdot 26 \right) \\ 1 \cdot 05 \left(0 \cdot 72 - 1 \cdot 53 \right) \\ 0 \cdot 99 \left(0 \cdot 91 - 1 \cdot 09 \right) \\ 0 \cdot 99 \left(0 \cdot 67 - 1 \cdot 47 \right) \\ 0 \cdot 99 \left(0 \cdot 67 - 1 \cdot 47 \right) \\ 2 \cdot 71 \left(1 \cdot 54 - 47 \cdot 78 \right) \\ 1 \cdot 02 \left(0 \cdot 89 - 1 \cdot 18 \right) \\ 1 \cdot 02 \left(0 \cdot 99 - 1 \cdot 11 \right) \\ 1 \cdot 05 \left(0 \cdot 99 - 1 \cdot 11 \right) \end{array}$	0-33	1.0 	3
Heterogeneity among va heterogeneity among va heterogeneity among va heterogeneity among va Sourzzo (1993) Sow (2017) Su-Arehawaratana (1992) Suharyono (1992) Suharyono (1992) Suharyono (1992) Suharyono (1992) Suharyono (1992) Summary DPV3 Ataciae (1988) NHO (1995) Summary Detroit (1995) Summary Detro	veccines (Qm 7-3 um Peru Mali) Thailand Indonesia Indonesia India India India Brazil Brazil Gambia Acccines (Qm 2-0	Adult Adult Adult Adult 60-108 60-108 6-12 1-12 3-36 9.6 0 0-1-5 8, df 1), p=0-149; r	1× CVD103- 1× CVD103- 0× tOPV 4× tOPV 4× tOPV 4× tOPV 4× tOPV	ty (Qe 0-22, df 1), p=0-6 HgR High (10 ⁹) HgR High (2x10 ⁹) HgR High (2x10 ⁹) HgR High (10 ¹⁰) High (0-2 mL) High (0-4 mL) High (0-4 mL) High (0-4 mL) High (6x10 ⁵)† High (6x10 ⁵)‡ High (6x10 ⁵)‡	28/39 (72%) 40/48 (83%) 17/40 (43%) 30/33 (91%) 26/32 (81%) 21/35 (60%) 27/28 (96%) 14/25 (56%) 31/73 (42%) 125/195 (64%) 151/161 (71%)	Std (10 ⁸) Std (2x10 ⁸) Std (5x10 ⁹) Std (5x10 ⁹) Std (5x10 ⁹) Std (0-2 mL) Std (0-2 mL) Std (0-2 mL) Std (3x10 ⁵) Std (3x10 ⁵)	19/39 (49%) 33/46 (72%) 13/39 (33%) 27/31 (87%) 21/28 (75%) 24/42 (57%) 33/34 (97%) 49/87 (56%) 13/83 (15%) 111/176 (63%) 111/176 (63%)	1-47 (1-01-2-15) 1-16 (0-93-1-45) 1-28 (0.72-2-26) 1-04 (0-88-1-24) 1-08 (0-83-1-42) 1-12 (1-00-1-26) 1-05 (0.72-1-53) 0-99 (0-91-1-09) 0-99 (0-67-1-47) 2-71 (1-54-4-78) 1-02 (0-89-1-18) 1-02 (0-95-1-09) 1-05 (0-99-1-11)	0-33		3
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Accreased vaccine inocul inolera isourzo (1993) iow (2017) iu-Arehawaratana (1992) iuharyono (1995) VHO (1995) Sturmary DVerall deterogeneity among va PV valence DPV3 isturias (2016) iatriarca (1988) ialeem (2017) utter (2000) iutter (2010) S Vaggie (2012)	Accines (Qm 7-3 um Peru Mali) Thailand Indonesia Indonesia India India Brazil Brazil Gambia Accines (Qm 2-0 Multiple Brazil Pakistan Oman India South Africa	Adult Adult Adult Adult 60-108 60-108 60-108 60-108 6-12 1-12 3-36 9-6 0 0-1-5 8, df 1), p=0-149; r 1-4 9-6 0 9 0 0	1× CVD103- 3× tOPV 1× tOPV 4× tOPV 4× tOPV 4× tOPV 1× OPV	ty (Qe 0-22, df 1), p=0-6 HgR High (10 ⁹) HgR High (2x10 ⁹) HgR High (2x10 ⁹) HgR High (10 ¹⁰) High (0-2 mL) High (0-4 mL) High (0-4 mL) High (6x10 ⁵) High (6x10 ⁵)‡ High (6x10 ⁵)‡ ity (Qe 14-84, df 9), p=0 bOPV mOPV3 bOPV mOPV3 mOPV3 mOPV3 mOPV3	28/39 (72%) 40/48 (83%) 17/40 (43%) 30/33 (91%) 26/32 (81%) 27/28 (96%) 14/25 (56%) 31/73 (42%) 125/195 (64%) 151/161 (71%) *096 *096 *096 *095	Std (10 ⁸) Std (2x10 ⁸) Std (5x10 ⁹) Std (5x10 ⁹) Std (5x10 ⁹) Std (0-1 mL) Std (0-2 mL) Std (0-2 mL) Std (3x10 ⁵) Std (3x10 ⁵) Std (3x10 ⁵) Std (3x10 ⁵) Std (3x10 ⁵)	19/39 (49%) 33/46 (72%) 13/39 (33%) 27/31 (87%) 21/28 (75%) 33/34 (97%) 49/87 (55%) 13/83 (16%) 111/176 (63%) 111/179 (70%) 87/88 (99%) 13/83 (16%) 114/134 (85%) 155/177 (88%) 87/168 (52%) 39/184 (21%)	$\begin{array}{c} 1.47 (1.01-2.15) \\ 1.16 (0.93-1.45) \\ 1.28 (0.72-2.26) \\ 1.04 (0.88-1.24) \\ 1.08 (0.83-1.42) \\ 1.12 (1.00-1.26) \\ 1.05 (0.72-1.53) \\ 0.99 (0.91-1.09) \\ 0.99 (0.67-1.47) \\ 2.71 (1.54-4.78) \\ 1.02 (0.87-1.19) \\ 1.02 (0.89-1.18) \\ 1.02 (0.95-1.09) \\ 1.05 (0.99-1.11) \\ 1.05 (0.99-1.11) \\ 1.05 (0.99-1.12) \\ 1.05 (0.99-1.13) \\ 1.03 (1.93-5.70) \\ 1.10 (1.01-1.20) \\ 1.05 (0.98-1.13) \\ 1.63 (1.39-1.91) \\ 2.73 (2.02-3.70) \end{array}$	0-33		3
<pre>/verail ///// ///// ///// ////// ///// ///// ////</pre>	Accines (Qm 7-3 UM Peru Mali) Thailand Indonesia India India India Brazil Gambia Accines (Qm 2-0 Multiple Brazil Pakistan Oman India South Africa	5, df 2), p=0-025; rd Adult Adult Adult 60-108 6-12 1-12 3-36 9.6 0 0-1-5 8, df 1), p=0-149; r 1-4 9.6 0 9 0 0	1× CVD103- 1× CVD104- 1× CVD105- 3× tOPV 3× tOPV 4× tOPV 4× tOPV 4× tOPV 4× tOPV 1× OPV	ty (Qe 0-22, df 1), p=0-6 HgR High (10 ⁹) HgR High (2x10 ⁹) HgR High (2x10 ⁹) HgR High (10 ¹⁰) High (0-2 mL) High (0-4 mL) High (0-4 mL) High (0-4 mL) High (0-4 mL) High (6x10 ⁵)† High (6x10 ⁵)‡ ity (Qe 14-84, df 9), p=0 bOPV mOPV3 bOPV mOPV3 mOPV3 mOPV3 mOPV3	28/39 (72%) 40/48 (83%) 17/40 (43%) 30/33 (91%) 26/32 (81%) 21/35 (60%) 27/28 (96%) 14/25 (56%) 31/73 (42%) 125/195 (64%) 151/161 (71%) *096 195/198 (98%) 41/79 (52%) 135/144 (94%) 139/165 (84%) 113/195 (58%)	Std (10 ⁸) Std (2x10 ⁸) Std (5x10 ⁹) Std (5x10 ⁹) Std (5x10 ⁹) Std (0-2 mL) Std (0-2 mL) Std (3x10 ⁵) Std (3x10 ⁵) Std (3x10 ⁵) Std (3x10 ⁵) Std (3x10 ⁵)	19/39 (49%) 33/46 (72%) 13/39 (33%) 27/31 (87%) 21/28 (75%) 33/34 (97%) 49/87 (56%) 13/83 (16%) 111/176 (63%) 111/179 (70%) 87/88 (99%) 13/83 (16%) 114/134 (85%) 155/177 (88%) 87/168 (52%) 39/184 (21%)	$\begin{array}{c} 1.47 (1.01-2.15) \\ 1.16 (0.93-1.45) \\ 1.28 (0.72-2.26) \\ 1.04 (0.88-1.24) \\ 1.08 (0.83-1.42) \\ 1.12 (1.00-1.26) \\ 1.05 (0.72-1.53) \\ 0.99 (0.91-1.09) \\ 0.99 (0.67-1.47) \\ 2.71 (1.54-4.78) \\ 1.02 (0.87-1.19) \\ 1.02 (0.89-1.18) \\ 1.02 (0.95-1.09) \\ 1.05 (0.99-1.11) \\ 1.05 (0.99-1.11) \\ 1.05 (0.99-1.12) \\ 3.31 (1.93-5.70) \\ 1.10 (1.01-1.20) \\ 1.05 (0.98-1.13) \\ 1.63 (1.39-1.91) \\ 2.73 (2.02-3.70) \\ 1.54 (1.04-2.28) \\ \end{array}$	0-33		3

Figure 2: Effect of adjuncts or vaccine composition on seroconversion to oral vaccines

Forest plot showing the effects of zinc supplementation, probiotics, increased vaccine inoculum, and OPV valence on seroconversion to OCV, RVV, or OPV. Bbb01=Bifidobacterium breve 01.

Lc ATCC=Lactobacillus casei ATCC. LGG=lactobacillus GG. OPV=oral poliovirus vaccine. bOPV=bivalent OPV. mOPV=monovalent OPV. tOPV=trivalent OPV. RVV=rotavirus vaccine. RV1=Rotarix monovalent vaccine. Qm=Q statistic for moderator effect. *This study also examined another probiotic, Lactobacillus casei, with similar results. We excluded these data from the forest plot to avoid replication of the control group. †This study also included an arm comparing bOPV with tOPV. We excluded these data from the forest plot to avoid replication of the control group.

19 studies investigated changes to vaccine dosing; 14 were eligible for meta-analysis, assessing additional doses (n=8; one on OCV, six on RVV, one on OPV) or additional birth dosing (n=6; five on OPV, one on RVV). There was weak evidence that increasing the number of doses improved seroconversion (RR 1.12, 95% CI



0.96-1.30; figure 3). Two studies reporting RVV efficacy as the primary outcome also showed weak evidence of benefit in increasing from two to three doses.^{36,37} There was weak evidence that an additional birth dose of OPV increased OPV3 seroconversion (RR 1.08, 95% CI 0.99-1.18), and findings remained similar in the combined meta-analysis, which included a rotavirus study³⁸ (1.06, 95% CI 0.98-1.14; appendix p 17).

17 studies reported altered timing of vaccine administration (five narrowed dose interval, four delayed first dose, one early first dose, and seven staggered RVV and OPV administration): 15 were eligible for metaanalysis. We found no evidence that narrowing the dosing interval (from 4-8 weeks to 1-4 weeks, assessed in one study of RVV, two of OPV) benefited seroconversion (RR 0.98, 95% CI 0.94-1.02; appendix p 19), although these trials were primarily designed as non-inferiority studies. Conversely, delaying the first dose of RVV by 4 weeks (four studies) significantly increased seroconversion rates (1.37, 1.15-1.61; figure 3). We found no evidence that staggered administration of RVV and OPV affected OPV seroconversion (1.00, 0.98-1.02), with consistent findings across all OPV serotypes (appendix p 21). By contrast, there was some evidence that children given RVV alone or separated from OPV were more likely to seroconvert against rotavirus compared with children given RVV and OPV concomitantly (1.21, 95% CI 1.00–1.47; figure 3).

Three studies examined the effect of anthelmintics on oral vaccine responses³⁹⁻⁴¹ with no overall effect on seroconversion to OCV (RR 1·26, 95% CI 0·63–2·53) or typhoid (appendix p 15). Only one antibiotic study was identified.⁴² This randomised controlled trial found no effect of oral azithromycin on OPV immunogenicity, although antibiotics reduced faecal biomarkers of environmental enteric dysfunction. There was no evidence that soy-based formulas,⁴³ compared with human milk or conventional formula, affected OPV immunogenicity.

Figure 3: Effect of dosing on seroconversion to oral vaccines

Forest plot showing the effects of delaying the first dose, separating RVV from OPV, and giving extra doses on seroconversion to OCV, OPV, or RVV. IPV=inactivated poliovirus vaccine OPV=oral poliovirus vaccine bOPV=bivalent OPV. mOPV=monovalent OPV. tOPV=trivalent OPV. RVV=rotavirus vaccine. RV1=Rotarix monovalent vaccine. RV5=RotaTeq pentavalent vaccine. Qe=Q statistic for residual heterogeneity. Qm=Q statistic for moderator effect. *Intervention and control group recruited separately. †Centrifuged. ‡Filtered. §Exact sample sizes were not reported for immunogenicity data and were therefore estimated by assuming that loss-to-follow-up rates reported in figure 1 of the trial report were evenly distributed across groups. ‡Study included both a 6-week plus 10-week dose schedule and a 10-week plus 14-week dose schedule. The 10-week plus 14-week schedule was selected as the control group to ensure consistency with other studies and to delineate the effect of extra doses from delayed doses (considered in a separate comparison). ¶Immunogenicity data extracted from Madhi et al¹⁴⁵ and Cunliffe et al.¹⁴⁶ Exact sample sizes were not reported for Malawi data: we therefore assumed that the 85 RVV recipients were distributed 1:1 across the 2-dose and 3-dose schedules (n=42 per group) and used the reported seroconversion rates (47.2% and 57.1%) to estimate the number of infants who seroconverted.

One study of anti-human γ -globulin horse serum and another of inactivated *Escherichia coli* K12 showed a positive effect on OPV and OCV responses, respectively;^{44,45} however, both were downgraded in GRADE assessment to a score of one or less (very low quality evidence) on the basis of study quality and indirectness.

Overall, there was considerable heterogeneity between studies. For three interventions (extra dose or doses, probiotics, and separating RVV from OPV), the intervention effect differed significantly according to vaccine type (appendix p 26); for four interventions (breastfeeding, OPV valence, separating RVV from OPV, and zinc), there was significant residual heterogeneity not explained by vaccine type (appendix p 26). For zinc, the effect on vaccine response was significantly greater for older than for younger children (appendix p 27). For OPV valence and separating RVV from OPV, background immunogenicity was negatively correlated with in-tervention effect (appendix p 27). Notably, the beneficial effect of staggered administration for RVV seroconversion was strongest in two studies with low background immunogenicity that administered only a single dose of RVV.46,47 For withholding breastfeeding, neither age, setting, nor background immunogenicity were sig-nificant secondary moderators (appendix p 27).

Because of variation in timing of post-vaccine titre measurements, we did a sensitivity analysis, excluding 19 studies that measured seroconversion outside our prespecified windows (appendix p 8). The inferences for each intervention remained unaltered in meta-analysis (appendix p 28).

Summary effect sizes for each meta-analysis, grouped by vaccine, are shown in figure 4. Overall, RRs for seroconversion across all interventions ranged between 0.93 and 1.54. For OCV, there was weak evidence that all interventions (except probiotics) improved seroconversion. For RVV, delayed administration of the first dose was the most effective intervention; there was some evidence that staggered RVV and OPV and probiotics increased RVV seroconversion. For OPV, adjusting valence was the most effective intervention. Extra doses of OPV were also effective in one small study.⁴⁸

Discussion

Oral vaccine underperformance is a major public health problem in LMICs. In this systematic review, we identified 87 studies assessing interventions aimed at improving oral vaccine immunogenicity, done over the past four decades. Typically, there were few studies for a given intervention and effect sizes were small. Delayed RVV administration and altered OPV valence were the only effective approaches identified from the available evidence.

Our finding that delaying the first dose of RVV improves seroconversion is consistent with another



Figure 4: Meta-analysis summary

Summary effect sizes from forest plots for each intervention according to vaccine type (OCV, RVV, and OPV). Error bars represent 95% CI and the size of the circles corresponds to the number of participants. OCV=oral cholera vaccine. OPV=oral poliovirus vaccine. RVV=rotavirus vaccine.

systematic review.19 The mechanism for increased immunogenicity when RVV is delayed is probably a combination of less interference from maternal antibodies⁴⁹ and maturation of infant immune function.⁵⁰ Delayed RVV administration might also mitigate the inhibitory effect of OPV; for example, we found some evidence (driven primarily by studies involving a single RVV dose) that concurrent administration of OPV and RVV leads to lower seroconversion to RVV, consistent with in-vitro observations.51 The decision to delay administration of RVV on the basis of our findings should be weighed against the risks of postponing protection against early natural infection, which might increase the risk of severe disease. It would be useful to compare the frequency and severity of infections and cost-effectiveness between the current approach and a delayed strategy using modelling, to better inform scheduling decisions. Reducing OPV valence also showed clear evidence of benefit for seroconversion. Enhanced immunogenicity of monovalent or bivalent OPV compared with trivalent OPV reflects the inhibitory effect of Sabin 2 vaccine virus on the uptake of other serotypes. This finding suggests that the switch from trivalent OPV to bivalent OPV (which does not contain Sabin 2 virus) in 2016-motivated primarily by the need to mitigate the risk of vaccine-derived poliovirus outbreaks52-has probably improved immunity to seretypes 1 and 3.

Several interventions showed weak evidence of benefit, but effect sizes were generally small and the lower bound of the 95% CI did not exclude unity. There was weak evidence of increased seroconversion with oral vaccine buffering, which is designed to protect key components from destruction by gastric acid. However, administering oral vaccines that incorporate sufficient buffer can be challenging in young children, and there might be a reduced effect in this age group because they have little gastric acid.

We found some evidence that increasing vaccine inoculum improved seroconversion to OCV, but not to OPV. For OCV, increasing the inoculum from 1×10^8 to 1×10^9 colony-forming units (CFUs) modestly improved seroconversion rates (RRs of $1 \cdot 16 - 1 \cdot 47$), while further increases (from 5×10^9 to 1×10^{10} CFUs) did not confer any additional benefit.⁵³ Together, these findings suggest that OCV immunogenicity might plateau as the administered dose increases. OPV already contains more than 1×10^5 median cell-culture infectious doses of each Sabin poliovirus serotype, and any further increases in the inoculum might have limited effect.

There was some evidence that additional oral vaccine doses were beneficial for OPV in the single eligible study. This is consistent with the observation from supplementary immunisation programmes that additional doses are immunogenic and that efficacy increases with a constant per-dose response rate.48,54 For RVV, we observed no significant benefit of giving additional Rotarix vaccine (3-5 doses vs 1-3 doses) up until age 22 weeks. However, RVV studies typically compared a schedule of RVV given at age 6, 10, and 14 weeks with RVV given at age 10 weeks and 14 weeks (rather than the standard schedule given at age 6 weeks and 10 weeks for Rotarix), which might explain the lack of benefit observed. A booster dose of RVV at age 9 months induced IgA seroconversion in more than 40% of Bangladeshi infants who had received two previous doses of vaccine,55

highlighting the potential benefit of additional doses in later infancy.

Finally, our meta-analysis showed that a birth dose of OPV had no overall effect on seroconversion rates. However, these data do not capture the benefits of vaccinating at birth to improve coverage.⁵⁶ Moreover, a benefit of vaccination in the neonatal period is protection from early natural infection, which occurs frequently in low-income countries.⁵⁷

For most interventions, we found no evidence of effect on oral vaccine performance. In the meta-analysis, there was no benefit of vitamin A supplementation for OPV or OCV immunogenicity in Asia or Africa. We also found no overall effect of zinc supplementation on oral vaccine responses across five studies, despite its role in mucosal protection.⁵⁸ However, secondary analyses showed that zinc supplementation had a greater effect on oral vaccine responses among older children. Vitamin A and zinc deficiency vary with age and setting, and true deficiency might have been under-represented in the included trials. Three randomised trials showed no effect of temporarily withholding breastfeeding on RVV, live-attenuated OCV (CVD 103-HgR), or OPV immunogenicity. In one trial in Pakistan, there was, paradoxically, weak evidence for increased RVV seroconversion rates in the group with liberalised breastfeeding.⁵⁹ Notably, breastfeeding was only withheld for short periods (mean $3 \cdot 2$ h); however, the feasibility and value of withholding breastfeeding for longer is doubtful. Instead, other novel methods such as vector systems should be pursued to tackle maternal antibody interference.60 The results from our metaanalysis show no significant benefits overall for probiotics, despite findings from individual studies that probiotics can increase levels of specific antibodies to oral vaccines. This finding might be due to the heterogeneity of probiotic strains, reflecting an incomplete understanding of which are most likely to confer benefits. In a previous systematic review of the effects of probiotics on oral and parenteral vaccines, a total of 40 different probiotics were tested, with variations in dose, purity, and timing of administration.¹⁸ The review concluded that the potential benefit of probiotics was strongest for oral vaccines, although only two studies were done among children.

Our review highlights several interventions that merit further study. First, although we found no overall benefit from increasing the number of RVV doses, additional doses given at birth or later in infancy may have a greater impact, particularly as WHO guidelines now allow for later dosing of RVV.⁶¹ There is also interest in early oral vaccination using neonatal strains of rotavirus. One candidate (RV3-BB) was shown to be efficacious against severe gastroenteritis in Indonesian infants.⁶²

Second, of the six probiotic studies included in the qualitative analysis, four were done in European countries and three of these recruited adult participants.

It is therefore difficult to draw conclusions about the value of probiotics in improving oral vaccine efficacy among infants in LMICs. Moreover, the association between bacterial microbiota composition and oral vaccine responses remains equivocal.⁶³ Further studies might help to better define microbiota-directed therapies that will benefit vaccine efficacy.

Third, we did not find any studies exploring the effects of water, sanitation, or hygiene on oral vaccine responses, although one study is underway in Zimbabwe.64 Fourth, no studies investigated adjustments to RVV inoculum (with the exception of early pre-licensure immunogenicity dose-finding trials, which were excluded).65,66 Our search of the grey literature highlighted one trial, underway in Bangladesh, exploring the effects of an increased inoculum on RVV immunogenicity (NCT02992197). Finally, only one antenatal intervention trial was identified in this systematic review.26 In this study, OPV seroconversion was similar among infants of mothers randomised to receive vitamin A or placebo in pregnancy. The antenatal period might provide a window of opportunity to assess the effect of maternal interventions such as macronutrients, micronutrients, probiotics, or antibiotics, since oral vaccine underperformance occurs so early in infancy.

Our study has several limitations. First, we used seroconversion as our primary outcome for metaanalysis; however, serological markers do not always correlate with protection from disease, particularly for RVV, for which rotavirus-specific IgA is a poor correlate of protection in low-income countries.67 Second, there were differences in study design and significant heterogeneity between studies for several interventions, despite our strict inclusion criteria, probably reflecting the range of ages, settings, and vaccine types across studies. However, we undertook meta-regression to provide insights into factors that drive heterogeneity and did sensitivity analyses restricted to studies that measured vaccine immunogenicity during a narrow time window, which did not change our inferences. Third, several studies reported high seroconversion rates in intervention and control groups, which might have obscured detection of a beneficial treatment effect. For example, effect size was negatively correlated with background immunogenicity for valence and separation of RVV from OPV. Studies with high background immunogenicity might also have contributed to the potential publication bias observed for several interventions because their small effect sizes and standard errors might create an apparent skew in funnel plots and Egger's test p values. Finally, comparisons often included small numbers of studies, subdivided by vaccine type, which were sometimes done among adults in high-income countries. Therefore, the available evidence was sometimes insufficient to determine the generalisability of interventions for the most relevant populations of interest (mostly infants in LMICs), which

is apparent from the substantial proportion of studies that were downgraded for indirectness in the GRADE assessment.

Oral vaccines provide tremendous benefits in developing countries; however, they are failing to reach their full potential. Very few strategies substantially improve oral vaccine immunogenicity, and those that do have modest effects. There is a need to better understand the causes of oral vaccine failure to inform more effective interventions; however, overcoming the multiple factors that probably underlie oral vaccine failure in LMICs might be difficult.3 In addition to optimising the effectiveness of intervention approaches, other factors such as safety, feasibility, and affordability should be considered. Immunisation strategies in LMICs must therefore also consider alternatives to oral vaccines. Bypassing the gut through parenteral administration, for example, is an alternative strategy not evaluated in this review. In 2018, a Vi-polysaccharide tetanus-toxoid conjugate typhoid vaccine received WHO pre-qualification, supported by field estimates showing long-term protection,68 and a parenteral rotavirus vaccine has been shown to be immunogenic in South African infants.69 Another strategy being explored is the use of mucosal adjuvants such as dmLT, a detoxified version of Escherichia coli enterotoxin, combined with inactivated poliovirus vaccine.70 However, whether these interventions can be scaled up effectively is unclear. Meanwhile, introducing changes in oral vaccine scheduling, such as deferred RVV dosing, after a careful assessment of the costs and benefits, could improve the effect of oral vaccines for children in LMICs countries and reduce the global burden of diarrhoeal disease.

Contributors

JAC conceived the idea for the review. JAC and EPP designed and undertook the literature review and extracted the data with help from AJP. EPP coded the statistical analysis, figures, and appendix with support from NCG. JAC, EPP, BDK, NCG, and AJP analysed and interpreted the data. JAC wrote the first draft of the manuscript with input from AJP. All authors reviewed and revised subsequent drafts.

Declaration of interests

We declare no competing interests.

For the **extracted data** see Extracted data for all included studies are available online. The analysis

https://osf.io/bemw6/

For the **analysis code** see goo.gl/WqHR7L

For figures and statistical outputs see https://eparker12. github.io/oral_vaccine_ interventions_ metaanalysis_2018/ code is available. All figures and statistical outputs are available in online.

Acknowledgments

We thank Frances Delwiche (Library Associate Professor, University of Vermont) for her advice on the literature search strategy. JAC (grant 201293/Z/16/Z) and AJP (grant 108065/Z/15/Z) are funded by the Wellcome Trust. NCG is funded by the Bill & Melinda Gates Foundation, Medical Research Council, and WHO Polio Research Committee.

References

- GBD 2016 Causes of Death Collaborators. Global, regional, and national age-sex specific mortality for 264 causes of death, 1980–2016: a systematic analysis for the Global Burden of Disease Study 2016. *Lancet* 2017; **390:** 1151–210.
- 2 Tate JE, Burton AH, Boschi-Pinto C, Parashar UD. Global, regional, and national estimates of rotavirus mortality in children <5 years of age, 2000–13. *Clin Infect Dis* 2016; 62 (suppl 2): S96–105.

- Parker EP, Ramani S, Lopman BA, et al. Causes of impaired oral vaccine efficacy in developing countries. *Future Microbiol* 2018; **13**: 97–118.
- Armah GE, Sow SO, Breiman RF, et al. Efficacy of pentavalent rotavirus vaccine against severe rotavirus gastroenteritis in infants in developing countries in sub-Saharan Africa: a randomised, double-blind, placebo-controlled trial. *Lancet* 2010; 376: 606–14.
- 5 Zaman K, Dang DA, Victor JC, et al. Efficacy of pentavalent rotavirus vaccine against severe rotavirus gastroenteritis in infants in developing countries in Asia: a randomised, double-blind, placebo-controlled trial. *Lancet* 2010; **376**: 615–23.
- 6 Ruiz-Palacios GM, Perez-Schael I, Velazquez FR, et al. Safety and efficacy of an attenuated vaccine against severe rotavirus gastroenteritis. N Engl J Med 2006; 354: 11–22.
- 7 Vesikari T, Matson DO, Dennehy P, et al. Safety and efficacy of a pentavalent human-bovine (WC3) reassortant rotavirus vaccine. N Engl J Med 2006; 354: 23–33.
- 8 Parker EP, Ramani S, Lopman BA, et al. Causes of impaired oral vaccine efficacy in developing countries. *Future Microbiol* 2017; 13: 91–118.
- 9 Parker EP, Kampmann B, Kang G, Grassly NC. Influence of enteric infections on response to oral poliovirus vaccine: a systematic review and meta-analysis. J Infect Dis 2014; 210: 853–64.
- 10 Taniuchi M, Platts-Mills JA, Begum S, et al. Impact of enterovirus and other enteric pathogens on oral polio and rotavirus vaccine performance in Bangladeshi infants. *Vaccine* 2016; 34: 3068–75.
- 11 Harris VC, Armah G, Fuentes S, et al. Significant correlation between the infant gut microbiome and rotavirus vaccine response in rural Ghana. J Infect Dis 2017; 215: 34–41.
- 12 Church JA, Parker EP, Kosek MN, et al. Exploring the relationship between environmental enteric dysfunction and oral vaccine responses. *Future Microbiol* 2018; 13: 1055–70.
- 13 Appaiahgari MB, Glass R, Singh S, et al. Transplacental rotavirus IgG interferes with immune response to live oral rotavirus vaccine ORV-116E in Indian infants. *Vaccine* 2014; 32: 651–56.
- 14 John TJ, Devarajan LV, Luther L, Vijayarathnam P. Effect of breast-feeding on seroresponse of infants to oral poliovirus vaccination. *Pediatrics* 1976; 57: 47–53.
- 15 Kazi AM, Cortese MM, Yu Y, et al. Secretor and salivary ABO blood group antigen status predict rotavirus vaccine take in infants. J Infect Dis 2017; 215: 786–89.
- 16 Kaufman DR, De Calisto J, Simmons NL, et al. Vitamin A deficiency impairs vaccine-elicited gastrointestinal immunity. J Immunol 2011; 187: 1877–83.
- Shankar AH, Prasad AS. Zinc and immune function: the biological basis of altered resistance to infection. *Am J Clin Nutr* 1998; 68 (suppl 2): S447–63.
- 18 Zimmermann P, Curtis N. The influence of probiotics on vaccine responses—a systematic review. Vaccine 2018; 36: 207–13.
- 19 Gruber JF, Gruber LM, Weber RP, Becker-Dreps S, Jonsson Funk M. Rotavirus vaccine schedules and vaccine response among infants in low- and middle-income countries: a systematic review. Open Forum Infect Dis 2017; 4: ofx066.
- 20 Tissera MS, Cowley D, Bogdanovic-Sakran N, et al. Options for improving effectiveness of rotavirus vaccines in developing countries. *Hum Vaccin Immunother* 2017; 13: 921–27.
- 21 Cochrane Handbook for Systematic Reviews of Interventions. In: Higgins J, ed. 763 Preparing for data extraction: The Cochrane Collaboration; 2011. http://handbook-5-1.cochrane.org/ (accessed March 28, 2017).
- 22 Ryan R HS. How to GRADE the quality of evidence. 2016. http://cccrg.cochrane.org/author-resources (accessed March 30, 2017).
- 23 Viechtbauer W. Conducting meta-analyses in R with the metafor package. J Stat Softw 2010; 36: 1–48.
- 24 Sidik K, Jonkman JN. A comparison of heterogeneity variance estimators in combining results of studies. *Stat Med* 2007; 26: 1964–81.
- 25 Albert MJ, Qadri F, Wahed MA, et al. Supplementation with zinc, but not vitamin A, improves seroconversion to vibriocidal antibody in children given an oral cholera vaccine. J Infect Dis 2003; 187: 909–13.

- 26 Bhaskaram P, Balakrishna N. Effect of administration of 200000 IU of vitamin A to women within 24 hrs after delivery on response to PPV administered to the newborn. *Indian Pediatr* 1998; 35: 217–22.
- 27 Broome CS, McArdle F, Kyle JA, et al. An increase in selenium intake improves immune function and poliovirus handling in adults with marginal selenium status. *Am J Clin Nutr* 2004; 80: 154–62.
- 28 Taffs RE, Enterline JC, Rusmil K, et al. Oral iodine supplementation does not reduce neutralizing antibody responses to oral poliovirus vaccine. Bull World Health Organ 1999; 77: 484–91.
- 29 Wolvers DAW, van Herpen-Broekmans WMR, Logman MHGM, van der Wielen RPJ, Albers R. Effect of a mixture of micronutrients, but not of bovine colostrum concentrate, on immune function parameters in healthy volunteers: a randomized placebo-controlled study. Nutr J 2006; 5: 28.
- 30 de Vrese M, Rautenberg P, Laue C, Koopmans M, Herremans T, Schrezenmeir J. Probiotic bacteria stimulate virus-specific neutralizing antibodies following a booster polio vaccination. *Eur J Nutr* 2005; 44: 406–13.
- 31 Isolauri E, Joensuu J, Suomalainen H, Luomala M, Vesikari T. Improved immunogenicity of oral D x RRV reassortant rotavirus vaccine by *Lactobacillus casei* GG. *Vaccine* 1995; 13: 310–12.
- 32 Lazarus RP, John J, Shanmugasundaram E, et al. The effect of probiotics and zinc supplementation on the immune response to oral rotavirus vaccine: a randomized, factorial design, placebo-controlled study among Indian infants. *Vaccine* 2018; 36: 273–79.
- 33 Matsuda F, Chowdhury MI, Saha A, et al. Evaluation of a probiotics, *Bifidobacterium breve* BBG-01, for enhancement of immunogenicity of an oral inactivated cholera vaccine and safety: a randomized, double-blind, placebo-controlled trial in Bangladeshi children under 5 years of age. Vaccine 2011; 29: 1855–58.
- 34 Levine MM, Ferreccio C, Black RE, Germanier R. Large-scale field trial of Ty21a live oral typhoid vaccine in enteric-coated capsule formulation. *Lancet* 1987; 1: 1049–52.
- 35 Simanjuntak CH, Paleologo FP, Punjabi NH, et al. Oral immunisation against typhoid fever in Indonesia with Ty21a vaccine. *Lancet* 1991; 338: 1055–59.
- 36 Lanata CF, Black RE, Flores J, et al. Immunogenicity, safety and protective efficacy of one dose of the rhesus rotavirus vaccine and serotype 1 and 2 human-rhesus rotavirus reassortants in children from Lima, Peru. Vaccine 1996; 14: 237–43.
- 37 Madhi SA, Cunliffe NA, Steele D, et al. Effect of human rotavirus vaccine on severe diarrhea in African infants. N Engl J Med 2010; 362: 289–98.
- 38 Bines JE, Danchin M, Jackson P, et al. Safety and immunogenicity of RV3-BB human neonatal rotavirus vaccine administered at birth or in infancy: a randomised, double-blind, placebo-controlled trial. *Lancet Infect Dis* 2015; 15: 1389–97.
- 39 Bhuiyan TR, Choudhury FK, Khanam F, et al. Evaluation of immune responses to an oral typhoid vaccine, Ty21a, in children from 2 to 5 years of age in Bangladesh. *Vaccine* 2014; 32: 1055–60.
- 40 Bruckner S, Agnandji ST, Berberich S, et al. Effect of antihelminthic treatment on vaccine immunogenicity to a seasonal influenza vaccine in primary school children in Gabon: a randomized placebo-controlled trial. *PLoS Negl Trop Dis* 2015; **9**: e0003768.
- 41 Cooper PJ, Chico ME, Losonsky G, et al. Albendazole treatment of children with ascariasis enhances the vibriocidal antibody response to the live attenuated oral cholera vaccine CVD 103-HgR. J Infect Dis 2000; 182: 1199–206.
- 42 Grassly NC, Praharaj I, Babji S, et al. The effect of azithromycin on the immunogenicity of oral poliovirus vaccine: a double-blind randomised placebo-controlled trial in seronegative Indian infants. *Lancet InfectDis* 2016; **16**: 905–14.
- 43 Ostrom KM, Cordle CT, Schaller JP, et al. Immune status of infants fed soy-based formulas with or without added nucleotides for 1 year: part 1: vaccine responses, and morbidity. *J Pediatr Gastroenterol Nutr* 2002; **34**: 137–44.
- 44 Domok I, Balayan MS, Fayinka OA, Skrtic N, Soneji AD, Harland PS. Factors affecting the efficacy of live poliovirus vaccine in warm climates. Efficacy of type 1 Sabin vaccine administered together with antihuman gamma-globulin horse serum to breast-fed and artificially fed infants in Uganda. Bull World Health Organ 1974; 51: 333–47.

- 45 Wasserman SS, Kotloff KL, Losonsky GA, Levine MM. Immunologic response to oral cholera vaccination in a crossover study: a novel placebo effect. Am J Epidemiol 1993; 138: 988–93.
- 46 Giammanco G, De Grandi V, Lupo L, et al. Interference of oral poliovirus vaccine on RIT 4237 oral rotavirus vaccine. *Eur J Epidemiol* 1988; 4: 121–23.
- 47 Vodopija I, Baklaic Z, Vlatkovic R, Bogaerts H, Delem A, Andre FE. Combined vaccination with live oral polio vaccine and the bovine rotavirus RIT 4237 strain. *Vaccine* 1986; 4: 233–36.
- 48 John TJ. Antibody response of infants in tropics to five doses of oral polio vaccine. BMJ 1976; 1: 812.
- 49 Mwila K, Chilengi R, Simuyandi M, Permar SR, Becker-Dreps S. Contribution of maternal immunity to decreased rotavirus vaccine performance in low- and middle-income countries. *Clin Vaccine Immunol* 2017; 24: e00405–16.
- 50 Siegrist CA, Aspinall R. B-cell responses to vaccination at the extremes of age. Nat Rev Immunol 2009; 9: 185–94.
- 51 Wang H, Moon S, Wang Y, Jiang B. Multiple virus infection alters rotavirus replication and expression of cytokines and Toll-like receptors in intestinal epithelial cells. *Virus Res* 2012; 167: 48–55.
- 52 WHO. Global Polio Eradication Initiative (2013). Polio eradication and endgame strategic plan (2013–2018). World Health Organization, Geneva; 2013.
- 53 Subaryono, Simanjuntak C, Witham N, et al. Safety and immunogenicity of single-dose live oral cholera vaccine CVD 103-HgR in 5–9-year-old Indonesian children. *Lancet* 1992; 340: 689–94.
- 4 Grassly NC, Fraser C, Wenger J, et al. New strategies for the elimination of polio from India. *Science* 2006; 314: 1150–53.
- 55 Zaman K, Fleming JA, Victor JC, et al. Noninterference of rotavirus vaccine with measles-rubella vaccine at 9 months of age and improvements in antirotavirus immunity: a randomized trial. J Infect Dis 2016; 213: 1686–93.
- 56 Clark A, Sanderson C. Timing of children's vaccinations in 45 low-income and middle-income countries: an analysis of survey data. *Lancet* 2009; **373**: 1543–49.
- 57 Taniuchi M, Sobuz SU, Begum S, et al. Etiology of diarrhea in Bangladeshi infants in the first year of life analyzed using molecular methods. J Infect Dis 2013; 208: 1794–802.
- 58 Lazzerini M, Wanzira H. Oral zinc for treating diarrhoea in children. Cochrane Database Syst Rev 2016; 12: CD005436.
- 59 Ali A, Kazi AM, Cortese MM, et al. Impact of withholding breastfeeding at the time of vaccination on the immunogenicity of oral rotavirus vaccine--a randomized trial. *PLoS One* 2015; 10: e0127622.
- 50 Niewiesk S. Maternal antibodies: clinical significance, mechanism of interference with immune responses, and possible vaccination strategies. *Front Immunol* 2014; 5: 446.
- 61 WHO. Recommendations for routine immunization-summary tables. http://www.who.int/immunization/policy/immunization_ tables/en/ (accessed May 30, 2017).
- 62 Bines JE, At Thobari J, Satria CD, et al. Human neonatal rotavirus vaccine (RV3-BB) to target rotavirus from birth. N Engl J Med 2018; 378: 719–30.
- 63 Parker EPK, Praharaj I, Zekavati A, et al. Influence of the intestinal microbiota on the immunogenicity of oral rotavirus vaccine given to infants in south India. *Vaccine* 2018; 36: 264–72.
- 64 Church J. The impact of improved water, sanitation and hygiene on oral rotavirus vaccine immunogenicity in Zimbabwean infants: an analysis plan. *Open Sci Framework*; 2018; published online April 30. DOI:10.17605/OSF.IO/BNTU7 (preprint).
- 65 Phua KB, Quak SH, Lee BW, et al. Evaluation of RIX4414, a live, attenuated rotavirus vaccine, in a randomized, double-blind, placebo-controlled phase 2 trial involving 2464 Singaporean infants. J Infect Dis 2005; 192 (suppl 1): S6–16.
- 66 Salinas B, Perez Schael I, Linhares AC, et al. Evaluation of safety, immunogenicity and efficacy of an attenuated rotavirus vaccine, RIX4414: a randomized, placebo-controlled trial in Latin American infants. *Pediatr Infect Dis J* 2005; 24: 807–16.
- 7 Lee B, Carmolli M, Dickson DM, et al. Rotavirus-specific immunoglobulin A responses are impaired and serve as a suboptimal correlate of protection among infants in Bangladesh. *Clin Infect Dis* 2018; 67: 186–92.

- 68 Voysey M, Pollard AJ. Sero-efficacy of Vi-polysaccharide tetanus-toxoid typhoid conjugate vaccine (Typbar-TCV). *Clin Infect Dis* 2018; 67: 18–24.
- 69 Groome MJ, Koen A, Fix A, et al. Safety and immunogenicity of a parenteral P2-VP8-P[8] subunit rotavirus vaccine in toddlers and infants in South Africa: a randomised, double-blind, placebo-controlled trial. *Lancet Infect Dis* 2017; 17: 843–53.
- 70 Norton EB, Bauer DL, Weldon WC, Oberste MS, Lawson LB, Clements JD. The novel adjuvant dmLT promotes dose sparing, mucosal immunity and longevity of antibody responses to the inactivated polio vaccine in a murine model. *Vaccine* 2015; 33: 1909–15.