

1 Title:

2 Presymptomatic atrophy in autosomal dominant Alzheimer's disease: a serial MRI
3 study

4

5 Authors:

6 Kirsi M. Kinnunen^{a,1}, David M. Cash^{a,b,1}, Teresa Poole^{a,c}, Chris Frost^{a,c}, Tammie L.
7 S. Benzinger^d, R. Laila Ahsan^a, Kelvin K. Leung^a, M. Jorge Cardoso^{a,b}, Marc
8 Modat^{a,b}, Ian B. Malone^a, John C. Morris^d, Randall J. Bateman^d, Daniel S. Marcus^d,
9 Alison Goate^e, Stephen Salloway^f, Stephen Correia^f, Reisa A. Sperling^g, Jasmeer P.
10 Chhatwal^g, Richard Mayeux^h, Adam M. Brickman^h, Ralph N. Martinsⁱ, Martin R.
11 Farlow^j, Bernardino Ghetti^j, Andrew J. Saykin^j, Clifford R. Jack Jr^k, Peter R.
12 Schofield^{l,m}, Eric McDade^d, Michael W. Weinerⁿ, John M. Ringman^o, Paul M.
13 Thompson^p, Colin L. Masters^q, Christopher C. Rowe^r, Martin N. Rossor^a, Sebastien
14 Ourselin^{a,b}, Nick C. Fox^{a*}, for the Dominantly Inherited Alzheimer Network (DIAN)

15 ¹*These authors contributed equally to this work*

16

17 ^aDementia Research Centre, UCL Institute of Neurology, London, WC1N 3BG, UK,

18 ^bTranslational Imaging Group, UCL Centre for Medical Image Computing, London,

19 NW1 2HE, UK, ^cLondon School of Hygiene & Tropical Medicine, WC1E 7HT,

20 London, UK, ^dDepartments of Radiology, Neurology, and Neurological Surgery &

21 Psychiatry, Washington University School of Medicine, St. Louis, MO 63110, USA,

22 ^eDepartment of Neuroscience, Icahn School of Medicine at Mount Sinai, New York,

23 NY 10029-5674, USA, ^fBrown University-Butler Hospital, Providence, RI 02903,

24 USA, ^gDepartment of Neurology, Massachusetts General Hospital, Harvard Medical

25 School, Boston, MA 02114, USA, ^hDepartment of Neurology, Columbia University

1 Medical Center, New York, NY 10032, USA, ⁱSchool of Medical Sciences, Edith
2 Cowan University, Joondalup, WA 6027, Australia, ^jDepartments of Neurology,
3 Pathology and Laboratory Medicine and Radiology and Imaging Sciences, Indiana
4 University School of Medicine, Indianapolis, IN 46202, USA, ^kDepartment of
5 Radiology, Mayo Clinic, Rochester, MN 55905, USA, ^lNeuroscience Research
6 Australia, Randwick NSW 2031, Australia, ^mSchool of Medical Sciences, University
7 of New South Wales, Sydney, NSW 2052, Australia, ⁿDepartment of Radiology,
8 School of Medicine, University of California, San Francisco, CA 94143-0628, USA,
9 ^oDepartment of Neurology, Keck USC School of Medicine, Los Angeles, CA 90089,
10 USA, ^pImaging Genetics Center, Keck School of Medicine, University of Southern
11 California, Marina del Rey, CA 90292, USA, ^qThe Florey Institute, The University of
12 Melbourne, Heidelberg VIC 3084, Australia, ^rDepartment of Nuclear Medicine and
13 Centre for PET and Department of Medicine, University of Melbourne, Austin Health,
14 Heidelberg, VIC 3084, Australia

15

16 Corresponding author:

17 David M. Cash

18 Dementia Research Centre, UCL Institute of Neurology

19 Box 16, The National Hospital for Neurology and Neurosurgery

20 Queen Square

21 London WC1N 3BG

22 United Kingdom

23 Telephone: +44 203 448 3054

24 Fax: +44 (0)20 3448 3104

25 Email: d.cash@ucl.ac.uk

1 Abstract

2 INTRODUCTION: Identifying at what point atrophy rates first change in Alzheimer's
3 disease is important for informing design of presymptomatic trials.

4 METHODS: Serial T1-weighted MRI scans of 94 participants (28 non-carriers, 66
5 carriers) from the Dominantly Inherited Alzheimer Network (DIAN) were used to
6 measure brain, ventricular and hippocampal atrophy rates. For each structure, non-
7 linear mixed effects models estimated the change-points when atrophy rates deviate
8 from normal and the rates of change before and after this point.

9 RESULTS: Atrophy increased after the change-point, which occurred 1-1.5 years
10 (assuming a single step change in atrophy rate) or 3-8 years (assuming gradual
11 acceleration of atrophy) before expected symptom onset. At expected symptom
12 onset, estimated atrophy rates were at least 3.6 times those before the change-point.

13 DISCUSSION: Atrophy rates are pathologically increased up to seven years before
14 "expected onset". During this period, atrophy rates may be useful for inclusion and
15 tracking of disease progression.

16

17

18 Keywords: Longitudinal, Atrophy, Alzheimer's disease, Dementia, Autosomal
19 dominant, Neuroimaging, MRI, Boundary Shift Integral, Non-linear modeling,
20 Change-point

1 1. Background

2 Testing potentially disease-modifying treatments for Alzheimer's disease (AD) during
3 the preclinical phase [1] presents challenges of recruitment and staging of
4 asymptomatic individuals, as well as determining suitable measures for assessing
5 disease modification. One recruitment strategy is to study members of families
6 known to carry a pathogenic mutation in a gene – *presenilin 1 (PSEN1)*, *presenilin 2*
7 (*PSEN2*) or *amyloid precursor protein (APP)* – that causes autosomal dominant AD
8 (ADAD). These mutations have almost 100% penetrance and ~50% of at-risk
9 individuals are carriers. ADAD typically has an early and relatively predictable age at
10 symptom onset [2,3]. The Dominantly Inherited Alzheimer Network (DIAN) is a
11 multicentre observational study of individuals at risk of, or affected by, ADAD. DIAN
12 performs longitudinal assessments of imaging, fluid biomarkers, and cognitive
13 function, which reflect pathological features in ADAD [4] and sporadic AD [5]. In
14 particular, cerebral atrophy measures derived from volumetric magnetic resonance
15 imaging (MRI) are used as biomarkers of neurodegeneration and as outcome
16 measures in trials [6].

17

18 Longitudinal data from presymptomatic ADAD individuals provide a unique
19 opportunity to determine when atrophy rates begin to diverge from normal. Previous
20 cross-sectional, or small longitudinal studies report a wide range of estimates of this
21 point of divergence: from 10 years before [4,7] to 7 years after [8] expected clinical
22 onset (as determined by the affected parent's age at onset).

Abbreviations: DIAN = Dominantly Inherited Alzheimer Network; ADAD = autosomal dominantly inherited familial AD; PSEN1 = presenilin 1; PSEN2 = presenilin 2; APP = amyloid precursor protein; EAO = expected age at onset; EYO = estimated years to expected symptom onset; NC = mutation non-carriers; pMut+ = presymptomatic mutation carriers; qMut+ = questionably or mildly symptomatic mutation carriers; sMut+ = overtly symptomatic mutation carriers.

1
2 We used serial MRI data from DIAN to model cerebral atrophy rates during
3 presymptomatic and early symptomatic stages of ADAD. We assessed whole brain
4 and hippocampal atrophy and ventricular expansion, three well-established imaging
5 measures used as exploratory endpoints in clinical trials [6]. We hypothesize that
6 presymptomatic carriers have similar atrophy rates to non-carriers up until a 'change-
7 point' when the biomarker starts to diverge from normal. This hypothesis is
8 consistent with models of sporadic AD [5] that assume a sigmoidal trajectory, and
9 cross-sectional findings from the DIAN cohort [4,7]. We used two non-linear mixed
10 effects models (Supplementary Appendix A) to estimate the timing of change-points
11 relative to expected symptom onset, and atrophy rates before and after these
12 change-points. The first model assumes that the atrophy rate undergoes a single
13 'step change' to a new, stable value; whereas the second model assumes a 'gradual
14 acceleration' in atrophy rate after the change-point. These models help characterize
15 when therapeutic effects on brain atrophy could potentially be observed in
16 presymptomatic ADAD and could help focus future sample size calculations for
17 upcoming prevention trials.

18

19 2. Methods

20 2.1 Participants and Procedures

21 All participants were members of DIAN [9], and details of participating sites are
22 available (<http://dian-info.org/>). The study received prior approval from appropriate
23 Institutional Review Boards and Ethics Committees at each site. Informed consent
24 was obtained from all participants.

25

1 Genotyping was performed to determine the presence of an ADAD mutation for each
2 at-risk participant. A semi-structured interview assessed the expected age at onset
3 (EAO), based on when the affected parent first showed progressive cognitive
4 decline. Expected years to symptom onset (EYO) is the difference between age at
5 scan and EAO [3]. Negative values indicate years before expected onset and
6 positive values years after.

7
8 At the sixth data freeze (July 2013), there were 102 participants with two or more
9 MRI scans available and complete data (mutation status, age, EAO, and global
10 Clinical Dementia Rating (CDR) score [10]).

11 12 2.2 Volumetric MRI

13 Volumetric T1-weighted scans were acquired on 3 Tesla MRI scanners using
14 Alzheimer's Disease Neuroimaging Initiative (ADNI) standardized protocols [11] and
15 corrected for intensity inhomogeneity [12]. Whole brain and hippocampal regions
16 were automatically segmented [13–15]. Lateral ventricles were delineated semi-
17 automatically by an expert rater. Baseline volumetric measures were corrected for
18 total intracranial volume (TIV), calculated using an automated technique [16]. For
19 each structure, volume change was directly measured using a group-wise
20 implementation [17–19] of the Boundary Shift Integral (BSI) [20] to ensure
21 longitudinal consistency. A trained image analyst, blinded to participants' mutation
22 and clinical status, reviewed all raw and processed images.

23 24 2.3 Clinical Classification

1 Participants were classified into four groups, based on mutation status, global CDR
2 score, and actual age at onset (where this had occurred), determined by Uniform
3 Data Set form B9, “Clinical Judgment of symptoms” [21]:

4

5 • **Mutation non-carriers (NC)**; our control group.

6 • **Presymptomatic mutation carriers (pMut+)**; included mutation carriers with
7 a global CDR score of 0 at both their first two visits.

8 • **Questionably or mildly symptomatic mutation carriers (qMut+)**; included
9 participants with at least one global CDR score of 0.5 during their first two
10 visits, with the other visit being either 0 or 0.5. We excluded from this group
11 participants who had a reported onset more than four years before study
12 entry.

13 • **Overtly symptomatic mutation carriers (sMut+)**; included participants with
14 a CDR score of 1.0 or greater at either (or both) of their first two visits or who
15 were more than four years after reported onset at study entry.

16

17 Eight participants were excluded from the analysis: seven (one NC, four pMut+, one
18 qMut+, one sMut+) were identified during initial visual review of the image data and
19 excluded due to non-Alzheimer’s pathology (e.g. infarct, neoplasm), imaging
20 artifacts, or acquisition-related changes likely to result in unreliable atrophy
21 measures. An additional participant (qMut+) was excluded due to moderate motion
22 artefact on follow-up imaging and implausible growth in brain and hippocampi. As
23 part of the sensitivity analysis, we re-ran the model including this participant
24 (Supplementary Appendix B).

25

1 Two participants who initially satisfied the qMut+ criteria were retrospectively re-
2 classified as sMut+, as both participants had consistent evidence of cognitive decline
3 over a sustained period.

4
5 Our final sample therefore included 94 participants: 24 pMut+, 18 qMut+, 24 sMut+,
6 and 28 NC. Of the 66 carriers, 54 had mutations in PSEN1, three in PSEN2, and
7 nine in APP. There were 66 participants with two MR scans, 20 with three, and eight
8 with four scans. The scan interval between baseline to follow-up ranged from 0.9 to
9 3.3 years, and was independent of carrier status or clinical severity. Two participants
10 (one qMut+ and one sMut+) had inadequate image quality for analyses involving
11 hippocampi.

12

13 2.4 Statistical analysis

14 To compare baseline values between each of the three mutations groups (pMut+,
15 qMut+, sMut+) and the non-carrier group, ANOVA models were used for age, EYO,
16 and TIV, while logistic regression was used for APOE ϵ 4 positivity and sex. A
17 generalized least squares linear regression model that allows different group-specific
18 residual variances was used to compare baseline volumes (standardized to mean
19 TIV) between each of the three carrier groups and non-carriers.

20

21 The change-point model [22–24] was used to explore brain, ventricular and
22 hippocampal atrophy rates (Supplementary Appendix A provides a detailed model
23 description). As the focus of our study was the presymptomatic and earliest
24 symptomatic stages of ADAD, the model included non-carriers (NC),
25 presymptomatic, and questionably symptomatic carriers (pMut+/qMut+).

1

2 Figure 1 provides a schematic representation of the 'step change' and 'gradual
3 acceleration' change-point models. In both, β represents the shared atrophy rate for
4 NC and pMut+/qMut+ groups before the change-point, which takes place δ years
5 before or after the EAO. Due to limited data, δ (for a specific brain structure) was
6 assumed to be the same for all pMut+/qMut+ individuals.

7

8 For the 'step change' model, γ is the change in atrophy rate for the pMut+/qMut+
9 group after the change-point. In the 'gradual acceleration' model, the atrophy rate for
10 the pMut+/qMut+ group accelerates after the change-point by a value of 2γ per year.
11 With each model, we estimated β , γ and δ for each region, and using these we
12 estimated atrophy rates at various points before and after EAO.

13

14 Our change-point model was not designed to estimate atrophy rates several years
15 after symptom onset; to do so risked distorting a model that was designed to focus
16 on the progression from early changes to clinical symptoms. Thus, a separate linear
17 mixed-effects random-slopes model (with no change-point) was used to model
18 atrophy rates of the sMut+ group, assuming all observations were after the change-
19 point.

20

21 The change-point models are non-linear extensions of a previously described linear
22 mixed-effects random-slopes model [25] (Supplementary Appendix A). Atrophy
23 measures were log-transformed to provide symmetric approximations of percentage
24 change from baseline. The change-point models were implemented using SAS

1 (version 9.4) procedure NLMIXED, which simultaneously estimated β , γ and δ .
2 Robust estimates of uncertainty for these coefficients were obtained through
3 bootstrapping [26,27], with 10,000 replicates and using bias corrected and
4 accelerated (BCa) 95% confidence intervals. Sensitivity of the estimates and
5 confidence intervals to outliers was explored (see Supplementary Appendix B).

6

7 3. Results

8 Table 1 summarizes demographic and clinical data. The sMut+ group was, as
9 expected, older than the non-carriers, with smaller brain and hippocampal volumes,
10 and larger ventricular volumes (all TIV-adjusted), reflecting pathological losses and
11 larger TIV, which likely reflects the higher (albeit statistically non-significant)
12 proportion of males in this group. The qMut+ group had smaller hippocampal
13 volumes and larger ventricular volumes compared to non-carriers, while the preMut+
14 group just had smaller right hippocampal volumes.

15

16 Table 2 shows the change-point model results for each structure. In the 'step
17 change' model, the pre-change atrophy rate (β) was statistically significant in every
18 structure except the right hippocampus. In all regions, there were significant
19 increases in atrophy rate (γ) after the change-point. This is demonstrated by
20 deriving, from the results of the model, a ratio between the atrophy rate at EAO (1-0
21 years before) to the pre-change atrophy rate. This ratio was 4.0 for whole brain, 4.5
22 for ventricles, and 9.0 for left hippocampus, but it could not be produced for right
23 hippocampus as the estimated pre-change atrophy rate was small and not
24 statistically significantly different from zero. However, the increase in atrophy rate (γ)
25 after the change-point for the right hippocampus was larger than the corresponding

1 coefficient in the results for the left hippocampus. The estimated change-point (δ) for
2 brain, ventricle and left hippocampus was 1.4 years before EAO and 1.1 years
3 before EAO for the right hippocampus. For whole brain and left hippocampus, the
4 confidence intervals for δ did not span zero, providing evidence that they occurred
5 before EAO. Estimates of the ventricular change-point had greater uncertainty (-1.1
6 to 13.5 years) than the other structures. Table 2 provides estimates for rates of
7 change at various times before and after EAO.

8
9 As with the 'step change' model, in the 'gradual acceleration' model all structures
10 except the right hippocampus had statistically significant pre-change atrophy rates.
11 All regions had coefficients (γ) indicating statistically significant increased
12 neurodegeneration after the change-point. The ratio of atrophy rate at EAO to the
13 pre-change rate was 3.6 for whole brain, 4.1 for ventricles, and 5.1 for left
14 hippocampus. The ratio for the right hippocampus was also not available due to the
15 small, non-significant pre-change atrophy rate, but the coefficient (γ) indicated that
16 the right hippocampus had a similar increase towards neurodegeneration as the left.
17 The change-point estimates (δ) for the whole brain and ventricles were 3.0-4.6 years
18 earlier than for the hippocampi. For all structures, the confidence intervals for δ did
19 not span zero. Figure 2 shows estimated atrophy rates and 95% confidence intervals
20 from both models in relation to EYO.

21
22 In the sensitivity analysis, we re-ran the model including the participant with
23 movement artefact and clinically implausible data (Supplementary Appendix B). The
24 pattern of the results was not materially altered although the statistical significance of
25 some parameter estimates was lost.

1

2 The estimated rates of change in sMut+ participants were approximately double
3 those found in pMut+/qMut+ carriers at EAO using the change-point models. The
4 symptomatic rates were: -2.41% (95% CI: -2.88, -1.95) per year for whole brain,
5 15.0% (95% CI: 12.6,17.5) for ventricles, -4.70% (95% CI: -6.39, -3.01) for left
6 hippocampus, and -4.64% (95% CI: -5.68, -3.60) for right hippocampus.

7

8 4. Discussion

9 The goal of this study was to estimate when brain, ventricular and hippocampal
10 volume changes in ADAD diverge from non-carriers, and to model the rates before
11 and after this transition using serial MRI data from the DIAN cohort. We designed
12 two non-linear mixed effects models: one assuming a single 'step-change' and
13 another assuming a 'gradual acceleration' in rates of atrophy after the change-point.
14 This type of model has previously been used to investigate the trajectories of
15 cognitive decline [23,28] and atrophy rates [29,30]. In all cases, there was evidence
16 of increased atrophy after the change-point, suggesting that our models better reflect
17 the non-linear nature of atrophy in early-stage disease than a linear relationship
18 would. The 'gradual acceleration' model found evidence for all assessed regions that
19 atrophy rates diverge from normal values before symptom onset, with the change-
20 point occurring 3.0 to 7.6 years before EAO. The 'step change' model found a
21 change-point of 1.4 years before EAO for whole brain and left hippocampus but was
22 unable to show evidence of a change-point preceding EAO for ventricles or right
23 hippocampus.

24

25 4.1 Interpreting the change-point model results

1 A key advantage of using two different change-point models is that they provide
2 complementary information about the timing of the change-point. The 'step change'
3 model provides the most conservative estimate of when atrophy rates diverge. In
4 contrast, the 'gradual acceleration' model is probably more biologically plausible,
5 based on previous results in ADAD [4,7,31,32] and by the well-characterised spatial
6 spread of neurodegeneration [33] that typically begins in the medial temporal lobe
7 and gradually spreads into neocortical regions. However, there are caveats to the
8 gradual acceleration model used. The non-linear nature of the atrophy may vary
9 between individuals and a quadratic may not be the most appropriate fit. However,
10 given the size of the dataset, this approach minimizes risk of overfittings. Change-
11 point models also avoid some of the pitfalls that can occur when including polynomial
12 terms in a linear regression to model this non-linear relationship [34]. While a
13 quadratic term could better capture the increase in atrophy rate observed around
14 expected onset, it may also produce artefacts of increased atrophy in carriers who
15 are decades before their expected onset.

16

17 Unlike linear models, change-point models can capture the different phases of
18 atrophy/expansion during the long period of presymptomatic disease progression.
19 Both models provide similar estimates of β (see Table 2), the pre-change atrophy
20 rate. This suggested age-related changes broadly consistent with previous aging
21 studies [35–37] showing small but significant rates of whole-brain atrophy of the
22 order of 0.2-0.6%/year and hippocampal atrophy of the order of 0.3-0.4%/year for
23 similar age ranges to this cohort. From both models, there was evidence of
24 increased atrophy after the change-point in all regions.

25

1 4.2 Estimating onset of pathological atrophy

2 It is unclear when disease-related atrophy first becomes evident in ADAD. Cross-
3 sectional results from *PSEN1* E280A mutation carriers [38,39] and DIAN [4,7]
4 suggest atrophy of hippocampi diverge from non-carriers ~6 years and 10 years
5 before symptom onset, respectively; earlier than in our models. However, initial
6 longitudinal results from DIAN [7] (N=53) identified increased atrophy rates only in
7 symptomatic carriers. A study of 13 presymptomatic *PSEN1* carriers found increased
8 cortical thickness at baseline but subsequent thinning of a number of cortical regions
9 [40], suggesting a non-linear nature to presymptomatic changes – with grey matter
10 increases preceding declines.

11
12 Most previous longitudinal volumetric MRI studies of ADAD mutation carriers have
13 been relatively small, single-site studies. One study following presymptomatic
14 participants to clinical onset indicated pathological hippocampal atrophy rates
15 appeared ~5.5 years before AD diagnosis [31]. Weston et al. [41] examined cortical
16 thickness longitudinally in presymptomatic carriers and detected significant losses in
17 the precuneus eight years before EAO. These values are consistent with our findings
18 using a gradual acceleration model where the change point was 7.6 years before
19 onset. However, another study of 16 ADAD mutation carriers (seven with long-term
20 follow-up) did not detect structural MRI changes until *after* symptom onset [8],
21 suggesting that a heterogeneity in these small cohorts and the methods used to
22 analyze them may generate markedly different results.

23
24 No prior ADAD study has used change-point models, making it difficult to compare
25 estimates. However, there are similarities between our findings and sporadic AD

1 studies that used similar approaches. A study of 79 elderly patients, 37 of whom
2 developed mild cognitive impairment (MCI), reported a ventricular expansion
3 change-point 2.3 years before MCI diagnosis [29]. Another longitudinal study
4 (N=296, 66 progressing to MCI) found a similar hippocampal atrophy change-point of
5 2-4 years before clinical onset [30]. Their estimate of a 0.2% per year pre-change
6 hippocampal atrophy rate accords with ours (0.2% left, 0.1% right). Their post-
7 change atrophy rate estimate for the right hippocampus (2.7%/year) was similar to
8 our value (2.5%) whereas their left hippocampal rate estimate (1.2%) was lower than
9 our (2.1%).

10

11 4.3 Predicting clinical onset in ADAD

12 An important challenge is what estimate to use for clinical onset before it has
13 occurred. Many studies, including ours, use an EAO based on when the affected
14 parent first developed symptoms consistent with progressive decline. Other
15 measures are based on the average across all previously affected family members,
16 or the reported age at onset in the literature for a particular mutation [3]. However,
17 each is an imperfect estimate of the future age at onset.

18

19 If future clinical trials use EYO as an inclusion criterion, then it is the distribution of
20 atrophy rates relative to EAO that is of importance. However, if we wish to
21 understand the etiology of the disease, then the distribution of atrophy rates relative
22 to actual onset is more informative, as change-points are likely to be more strongly
23 related to actual rather than expected age at onset. The effect of switching from
24 actual to expected onset in statistical models will change the form of the estimated
25 volume change over time, smoothing it to some degree. Without knowledge of actual

1 onset, this effect is not easily avoided. We did, however, attempt to reduce its impact
2 by excluding overtly symptomatic carriers from our change-point models.

3

4 Identifying precisely when clinical onset has occurred is not straightforward. To
5 facilitate standardization across sites, DIAN rigorously monitors how raters perform
6 CDR and other assessments [42]. In at-risk individuals, other factors can influence
7 cognitive function or behavioral changes, including stress, anxiety, and the constant
8 level of vigilance and introspection that participants experience. In this study, there
9 were six qMut+ participants who reverted from a baseline global CDR of 0.5 to 0 at
10 follow-up. These cases highlight the subtle nature of transitions from unimpaired to
11 “affected” and the potential confounds of mood disturbance and other factors. We
12 addressed this uncertainty by including questionably or mildly symptomatic carriers
13 in our change-point models.

14

15 4.4 Limitations and future work

16 Change-point models have been used to model atrophy rates in preclinical sporadic
17 AD [29,30]. We expand on these approaches by adapting the model for repeated
18 measures of direct change instead of individual volumetric measures and allowing
19 for either a ‘step change’ or ‘gradual acceleration’ after the change-point. Due to the
20 non-linear nature of our models, and the use of bootstrapping to obtain confidence
21 intervals for the model coefficients, these models are susceptible to influential
22 outliers, especially with smaller sample sizes (see the sensitivity analysis in
23 Supplementary Appendix B). Additional longitudinal data should provide improved
24 robustness against such issues.

25

1 No prior study has characterized the progression of atrophy in such a large cohort of
2 presymptomatic and earliest symptomatic ADAD. DIAN is currently recruiting
3 participants into a multicentre clinical trial [43], and the samples from our analysis
4 should more closely reflect a clinical trial setting. Whole brain, lateral ventricles, and
5 hippocampi are the most studied structures in sporadic AD, and are often used as
6 trial outcome measures. From the results, these atrophy measures appear to be
7 elevated compared to non-carriers approximately 5 years before expected onset,
8 making them best suited for prevention trials in ADAD from this period onward.
9 Given the evidence of presymptomatic atrophy in specific cortical regions [40,41],
10 future application of the change-point model could involve studying atrophy rates of
11 specific cortical structures, such as the precuneus and posterior cingulate. Atrophy in
12 these structures may appear earlier and thus be better suited for trials that target
13 presymptomatic patients. In addition, the model should incorporate information from
14 other biomarkers, including CSF amyloid and tau concentrations, to determine how
15 markers of these pathologies affect the timing of the change-point. Finally, it is
16 essential to understand which preclinical changes in ADAD generalize to sporadic
17 AD, as differences in the structures preferentially affected appear to exist [44].

18

19 4.5 Conclusions

20 Atrophy rates increase in ADAD some years before expected symptom onset. Using
21 two different change-point models, we can characterize when this change occurs.
22 The 'step-change' model provides a minimum estimate, 1.4 years before expected
23 onset. The 'gradual acceleration' model provides a more biologically plausible
24 approach towards how atrophy rates diverge from normal, with brain atrophy rates
25 showing pathological acceleration ~7.6 years before expected onset and

- 1 hippocampal rates changing ~3.0 years before expected onset. These models may
- 2 help predict the time to clinical onset for presymptomatic individuals with increased
- 3 atrophy and identify individuals for prevention trials.

1 Figure captions

2

3 **Figure 1: Schematic representation of the ‘step change’ (Figure 1a) and**

4 **‘gradual acceleration’ (Figure 1b) change-point models.**

5

6 **Figure 2: Rates of change estimated from the ‘step change’ and ‘gradual**

7 **acceleration’ models, as a function of the estimated years from symptom**

8 **onset (EYO) for the pMut+/qMut+ carriers.**

9 The figure shows the relationship between rate of annualized volume change (%)
10 and EYO. 95% confidence intervals are included, computed from the bootstrap
11 samples. While the schematics in Figure 1 display the decline in actual volume,
12 these graphs represent the rate of change in volume. A horizontal line indicates the
13 estimated atrophy rate (from the ‘step change’ model) for non-carriers and carriers
14 before the change-point before any deviation from normal rates of change. Vertical
15 dotted lines indicate the change-points for both the ‘step change’ and ‘gradual
16 acceleration’ models. For periods that include the change-point, the estimated rate of
17 atrophy is a weighted combination representing the transition from the pre-change-
18 point atrophy to the post-change-point atrophy. Top left: whole brain; top right: lateral
19 ventricles; bottom left: left hippocampus; bottom right: right hippocampus.

20

21

22

23

24

1 Acknowledgements

2

3 The study sponsors had no role in any aspects of designing or executing this study.

4 The authors had full access to the data used in the study and made the final decision

5 to submit for publication. Data collection and sharing for this project was supported

6 by The Dominantly Inherited Alzheimer's Network (DIAN, U19AG032438), funded by

7 the National Institute on Aging (NIA), the National Institute for Health Research

8 (NIHR) Queen Square Dementia Biomedical Research Unit, the Alzheimer's Society

9 (AS-PG-205) and the Medical Research Council's (MRC) Dementias Platform UK

10 (DPUK). The current study was undertaken at UCLH/UCL who received a proportion

11 of funding from the Department of Health's NIHR Biomedical Research Centres

12 funding scheme. The Dementia Research Centre (DRC) is supported by the UK

13 Dementia Research Institute, Alzheimer's Research UK (ARUK), Brain Research

14 Trust and The Wolfson Foundation. The DRC is also an ARUK coordinating centre,

15 and has received equipment funded by ARUK and the Brain Research Trust. KK

16 reports grants from Anonymous Foundation, during the conduct of the study; grants

17 from ARUK (ARUK-PCRF2014B-1), outside the submitted work. DMC reports grants

18 from Anonymous Foundation and the Alzheimer's Society (AS-PG-205), during the

19 conduct of the study; grants from Anonymous Foundation, Alzheimer's Research

20 UK, and Medical Research Council outside the submitted work. TLSB reports grants

21 from National Institutes of Health (NIH) (U19AG032438, UL1TR000448 and

22 5P30NS04805), during the conduct of the study; grants from Avid

23 Radiopharmaceuticals (Eli Lilly), other from Avid Radiopharmaceuticals (Eli Lilly),

24 other from Roche, other from Medscape LLC, other from Quintiles, outside the

25 submitted work. JCM reports grants from NIH (P50AG005681, P01AG003991,

1 P01AG026276, U19AG032438), during the conduct of the study. MNR reports
2 personal fees from Servier, grants from National Institute for Health Research
3 (NIHR), DIAN, GENFI, DPUK, outside the submitted work. NCF reports personal
4 fees (all paid to University College London directly) from Janssen/Pfizer, GE
5 Healthcare, IXICO, Johnson & Johnson, Genzyme, Eisai, Janssen Alzheimer's
6 Immunotherapy Research and Development, Lilly Research Laboratories (AVID) and
7 Eli Lilly, Novartis Pharma AG, outside the submitted work. In addition, NCF has a
8 patent QA Box issued. RJB reports grants from NIH/NIA U19 AG032438 and
9 Anonymous Foundation, during the conduct of the study; grants from Alzheimer's
10 Association, American Academy of Neurology, Anonymous Foundation,
11 AstraZeneca, BrightFocus Foundation, Cure Alzheimer's Fund, Glenn Foundation for
12 Medical Research, Merck, Metropolitan Life Foundation, NIH, grants from Pharma
13 Consortium (Biogen Idec, Elan Pharmaceuticals Inc., Eli Lilly and Co., Hoffman La-
14 Roche Inc., Genentech Inc., Janssen Alzheimer Immunotherapy, Mithridion Inc.,
15 Novartis Pharma AG, Pfizer Biotherapeutics R and D, Sanofi-Aventi, Eisai), Roche,
16 Ruth K. Broadman Biomedical Research Foundation, NIH/NINDS 2R01NS065667-
17 05, Alzheimer's Association, NIH/NIA (5K23AG030946, P50AG05681), non-financial
18 support from Avid Radiopharmaceuticals, other from C2N Diagnostics, NIH and
19 NIH/State Government Sources, personal fees and other from Washington
20 University, personal fees and non-financial support from Roche, IMI, Sanofi, Global
21 Alzheimer's Platform, FORUM, OECD and Boehringer Ingelheim, personal fees from
22 Merck, outside the submitted work. SS reports grants and personal fees from Lilly,
23 Biogen, Genentech, Roche and Merck, personal fees from Piramal and Forum,
24 grants from GE and Avid, outside the submitted work. AG reports grants from NIH
25 and Anonymous Foundation, during the conduct of the study; personal fees from

1 Cognition Therapeutics and Amgen, grants and non-financial support from
2 Genentech, grants from DIAN PharmaConsortium, outside the submitted work. In
3 addition, AG has a patent (6,083,694, 5,973,133) issued. RAS reports grants from
4 National Institute on Aging, Eli Lilly, Janssen, Bristol Myers Squibb, American Health
5 Assistance Foundation and Alzheimer's Association, during the conduct of the study;
6 personal fees from Eisa, Merck, Boehringer-Ingelheim, Genentech, Roche, Isis,
7 Janssen, Biogen, and Avid Radiopharmaceuticals, outside the submitted work. AMB
8 reports grants from NIH (AG034189, AG043337, AG016495, AG036469,
9 AG037212), during the conduct of study; personal fees from Keystone Heart,
10 personal fees from ProPhase, outside the submitted work. MRF, BG and AJS were
11 supported by NIH grant P30 AG010133 during the study; AJS was supported by
12 additional NIH grants (R01 AG019771, R01 LM011360, R44 AG049540, R01
13 CA129769, and U01 AG032984) during the conduct of the study, and also received
14 grant support from Eli Lilly and PET tracer support from Avid Radiopharmaceuticals,
15 outside the submitted work. CRJ reports grants from NIH, Alexander Family
16 Alzheimer's Disease professorship of the Mayo Foundation, other from Eli Lilly,
17 outside the submitted work. PRS reports grants from NIA, Anonymous Foundation,
18 and Wicking and Mason Trusts, during the conduct of the study; personal fees from
19 ICME Speakers & Entertainers and Janssen-Cilag Pty Ltd, outside the submitted
20 work. MWW reports grants from DOD, NIH/NIA, Veterans Administration,
21 Alzheimer's Association, and Alzheimer's Drug Discovery Foundation, during the
22 conduct of the study; personal fees from Synarc, Janssen, Alzheimer's Drug
23 Discovery Foundation, Neurotrope Bioscience, Merck, Avid, Biogen Idec,
24 Genentech, and Eli Lilly, outside the submitted work. JMR reports grants from NIH,
25 during the conduct of the study; other from Biogen Idec, other from Eli-Lilly, outside

1 the submitted work. All other authors have nothing to disclose. CCR reports personal
2 fees from Roche and GE Healthcare, grants from GE Healthcare, Avid
3 Radiopharmaceuticals and Piramal Imaging, outside the submitted work.

4

5 We gratefully acknowledge the altruism of the participants and their families and the
6 contributions of the DIAN research and support staff at each of the participating
7 sites. In addition, Shona Clegg, Casper Nielsen, Felix Woodward, Emily Manning,
8 Elizabeth Gordon and Josephine Barnes from the Dementia Research Centre
9 assisted with the quality control of automated segmentation and co-registration of
10 regions for longitudinal analysis. Cono Ariti and James Henry Roger from the London
11 School of Hygiene and Tropical Medicine assisted with SAS coding. All authors
12 reviewed the manuscript critically for scientific content and approved the final draft
13 before it was submitted for publication. DIAN Study investigators reviewed the
14 manuscript for consistency of data interpretation with previous DIAN Study
15 publications.

16

1 References

- 2 [1] Golde TE, Schneider LS, Koo EH. Anti- $\alpha\beta$ therapeutics in Alzheimer's disease:
3 the need for a paradigm shift. *Neuron* 2011;69:203–13.
4 doi:10.1016/j.neuron.2011.01.002.
- 5 [2] Ryan NS, Rossor MN. Correlating familial Alzheimer's disease gene mutations
6 with clinical phenotype. *Biomark Med* 2010;4:99–112. doi:10.2217/bmm.09.92.
- 7 [3] Ryman DC, Acosta-Baena N, Aisen PS, Bird T, Danek A, Fox NC, et al.
8 Symptom onset in autosomal dominant Alzheimer disease: A systematic
9 review and meta-analysis. *Neurology* 2014;83:253–60.
10 doi:10.1212/WNL.0000000000000596.
- 11 [4] Bateman RJ, Xiong C, Benzinger TLS, Fagan AM, Goate A, Fox NC, et al.
12 Clinical and Biomarker Changes in Dominantly Inherited Alzheimer's Disease.
13 *N Engl J Med* 2012;367:795–804. doi:10.1056/NEJMoa1202753.
- 14 [5] Jack CR, Knopman DS, Jagust WJ, Petersen RC, Weiner MW, Aisen PS, et
15 al. Tracking pathophysiological processes in Alzheimer's disease: an updated
16 hypothetical model of dynamic biomarkers. *Lancet Neurol* 2013;12:207–16.
17 doi:10.1016/S1474-4422(12)70291-0.
- 18 [6] Cash DM, Rohrer JD, Ryan NS, Ourselin S, Fox NC. Imaging endpoints for
19 clinical trials in Alzheimer's disease. *Alzheimers Res Ther* 2014;6:87.
20 doi:10.1186/s13195-014-0087-9.
- 21 [7] Benzinger TLS, Blazey T, Jack CR, Koeppe RA, Su Y, Xiong C, et al. Regional
22 variability of imaging biomarkers in autosomal dominant Alzheimer's disease.
23 *Proc Natl Acad Sci U S A* 2013;110:E4502-9. doi:10.1073/pnas.1317918110.
- 24 [8] Yau W-YW, Tudorascu DL, McDade EM, Ikonovic S, James JA, Minhas D,
25 et al. Longitudinal assessment of neuroimaging and clinical markers in

- 1 autosomal dominant Alzheimer's disease: a prospective cohort study. *Lancet*
2 *Neurol* 2015;14. doi:10.1016/S1474-4422(15)00135-0.
- 3 [9] Morris JC, Aisen PS, Bateman RJ, Benzinger TLS, Cairns NJ, Fagan AM, et
4 al. Developing an international network for Alzheimer research: The
5 Dominantly Inherited Alzheimer Network. *Clin Investig (Lond)* 2012;2:975–84.
6 doi:10.4155/cli.12.93.
- 7 [10] Morris JC. The Clinical Dementia Rating (CDR): current version and scoring
8 rules. *Neurology* 1993;43:2412–4.
- 9 [11] Jack CR, Bernstein MA, Borowski BJ, Gunter JL, Fox NC, Thompson PM, et
10 al. Update on the magnetic resonance imaging core of the Alzheimer's disease
11 neuroimaging initiative. *Alzheimers Dement* 2010;6:212–20.
12 doi:10.1016/j.jalz.2010.03.004.
- 13 [12] Sled JG, Zijdenbos AP, Evans AC. A nonparametric method for automatic
14 correction of intensity nonuniformity in MRI data. *IEEE Trans Med Imaging*
15 1998;17:87–97. doi:10.1109/42.668698.
- 16 [13] Leung KK, Barnes J, Modat M, Ridgway GR, Bartlett JW, Fox NC, et al. Brain
17 MAPS: an automated, accurate and robust brain extraction technique using a
18 template library. *Neuroimage* 2011;55:1091–108.
19 doi:10.1016/j.neuroimage.2010.12.067.
- 20 [14] Cardoso MJ, Leung K, Modat M, Keihaninejad S, Cash DM, Barnes J, et al.
21 STEPS: Similarity and Truth Estimation for Propagated Segmentations and its
22 application to hippocampal segmentation and brain parcellation. *Med Image*
23 *Anal* 2013;17:671–84. doi:10.1016/j.media.2013.02.006.
- 24 [15] Modat M, Ridgway GR, Taylor ZA, Lehmann M, Barnes J, Hawkes DJ, et al.
25 Fast free-form deformation using graphics processing units. *Comput Methods*

- 1 Programs Biomed 2010;98:278–84. doi:10.1016/j.cmpb.2009.09.002.
- 2 [16] Malone IB, Leung KK, Clegg S, Barnes J, Whitwell JL, Ashburner J, et al.
3 Accurate automatic estimation of total intracranial volume: a nuisance variable
4 with less nuisance. Neuroimage 2014. doi:10.1016/j.neuroimage.2014.09.034.
- 5 [17] Leung KK, Ridgway GR, Ourselin S, Fox NC. Consistent multi-time-point brain
6 atrophy estimation from the boundary shift integral. Neuroimage
7 2012;59:3995–4005. doi:10.1016/j.neuroimage.2011.10.068.
- 8 [18] Leung KK, Clarkson MJ, Bartlett JW, Clegg S, Jack CR, Weiner MW, et al.
9 Robust atrophy rate measurement in Alzheimer’s disease using multi-site
10 serial MRI: Tissue-specific intensity normalization and parameter selection.
11 Neuroimage 2010;50:516–23. doi:10.1016/j.neuroimage.2009.12.059.
- 12 [19] Leung KK, Barnes J, Ridgway GR, Bartlett JW, Clarkson MJ, Macdonald K, et
13 al. Automated cross-sectional and longitudinal hippocampal volume
14 measurement in mild cognitive impairment and Alzheimer’s disease.
15 Neuroimage 2010;51:1345–59. doi:10.1016/j.neuroimage.2010.03.018.
- 16 [20] Freeborough PA, Fox NC. The boundary shift integral: an accurate and robust
17 measure of cerebral volume changes from registered repeat MRI. IEEE Trans
18 Med Imaging 1997;16:623–9. doi:10.1109/42.640753.
- 19 [21] Morris JC, Weintraub S, Chui HC, Cummings J, DeCarli C, Ferris S, et al. The
20 Uniform Data Set (UDS): Clinical and Cognitive Variables and Descriptive Data
21 From Alzheimer Disease Centers. Alzheimer Dis Assoc Disord 2006;20:210–6.
22 doi:10.1097/01.wad.0000213865.09806.92.
- 23 [22] Van Den Hout A, Muniz-Terrera G, Matthews FE. Smooth random change
24 point models. Stat Med 2011;30:599–610. doi:10.1002/sim.4127.
- 25 [23] Hall CB, Lipton RB, Sliwinski M, Stewart WF. A change point model for

- 1 estimating the onset of cognitive decline in preclinical Alzheimer’s disease.
2 Stat Med 2000;19:1555–66.
- 3 [24] Hall CB, Ying J, Kuo L, Lipton RB. Bayesian and profile likelihood change point
4 methods for modeling cognitive function over time. Comput Stat Data Anal
5 2003;42:91–109. doi:10.1016/S0167-9473(02)00148-2.
- 6 [25] Frost C, Kenward MG, Fox NC. The analysis of repeated “direct” measures of
7 change illustrated with an application in longitudinal imaging. Stat Med
8 2004;23:3275–86. doi:10.1002/sim.1909.
- 9 [26] Hyslop T. SAS macros for bootstrap samples with stratification and multiple
10 observations per subject., 1995, p. 805–12.
- 11 [27] Barker N. A Practical Introduction to the Bootstrap Using the SAS System,
12 2005.
- 13 [28] Van Den Hout A, Muniz-Terrera G, Matthews FE. Change point models for
14 cognitive tests using semi-parametric maximum likelihood. Comput Stat Data
15 Anal 2013;57:684–98. doi:10.1016/j.csda.2012.07.024.
- 16 [29] Carlson NE, Moore MM, Dame A, Howieson D, Silbert LC, Quinn JF, et al.
17 Trajectories of brain loss in aging and the development of cognitive
18 impairment. Neurology 2008;70:828–33.
19 doi:10.1212/01.wnl.0000280577.43413.d9.
- 20 [30] Younes L, Albert M, Miller MI. Inferring changepoint times of medial temporal
21 lobe morphometric change in preclinical Alzheimer’s disease. NeuroImage Clin
22 2014;5:178–87. doi:10.1016/j.nicl.2014.04.009.
- 23 [31] Ridha BH, Barnes J, Bartlett JW, Godbolt A, Pepple T, Rossor MN, et al.
24 Tracking atrophy progression in familial Alzheimer’s disease: a serial MRI
25 study. Lancet Neurol 2006;5:828–34. doi:10.1016/S1474-4422(06)70550-6.

- 1 [32] Knight WD, Kim LG, Douiri A, Frost C, Rossor MN, Fox NC. Acceleration of
2 cortical thinning in familial Alzheimer's disease. *Neurobiol Aging*
3 2011;32:1765–73. doi:10.1016/j.neurobiolaging.2009.11.013.
- 4 [33] Braak H, Braak E. Neuropathological staging of Alzheimer-related changes.
5 *Acta Neuropathol* 1991;82:239–59. doi:10.1007/BF00308809.
- 6 [34] Fjell AM, Walhovd KB, Westlye LT, Østby Y, Tamnes CK, Jernigan TL, et al.
7 When does brain aging accelerate? Dangers of quadratic fits in cross-sectional
8 studies. *Neuroimage* 2010;50:1376–83.
9 doi:10.1016/j.neuroimage.2010.01.061.
- 10 [35] Allen JS, Bruss J, Brown CK, Damasio H. Normal neuroanatomical variation
11 due to age: The major lobes and a parcellation of the temporal region.
12 *Neurobiol Aging* 2005;26:1245–60. doi:10.1016/j.neurobiolaging.2005.05.023.
- 13 [36] Fraser MA, Shaw ME, Cherbuin N. A systematic review and meta-analysis of
14 longitudinal hippocampal atrophy in healthy human ageing. *Neuroimage*
15 2015;112:364–74. doi:10.1016/j.neuroimage.2015.03.035.
- 16 [37] Walhovd KB, Westlye LT, Amlie I, Espeseth T, Reinvang I, Raz N, et al.
17 Consistent neuroanatomical age-related volume differences across multiple
18 samples. *Neurobiol Aging* 2011;32:916–32.
19 doi:10.1016/j.neurobiolaging.2009.05.013.
- 20 [38] Quiroz YT, Stern CE, Reiman EM, Brickhouse M, Ruiz A, Sperling RA, et al.
21 Cortical atrophy in presymptomatic Alzheimer's disease presenilin 1 mutation
22 carriers. *J Neurol Neurosurg Psychiatry* 2013;84:556–61. doi:10.1136/jnnp-
23 2012-303299.
- 24 [39] Fleisher AS, Chen K, Quiroz YT, Jakimovich LJ, Gutierrez Gomez M, Langois
25 CM, et al. Associations Between Biomarkers and Age in the Presenilin 1

- 1 E280A Autosomal Dominant Alzheimer Disease Kindred. *JAMA Neurol*
2 2015;72:316. doi:10.1001/jamaneurol.2014.3314.
- 3 [40] Sala-Llonch R, Lladó A, Fortea J, Bosch B, Antonell A, Balasa M, et al.
4 Evolving brain structural changes in PSEN1 mutation carriers. *Neurobiol Aging*
5 2015;36:1261–70. doi:10.1016/j.neurobiolaging.2014.12.022.
- 6 [41] Weston PSJ, Nicholas JM, Lehmann M, Ryan NS, Liang Y, Macpherson K, et
7 al. Presymptomatic cortical thinning in familial Alzheimer disease: A
8 longitudinal MRI study. *Neurology* 2016;87:2050–7.
9 doi:10.1212/WNL.0000000000003322.
- 10 [42] Storandt M, Balota DA, Aschenbrenner AJ, Morris JC. Clinical and
11 psychological characteristics of the initial cohort of the Dominantly Inherited
12 Alzheimer Network (DIAN). *Neuropsychology* 2014;28:19–29.
13 doi:10.1037/neu0000030.
- 14 [43] Bateman RJ, Benzinger TL, Berry S, Clifford DB, Duggan C, Fagan AM, et al.
15 The DIAN-TU Next Generation Alzheimer’s prevention trial: Adaptive design
16 and disease progression model. *Alzheimer’s Dement* 2016:1–12.
17 doi:10.1016/j.jalz.2016.07.005.
- 18 [44] Cash DM, Ridgway GR, Liang Y, Ryan NS, Kinnunen KM, Yeatman T, et al.
19 The pattern of atrophy in familial Alzheimer disease: volumetric MRI results
20 from the DIAN study. *Neurology* 2013;81:1425–33.
21 doi:10.1212/WNL.0b013e3182a841c6.
- 22