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Invited Review

Severe malaria: what's new on the pathogenesis front?

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ABSTRACT

Plasmodium falciparum causes the most severe and fatal form of malaria in humans with over half a million deaths each year. Cerebral malaria (CM), a complex neurological syndrome of severe falciparum malaria, is often fatal and represents a major public health burden. Despite vigorous efforts, the pathophysiology of CM remains to be elucidated, thereby hindering the development of adjunctive therapies. In recent years, multidisciplinary and collaborative approaches have led to groundbreaking progress both in the laboratory and in the field. Here we review the latest breakthroughs in severe malaria pathogenesis, with a specific focus on new pathogenetic mechanisms leading to CM. The most recent findings point towards specific parasite phenotypes targeting brain microvasculature, endothelial dysfunction and subsequent oedema-induced brain swelling.

Keywords: *Plasmodium* spp.; Pathophysiology; Cerebral malaria; Endothelial dysfunction; Sequestration; Malaria in pregnancy

1. Introduction

Malaria is still a leading cause of morbidity and mortality in the developing world. The virulence of *Plasmodium falciparum* is caused by several factors including parasite proteins on the surface of infected erythrocytes (IE). These allow the binding of these cells to the microvascular endothelium of various organs and tissues during infection. Proteins of the *P. falciparum* erythrocyte membrane protein 1 (PfEMP1) family mediate this adhesion through specific binding to multiple cell receptors. These include intercellular adhesion molecule-1 (ICAM-1), CD36, E-selectin, neural cell adhesion molecule (NCAM) and CD31 (PECAM-1) for endothelial beds, as well as chondroitin sulfate A (CSA) for placental syncytiotrophoblasts. Binding to endothelium results in widespread sequestration of IE, which can lead to endothelial activation as well as pro-inflammatory and pro-coagulant responses.

Severe falciparum malaria encompasses a broad range of diseases, the development of which may be influenced by age, exposure and immune status (Wassmer et al., 2015). It includes complications that affect specific organs such as the brain in cerebral malaria (CM) or the placenta in malaria in pregnancy (MiP). Histopathology and laboratory studies allowed investigators to establish a causal link between placenta-specific sequestration of *P. falciparum* and MiP. Indeed, the ability of PfEMP1 variants to target different receptors, the expression of which varies depending on the organ, could explain why some patients with malaria develop organ-specific syndromes. Researchers have speculated that a specific PfEMP1 variant could bind receptors that are preferentially expressed in cerebral microvasculature, and could account for the focal manifestations observed in CM, the most lethal complication of *P. falciparum* infection. Two recent reports simultaneously shed new light on the pathogenetic mechanisms leading to CM. First, endothelial protein C receptor (EPCR) was identified as a binding partner for PfEMP1. Second, normally low levels of EPCR in brain microvessels were shown to be further down-regulated in CM, with a loss of EPCR and thrombomodulin at sites of IE sequestration. These studies provided new clues towards parasite and host cell interactions leading to CM, and connected for the first time brain-specific sequestration of EPCR-binding parasites to the loss of the protein C anti-coagulant function and endothelial cytoprotective pathways (Aird et al., 2014).

While the relative frequency of severe malaria is low, its reported case fatality rate has not substantially changed over decades, especially for CM (Manning et al., 2014). Due to the lack of specific neuro- and vasculoprotective therapies, treatments for CM are currently still precariously limited to antimalarial drugs and

emergency supportive care. The former are quickly dwindling, as the resistance of *P. falciparum* malaria against artemisinin combination treatments, the recommended first-line therapy for infected patients, is on the rise in southeastern Asia. Multi-drug-resistant falciparum malaria is increasingly difficult to treat and new antimalarials are not expected to become available within the next few years. This underlines the necessity for molecular markers for surveillance of partner drug resistance, in conjunction with the implementation of new biomarkers for early diagnosis and outcome prediction, as well as effective adjunct therapies.

Here we review some recent data with a focus on newly developed research approaches aimed at a better understanding of the pathogenetic mechanisms of severe malaria in general and CM in particular.

2. Parasite-brain microvasculature specificity in CM: a virulence factor?

The severity of *P. falciparum* is linked to sequestration of IEs within the microvasculature of various organs including the brain. This sequestration is driven both by the expressed *var* gene in the parasite, leading to the expression of a specific variant of PfEMP-1, and the presence of its associated receptors on microvascular walls (Hviid and Jensen, 2015). Since there are considerable variations in both adhesion molecule expression and functional properties of endothelial cells depending on their position within the vascular bed of a particular tissue, it has been hypothesised that CM may result from a brain endothelial-specific adhesive type of parasite (Moxon et al., 2014). Indeed, IE expressing the domain cassettes (DC) 8 and 13 of the cytoadherent ligand PfEMP-1 adhere to EPCR (Turner et al., 2013). By interfering with EPCR anti-coagulant and pro-endothelial barrier functions, IE adhesion could promote coagulation and vascular permeability that contribute to the pathogenesis of CM (Moxon et al., 2014). To better understand parasite factors that contribute to disease severity, Gillrie et al. (2015) developed in vitro binding models for different microvascular beds to examine the adhesion of DC8- and DC13-expressing parasite lines to endothelial cells from different microvasculature, and the consequences of EPCR engagement on endothelial cell function. They reported that IE from IT4var19 (DC8) and IT4var07 (DC13) parasite lines adhere to human brain, lung and dermal endothelial cells under shear stress. However, the relative contribution of EPCR to parasite cytoadherence on different types of endothelial cell varied.

Divergent functional outcomes for DC8 cysteine-rich interdomain region (CIDR) α 1.1 and DC13 CIDR α 1.4 domains were also observed. IT4var07 CIDR α 1.4 inhibited generation of activated protein C (APC) on lung

and dermal endothelial cells and blocked the APC-EPCR binding interaction on brain endothelial cells. IT4var19 CIDR α 1.1 inhibited thrombin-induced endothelial barrier dysfunction in lung endothelial cells, whereas IT4var07 CIDR α 1.4 inhibited the protective effect of APC on thrombin-induced permeability. Overall, these findings reveal a much greater complexity of how CIDR α 1-expressing parasites may modulate malaria pathogenesis through EPCR adhesion (Gillrie et al., 2015). DC8 PfEMP1 encode multiple endothelial binding domains, including binding activity for EPCR. These results show that PfEMP1 domains compete with protein C for EPCR binding but the extent of competition differs between domains.

Bernabeu et al. (2016) recently investigated these parasite virulence factors in adult patients in India and demonstrated that specific EPCR-binding parasites lead to severe malaria in that population. In addition, parasite phenotype and biomass are associated with patient hospitalization and disease severity. The authors show a broad range of EPCR binding activity from severe malaria isolates and even parasite domains that partially obstructed the interaction between EPCR and APC were sufficient to interfere with the cytoprotective functions of APC (Bernabeu et al., 2016). Taken together, their findings suggest that parasites may be under selection for phenotypic variation in a key host pathway that regulates coagulation and endothelial barrier properties, and has important implications for pathogenic mechanisms in severe malaria. Full-length sequences of PfEMP1 encoding transcripts were characterized in clinical isolates from children with severe malaria admitted to hospital in Tanzania, and EPCR-binding CIDR α 1 domains dominated PfEMP1 transcript profiles of children suffering from CM and/or severe malarial anaemia, further strengthening the evidence for a crucial pathogenic role of the PfEMP1–EPCR interaction in severe malaria (Jespersen et al., 2016).

EPCR is not the only receptor suspected to play an important role in the development of severe malaria. Studies of parasite isolates have demonstrated high rates of in vitro ICAM-1 binding among wild strains but reported correlations between ICAM-1 binding and disease severity have been inconsistent. Recent results suggest that ICAM-1 is a co-receptor for a subset of EPCR-binding parasites (Avril et al., 2016). Indeed, Avril et al. (2016) showed that parasites expressing DC13 have dual binding specificity for EPCR and ICAM-1, suggesting for the first time that ICAM-1-binding variants can be grouped into CD36 and EPCR co-receptor-binding traits. This leads to different cytoadherence abilities on TNF-stimulated endothelial cells, which has important implications for understanding parasite organ-specific microvascular bed tropism in pro-inflammatory conditions.

3. Microvascular endothelial dysfunction: new causes and repercussions

In the recent years, convincing evidence has been presented to support the role of both endothelial cell activation and platelets in modulating the pathogenesis of severe *P. falciparum* malaria. Thrombin, a common factor in both processes, is now thought to be a driver of pathology in CM. The relative contribution of EPCR-binding parasites versus loss of EPCR from the endothelial surface in mediating CM is not known, although both are associated with disease in clinical studies (Turner et al., 2013; Moxon et al., 2014). The key mediator of inflammation in both cases appears to be thrombin, which is a potent mediator of both pro-and anti-inflammatory pathways, depending on the context of endothelial signalling. Thrombin engages thrombomodulin on the plasma membrane of intact endothelium, where it promotes activation of protein C. The latter is accelerated by the presence of EPCR, which binds to protein C and presents it for optimal activation by the thrombin-thrombomodulin complex. Further to its anticoagulant activity, generated APC can also trigger numerous cell-signaling pathways initiating protective cellular responses upon exposure to pro-inflammatory, pro-apoptotic, or toxic insult. IEs expressing PfEMP1 compete with protein C and APC for EPCR, thereby down-regulating protein C activation by the thrombin-thrombomodulin complex. The resulting loss of EPCR-APC-induced cytoprotective signalling is consistent with a decrease in blood-brain barrier (BBB) properties, potentially leading to vasogenic oedema in CM.

Direct protein C pathway alteration by binding of EPCR-specific IEs is not the only cause of endothelial dysfunction in CM. Indeed, *P. falciparum* infection also initiates early endothelial and platelet activation, leading to coagulation dysregulation and microvascular lesions locally (Wassmer et al., 2011a). Furthermore, IEs elicit a myriad of signalling pathways leading to aberrant pro-coagulant effects, ultimately resulting in enhanced endothelial activation, damage and apoptosis (O'Sullivan et al., 2016). Lastly, severe falciparum malaria has been associated with a down-regulation of normal endogenous anticoagulant pathways. EC surface expression of thrombomodulin and EPCR are both reduced, likely through cytokine-enhanced shedding, leading to an increase of their soluble levels in plasma (Moxon et al., 2013). Together, these effects combine and lead to a significant reduction in generation of anti-inflammatory and cytoprotective APC on the endothelial surface. These findings suggest new avenues for acute therapeutic intervention and match well with post-mortem observations and magnetic resonance imaging (MRI) findings linked to mortality in CM.

Such an endothelial dysfunction is not only observed in severe falciparum malaria, but was also reported in severe *Plasmodium vivax* (Barber et al., 2015) and *Plasmodium knowlesi* (Yeo et al., 2007) cases, which raises the question of possible common pathogenic pathways in these various types of infection. All plasmodia species can cause severe and fatal malaria. In falciparum malaria, the most common cause of severe malaria, impaired tissue perfusion has long been recognised to arise from microvascular obstruction by IE adherent to endothelial cells. More recently recognised is the contribution of concurrent impairment of nitric oxide (NO) bioavailability, endothelial activation and microvascular dysfunction to impaired tissue perfusion and severe disease (Yeo et al., 2014). Angiopoietin-2, released from endothelial cell Weibel-Palade bodies and an NO-inhibited autocrine mediator of endothelial activation, is markedly elevated in severe falciparum malaria and consistently associated with impaired tissue perfusion and fatal outcome in both adult and paediatric severe malaria. This is independent of both total and sequestered parasite biomass, suggesting that microvascular obstruction and microvascular dysfunction make separate contributions to pathogenesis. Microvascular function, the capacity to increase flow and oxygen delivery in response to ischaemia, is decreased in severe falciparum malaria and associated with an increased risk of death. Endothelial activation, decreased endothelial NO bioavailability and microvascular dysfunction are also associated with impaired tissue perfusion in vivax and knowlesi malaria, a factor likely to contribute to organ dysfunction and severe malaria caused by infection with these species. Initial clinical trials of agents aimed at restoring NO bioavailability in severe falciparum malaria have recently been completed (Hawkes et al., 2011; Serghides et al., 2011), and microvascular function can be improved by L-arginine infusion in severe falciparum malaria (Yeo et al., 2013). These results suggest that new compounds aimed at increasing microvascular NO and microvascular functions have potential clinical benefits as adjunctive treatments in severe malaria from all species.

4. Rosetting and clumping: consequences for sequestration and microvascular obstruction

Parasite adhesion interactions in severe falciparum malaria are not restricted to the endothelium. Indeed, IEs can bind uninfected erythrocytes to form rosettes (Handunnetti et al., 1989), or platelets to form clumps (Pain et al., 2001). Both processes have been associated with severe malaria (Rowe et al., 1995; Pain et al., 2001) and CM (Carlson et al., 1990; Wassmer et al., 2008).

In this context, recent advances have been made in elucidating the molecular mechanisms underlying rosetting, with a view to development of rosette-reversing therapies. Rosetting generally requires the presence of soluble serum factors such as IgM, and the ability of IE to form rosettes and to bind non-immune IgM is correlated (Rowe et al., 2002). While the role of IgM binding in rosetting remains unclear, it appears to strengthen the bond of the central IE with the surrounding erythrocytes. Stevenson et al. (2015) recently reported that the serum protein α_2 -macroglobulin (α_2 M) is able to induce rosetting in vitro and ex vivo, using several parasite isolates. In contrast to IgM, α_2 M elicits rosetting alone, while the presence of IgM significantly lowers the concentration of α_2 M required. The authors of the study postulate that α_2 M allows the crosslinking of several individual PfEMP1 molecules, thereby increasing their combined avidity for carbohydrate receptors on surrounding erythrocytes and promoting the formation of rosettes. These results suggest that *P. falciparum* hijacks soluble host proteins for its own benefit, and avoids immune destruction by manipulating those to facilitate adhesion of IEs to low-affinity carbohydrate receptors. In addition, another report recently showed that PfEMP1 is not the only parasite ligand used in rosetting, potentially opening new rosette-disrupting approaches. Goel et al. (2015) showed in a very elegant study that the repetitive interspersed family of proteins (RIFINs) mediates binding of IEs with a preference for blood group A, leading to the formation of large rosettes of 10 or more IEs. This was not observed with IEs from group O. Indeed, blood group O is common in malaria endemic areas and IEs of that blood group were shown to form small, weak rosettes (Rowe et al., 2007). The role of RIFINs was confirmed by the disruption of rosettes in the presence of anti-RIFIN antibodies. These results suggest that RIFINs not only play a fundamental role in the development of severe malaria but also contribute to the virulence of *P. falciparum*.

Puzzlingly, formed rosettes have never been described in the peripheral bloodstream, suggesting that they sequester in the microvasculature and aggravate microvascular obstructions during severe malaria in general, and CM in particular. Understanding the mechanisms by which rosettes sequester (i.e., by direct binding of rosetting IE to endothelial cells, to platelets on endothelial cells or to non-rosetting cytoadherent IE is fundamental, as it may open new adjunct therapies to reduce the biomass of sequestered IE, a parameter associated with disease severity (Hendriksen et al., 2012). IE of the IT/R29 strain expressing a rosette-mediating PfEMP1 variant (IT4var09) were recently shown to cytoadhere to human brain microvascular endothelial cells using heparan sulfate proteoglycans as ligands (Adams et al., 2014). This process is distinct

from rosetting, which is primarily mediated by interactions with between the NTS-DBL1 α domain of PfEMP1 and complement receptor 1. This study shows for the first time that IT4var09-expressing parasites are capable of dual interactions with both endothelial cells and uninfected erythrocytes via distinct receptor-ligand interactions, and therefore could aggravate microvascular obstruction in severe malaria by facilitating the sequestration of platelet-mediated clumps.

One of the most recently described *P. falciparum* cytoadherence phenotypes is the ability of IEs to bind to platelets in suspension in vitro to form platelet-mediated clumps. Similar to rosetting, the formation of clumps is a common phenotype and has been reported in a variety of endemic settings (Rowe et al., 2009). This is mediated by the binding of IEs to platelet receptors which include CD36 (Pain et al., 2001), P-selectin (Wassmer et al., 2008) and globular C1q receptor (gC1qR) (Biswas et al., 2007). In all cases, the parasite ligands are unknown, although PfEMP1 is a likely candidate molecule. A recent study performed using clinical isolates from Mozambican patients to evaluate cytoadherence properties such as platelet-mediated clumping, rosetting and adhesion to purified receptors (CD36, ICAM1 and gC1qR) revealed that, compared with matched controls, prevalence of both rosetting and platelet-mediated clumping and adhesion to gC1qR was higher in severe cases (Mayor et al., 2011). Inhibition of these cytoadherence phenotypes may therefore reduce the occurrence or improve the prognosis of severe malaria outcomes. Similar to rosettes, platelet-mediated clumps have not been observed in the bloodstream and it is likely that they sequester in the microvasculature using IE or platelet receptors (or both) to cytoadhere on endothelial cells (Wassmer et al., 2011a). Since this may potentially aggravate microvascular obstruction in CM, further studies aimed at understanding the molecular process involved in their sequestration are warranted.

5. Clinical consequences of BBB opening in CM: vasogenic oedema and brain swelling

Following the report by Seydel et al. (2015), a study was initiated to investigate the different mechanisms potentially responsible for brain swelling in both pediatric and adult CM patients in India (Wassmer et al., 2015). Brain swelling was identified by MRI in over 50% of patients enrolled in the ongoing study, irrespective of their age group. The frequent occurrence of brain swelling in CM has been previously reported in separate studies on Indian adults using computed tomography (CT) (Mohanty et al., 2011), as well as in Bangladeshi adults (Maude et al., 2014) and Malawian children using MRI (Potchen et al., 2012). The cause of the swelling,

which can lead to brain herniation in fatal cases (Seydel et al., 2015), remains to be identified and may be due to cytotoxic or vasogenic causes. Recent results show that patients who develop CM exhibit an over-reaction of their endothelial cells to systemic inflammation, which is not observed in uncomplicated malaria (Wassmer et al., 2011b). Such vulnerability, coupled to a loss of cytoprotective EPCR in the brain, could lead to a disruption of the BBB and the leakage of fluids and proteins from the vascular system into the extracellular space. Also called vasogenic oedema, this process is consistent with the description of ring haemorrhages, a characteristic sign of BBB disruption, in pediatric CM (Taylor et al., 2004; Dorovini-Zis et al., 2011). In addition, these ring hemorrhages correlate positively between retinal and cerebral tissues, owing to their common embryological origin from the neuroectoderm (Barrera et al., 2015; Greiner et al., 2015). Malaria retinopathies seen in pediatric CM are therefore a direct reflection of the neurovascular disease process, and the high frequency of ring haemorrhages both in the brain and in the retina in fatal disease point towards the occurrence of vasogenic oedema. However, evidence for a generalised increase in BBB permeability leading to vasogenic oedema is still debated. A study performed in Thailand showed no evidence of BBB impairment, as radioactively-labelled albumin given intravenously was not found in the cerebrospinal fluid (CSF) of CM patients during coma (Warrell et al., 1986). Additionally, post-mortem analyses of adult Vietnamese patients who died of CM showed that its pathophysiology only involved subtle functional changes in BBB integrity (Brown et al., 2000), and evidence from a different cohort demonstrated that localised loss of vascular integrity did not correlate with the occurrence of pre-mortem coma (Medana et al., 2011). Brain swelling may also occur as a result of cytotoxic oedema, which can result from cells in the cerebral tissue being unable to maintain membrane potential after the failure of Na^+/K^+ ATP-dependent pumps, due to hypoxia or nutrient deprivation. This may cause a redistribution of water from the extra-cellular to the intracellular compartments, ultimately leading to cell swelling, cell death and tissue damage. Ischaemic or hypoxic insults due to mechanical effects of microvascular obstruction by IEs, rosettes and platelet-mediated clumps, as well as nutrient “steal” by local metabolically active parasites could account for cytotoxic oedema in CM. This mechanism is consistent with abundant sequestration in the cerebral microvasculature during the neurologic syndrome (White et al., 2013), and the first in vivo magnetic resonance study of experimental CM revealed a preponderant role for cytotoxic oedema in fatal outcome (Penet et al., 2005).

An increase in intravascular fluid volume within the brain due to sludging of blood flow with sequestration of

IEs, rosettes and platelet-mediated clumps could also cause brain swelling and would explain the diffuse mild brain swelling reported in severe malaria patients from Bangladesh (Maude et al., 2014), without evidence of either cytotoxic or vasogenic oedema. However, cytoadherence itself may not be directly or solely responsible for this clinical syndrome (Storm and Craig, 2014).

Lastly, it is entirely plausible that both mechanisms are not mutually exclusive. A study in live mice using high-field MRI with whole-brain coverage showed that vasogenic oedema occurs first in infected animals, and starts in the olfactory bulb before spreading deeper into the brain along a specific path called the rostral migratory stream, eventually reaching the brain stem. Microvascular pathology and ischemic brain injury develop only secondarily, after vasogenic oedema formation (Hoffmann et al., 2016). Additional MRI studies are currently underway in CM patients from Asia and Africa, and are aiming to elucidate these pathogenetic processes (Wassmer et al., 2015).

6. New adjunct therapies and critical care approaches in severe malaria

Even under optimal conditions, the case–fatality rate in severe malaria treated with either artemisinin derivatives or quinine remains high. In addition, multi-drug-resistant falciparum malaria is increasingly difficult to treat and new effective antimalarial agents are not expected to become available within the next few years. In an effort to reduce malaria-related mortality, numerous adjunctive therapies that may alter severe malaria-induced physiological abnormalities are being evaluated, some of which have been described in other sections of this review. Such therapies are, in nature, highly specific to distinct syndromes of severe malaria, as their aim is to target precise pathophysiological processes. For example, protein C system defects inflicted by the malaria parasite protein PfEMP1 in CM can be overcome by a soluble EPCR variant (Petersen et al., 2015), and could therefore represent a revolutionary approach to dampen the pathogenetic mechanisms and decrease mortality in affected patients. Promising advances in adjunct therapies for other severe malaria syndromes such as acute respiratory distress, acute kidney injury, severe anaemia and metabolic acidosis are still lacking.

The potential clinical benefit of fluid resuscitation was recently investigated and liberal fluid loading in adults with severe malaria showed no improvement in the acidosis and acute kidney injury, but increased the incidence of acute pulmonary oedema (Hanson et al., 2014). However, a more conservative fluid strategy,

using a simple weight-based algorithm, led to a low incidence of acute respiratory distress without significant deterioration in acid-base status, renal function, electrolyte profile or systemic haemodynamics, and was associated with increased survival (Aung et al., 2015).

In addition to *P. falciparum*, severe cases of *P. vivax* and *P. knowlesi* have also been reported, although sequestration is not a specific feature of either infection and the relative contribution of co-morbidities to clinical manifestations, particularly in vivax malaria, remains to be investigated (Wassmer et al., 2015). Both parasitic infections can cause acute pulmonary oedema with a clinical phenotype similar to that seen in severe falciparum malaria. No studies of fluid resuscitation have been performed in these patients, but those are warranted.

7. Novel parasite factors involved in malarial pathogenesis and possible therapeutic targets

In parallel to the development of adjunct therapies, a growing effort in the search for new and effective pharmacotherapies has been triggered by the emergence of multi-drug resistance in *P. falciparum*. While still in their infancy, these approaches are promising and provide a wide range of new therapeutic targets. Several new parasite factors were recently identified and have emerged as potential drug target candidates. Among those, G-quadruplex (G4) DNA motifs and RecQ helicases are newly described players in virulence gene control in *P. falciparum*. G4s are four-stranded structures formed by the stacking of quartets of guanines, and recent work has shown that they can form in vivo as well as in vitro, affecting DNA replication, transcription, translation and telomere maintenance. Harris and Merrick (2015) first demonstrated that DNA helicases, as well as histone-modifying proteins, can influence *var* gene dynamics in *P. falciparum*. Understanding the G4-mediated regulation of the parasite virulence may open the door to novel therapeutic interventions.

Plasmodium falciparum exports parasite-encoded proteins involved in structural and functional remodelling of the host cell. This process is essential for the development of the parasite and is also associated with its virulence (Maier et al., 2009). Molecular chaperones of the heat shock protein (Hsp) family are prominent members of the exportome, including various Hsp40s and one Hsp70. The first biochemical evidence for a specific functional co-chaperone interaction between the exported malarial PFA0660w and PfHsp70-x was recently reported (Daniyan et al., 2016). PFA0660w can stimulate the ATPase activity of PfHsp70-x and work additively with it in a co-chaperone/chaperone interaction, resulting in protein aggregation suppression. The

authors also showed that PFA0660w could potentially act independently as a chaperone. These findings support the proposed role of PfHsp70-x and PFA0660w in parasite protein trafficking and folding in IEs. Further studies are underway to determine the molecular basis for the specificity of this interaction, and to identify inhibitors capable of disrupting it. All of the modifications to the IEs are induced by parasite proteins, most of which initially traffic from the parasite via the secretory pathway to the parasitophorous vacuole. There, proteins interact with a translocon complex called PTEX (*Plasmodium* translocon of exported proteins) that transports them across the vacuole membrane into the IE. Blocking protein export through blocking PTEX function leads to the arrest of parasite growth and loss of virulence (Kalanon et al., 2016), making PTEX an excellent drug target (Gilson et al., 2016).

Post-translational modification of histones is one of the key gene regulation mechanisms during the intra-erythrocyte development cycle of *P. falciparum*. Studies of proteins, which recognise and interact with histone post-translational modifications, are pivotal for understanding *P. falciparum* pathogenesis. Bromodomain proteins bind to acetylated lysines, often on histones, and frequently play a role in regulation of gene expression. *Plasmodium falciparum*-specific bromodomain protein 1 (PfBDP1) binds to chromatin at transcriptional start sites of invasion-related genes and directly controls their expression. Conditional PfBDP1 knockdown causes a significant defect in parasite invasion and growth (Josling et al., 2015). In parallel to these studies, several small molecule inhibitors have recently been reported to have a high affinity and specificity to bromodomains, and could represent a new therapeutic avenue in *P. falciparum* infection (Padmanabhan et al., 2016).

The unique plasticity of the epigenetic regulation in *P. falciparum* has also emerged as a pivotal virulence and pathogenicity factor in recent years. Karmodyia et al. (2015) performed genome-wide mapping of multiple histone modifications of *P. falciparum* and reported H3K36me2 as a global repressive mark, with gene regulation being fine-tuned by the ratio of activation marks to H3K36me2 (Karmodyia et al., 2015). Moreover, *var* genes are mostly poised and marked by a unique set of activation (H4ac) and repression (H3K9me3) marks, which are mutually exclusive to other *P. falciparum* housekeeping genes. A better characterization of epigenetic regulation in *P. falciparum* will lead to the identification of potential therapeutic targets (Ay et al., 2015). Indeed, disrupting the function of proteins responsible for maintaining heterochromatin, such as HP1 (Brancucci et al., 2014), could be an effective strategy to block parasite

replication during the asexual cycle. PfAP2-G, a transcription factor shown to drive gametocytogenesis, also represents a promising target to disrupt malaria transmission (Coleman et al., 2014). Extensive characterisations of the parasite epigenetic factors, as well as its post-transcriptional and translational control processes, are likely to open new avenues for drug development against *P. falciparum*.

8. Pathologies other than CM: MiP

Plasmodium falciparum infection during pregnancy can result in ~~MiP~~, a pathology ~~resulting from~~caused by the accumulation of IEs in the placental intervillous space and the infiltration of maternal monocytes/macrophages (Rogerson et al., 2003), with detrimental outcomes for both the mother and the foetus. Expression of PfEMP1-var2csa at the surface of IEs mediates their adhesion to the placenta. Adaptive immunity is progressively acquired during sequential malaria infections in pregnancy and is mediated by the production of anti-VAR2CSA antibodies, which promote IE adhesion blocking and opsonisation (Desai et al., 2007). This naturally acquired immunity is the key basis for development of a vaccine to protect women during pregnancy, and VAR2CSA is currently the leading candidate (Fried and Duffy, 2015). However, recent findings indicate that broadly neutralising antibodies of multigravidae are not depleted on VAR2CSA recombinant antigens. Using a new approach to assess VAR2CSA domains for functional epitopes recognized by naturally acquired antibodies, Doritchamou et al. (2016) recently demonstrated that different Duffy binding-like (DBL) domain-specific IgG could react to both homologous as well as heterologous antigens and parasites, suggesting that conserved epitopes are shared between allelic variants. In addition, IE binding was blocked by ID1-DBL2-ID2a, DBL4 and DBL5-specific IgG, while partial cross-inhibition activity was observed with purified IgG specific to ID1-DBL2-ID2a and DBL4 antigens. Interestingly, plasma from patients still showed broadly adherence-blocking activity after complete depletion of these VAR2CSA specificities. These results suggest that VAR2CSA vaccines based on a single construct and variant might induce antibodies with limited broad blocking activity, and confirm that a multivalent vaccine comprised of up to five different alleles or with the addition of multiple placental malaria vaccine candidates may be needed to elicit the broad blocking activity observed in African multigravidae (Avril et al., 2010; Hommel et al., 2010).

The generation of protective vaccines is becoming a priority, especially in areas where the prevalence of malaria has decreased due to control and elimination campaigns. Indeed, a recent study conducted in

Mozambique showed a close relationship between antibody levels and the intensity of malaria transmission. Mayor et al. (2015) showed convincing evidence that a decline in the prevalence of malaria documented in the study area was accompanied by reductions in levels of IgG antibodies not only against VAR2CSA, but also against non-pregnancy-specific malaria antigens. In pregnant women with MiP, this was associated with an increase in parasite densities and a higher adverse effect of *P. falciparum* infection on maternal haemoglobin levels and newborn weights (Mayor et al., 2015). Although they also suggest that immunity may be regained as exposure increases, the findings of this study indicate that malaria control and elimination programmes could precede a resurgence of pregnancy-associated malaria pathologies.

Additionally, MiP has also recently been linked to placental pathology in a low malaria transmission area in Brazil, where *P. vivax* is predominant (Souza et al., 2013). While MiP was not associated with severe outcome in a second study performed by the same team in this region, an increased ratio of peripheral receptor tyrosine kinase Tie-2 to angiopoietin (Ang-1) was associated with the occurrence of MiP. Both Ang-1 and Ang-2 had similar magnitudes but inverse associations with placental barrier thickness. MiP is an effect modifier of the association between Ang-1 and placental barrier thickness (Ataide et al., 2015). These findings provide a possible pathway through which placental pathological changes occur during MiP.

Lastly, further to imbalances in cytokine cascades, IE cytoadhesion and angiogenic dysregulation, excessive or dysregulated complement activation as part of the host innate immune response to malaria infection can also exacerbate the severity of MiP, leading to poor pregnancy outcomes (McDonald et al., 2015b). Using an experimental model of MiP in conjunction with micro-CT and HPLC analysis of neurotransmitter levels, McDonald et al. (2015a) showed that complement activation, in particular C5a, contributes to foetal neuropathologic outcomes during MiP. The offspring of infected animals showed persistent neurocognitive deficits in memory and affective-like behaviour compared with unexposed controls (McDonald et al., 2015a). These impairments were linked with decreased tissue levels of neurotransmitters in regions of the brain associated with the deficits. The inhibition of maternal C5a complement receptor signaling restored the levels of neurotransmitters and reversed the associated phenotype, suggesting new targets for intervention in MiP aimed at decreasing foetal neuropathologic outcomes.

9. New investigative tools and experimental models

A vast array of new tools and models has recently become available to facilitate the investigation of severe malaria pathogenesis, with a particular focus on CM. These are detailed elsewhere (Sahu et al., 2015). Further to this, the rise of the 'omic' era during the past decade has provided the malaria research community with unprecedented approaches and technologies to better understand the biology, evolution and pathogenesis of different *Plasmodium* spp. The Malaria Host-Pathogen Interaction Centre (MaHPIC) is a large international systems biology consortium developed in 2012 and based in the USA (<http://www.systemsbiology.emory.edu/index.html>), which allowed the development of a variety of distinctive hypothesis-generating and hypothesis-driven collaborations with scientific teams in malaria-endemic regions. These collaborations involve investigators from several countries in South America, southeastern Asia and sub-Saharan Africa, with a focus on infections caused by the predominant, less predominant, or mixed species of *Plasmodium* (Anderson et al., 2015; Lapp et al., 2015), to study disease states, pathogenesis and physiological or immunobiological questions through the use of untargeted high-resolution metabolomics, as well as clinical and demographic metadata (Salinas et al., 2014). In addition, the MaHPIC team is intensively studying malaria using non-human primate model systems (macaque and New World monkey species, (Joyner et al., 2015)). An overarching goal, beyond the team's specific research quests, is to develop and make available unique large 'omic' datasets (e.g., transcripts, proteins, lipids, metabolites and immune responses), integrated models and tools for the use and benefit of the research community at large. This unique and integrated combination of 'omic' approaches represents a revolutionary platform to identify, develop and evaluate new diagnostic tools, antimalarial drugs and vaccines for different types of malaria parasites.

In parallel, live imaging, including multi-photon approaches, has contributed to the understanding of CM pathogenesis, notably its haemodynamics (Nacer et al., 2014) and its immune cell mobilisation (Pai et al., 2014) components. Recently, in experimental CM, vibrational spectroscopies provided evidence of peroxidative stress and protein oxidation within cerebellar gray matter, which were co-localised with elevated non-haeme iron at the site of microhaemorrhages. A novel combination of chemical probe-free, multimodal imaging to quantify molecular markers of disturbed energy metabolism and peroxidative stress thus provides new insights into understanding CM pathogenesis (Hackett et al., 2015).

10. Conclusions and future perspectives

Despite the recent leap in our understanding of pathogenetic mechanisms leading to severe malaria, the translational outputs to improve the clinical outcome of patients remain meager. Collaborative and multidisciplinary approaches using clinical samples from field sites in endemic areas, in vitro and ex vivo models, as well as animal models of the disease, are crucial to allow global advances in the fight not only against severe falciparum malaria, but also emerging public health issues such as severe malaria caused by *P. vivax* and *P. knowlesi*.

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References

- Adams, Y., Kuhnrae, P., Higgins, M.K., Ghumra, A., Rowe, J.A., 2014. Rosetting *Plasmodium falciparum*-infected erythrocytes bind to human brain microvascular endothelial cells in vitro, demonstrating a dual adhesion phenotype mediated by distinct *P. falciparum* erythrocyte membrane protein 1 domains. *Infect Immun* 82, 949-959.
- Aird, W.C., Mosnier, L.O., Fairhurst, R.M., 2014. *Plasmodium falciparum* picks (on) EPCR. *Blood* 123, 163-167.
- Anderson, D.C., Lapp, S.A., Akinyi, S., Meyer, E.V., Barnwell, J.W., Korir-Morrison, C., Galinski, M.R., 2015. *Plasmodium vivax* trophozoite-stage proteomes. *J Proteomics* 115, 157-176.
- Ataide, R., Murillo, O., Dombrowski, J.G., Souza, R.M., Lima, F.A., Lima, G.F., Hristov, A.D., Valle, S.C., Di Santi, S.M., Epiphanyo, S., Marinho, C.R., 2015. Malaria in Pregnancy Interacts with and Alters the Angiogenic Profiles of the Placenta. *PLoS Negl Trop Dis* 9, e0003824.
- Aung, N.M., Kaung, M., Kyi, T.T., Kyaw, M.P., Min, M., Htet, Z.W., Anstey, N.M., Kyi, M.M., Hanson, J., 2015. The Safety of a Conservative Fluid Replacement Strategy in Adults Hospitalised with Malaria. *PloS One* 10, e0143062.
- Avril, M., Bernabeu, M., Benjamin, M., Brazier, A.J., Smith, J.D., 2016. Interaction between Endothelial Protein C Receptor and Intercellular Adhesion Molecule 1 to Mediate Binding of *Plasmodium falciparum*-Infected Erythrocytes to Endothelial Cells. *mBio* 7, e00615.
- Avril, M., Cartwright, M.M., Hathaway, M.J., Hommel, M., Elliott, S.R., Williamson, K., Narum, D.L., Duffy, P.E., Fried, M., Beeson, J.G., Smith, J.D., 2010. Immunization with VAR2CSA-DBL5 recombinant protein elicits broadly cross-reactive antibodies to placental *Plasmodium falciparum*-infected erythrocytes. *Infect Immun* 78, 2248-2256.
- Ay, F., Bunnik, E.M., Varoquaux, N., Vert, J.P., Noble, W.S., Le Roch, K.G., 2015. Multiple dimensions of epigenetic gene regulation in the malaria parasite *Plasmodium falciparum*: gene regulation via histone modifications, nucleosome positioning and nuclear architecture in *P. falciparum*. *BioEssays : News Rev Mol Cell Dev Biol* 37, 182-194.

Barber, B.E., William, T., Grigg, M.J., Parameswaran, U., Piera, K.A., Price, R.N., Yeo, T.W., Anstey, N.M.,
 2015. Parasite biomass-related inflammation, endothelial activation, microvascular dysfunction and
 disease severity in vivax malaria. PLoS Pathog 11, e1004558.

Barrera, V., Hiscott, P.S., Craig, A.G., White, V.A., Milner, D.A., Beare, N.A., MacCormick, I.J., Kamiza, S.,
 Taylor, T.E., Molyneux, M.E., Harding, S.P., 2015. Severity of retinopathy parallels the degree of parasite
 sequestration in the eyes and brains of malawian children with fatal cerebral malaria. J Infect Dis 211,
 1977-1986.

Bernabeu, M., Danziger, S.A., Avril, M., Vaz, M., Babar, P.H., Brazier, A.J., Herricks, T., Maki, J.N., Pereira, L.,
 Mascarenhas, A., Gomes, E., Chery, L., Aitchison, J.D., Rathod, P.K., Smith, J.D., 2016. Severe adult
 malaria is associated with specific PfEMP1 adhesion types and high parasite biomass. Proc Natl Acad Sci
 (USA), 113, E3270-E3279.

Biswas, A.K., Hafiz, A., Banerjee, B., Kim, K.S., Datta, K., Chitnis, C.E., 2007. *Plasmodium falciparum* uses
 gC1qR/HABP1/p32 as a receptor to bind to vascular endothelium and for platelet-mediated clumping.
 PLoS Pathog 3, 1271-1280.

Brancucci, N.M., Bertschi, N.L., Zhu, L., Niederwieser, I., Chin, W.H., Wampfler, R., Freymond, C., Rottmann,
 M., Felger, I., Bozdech, Z., Voss, T.S., 2014. Heterochromatin protein 1 secures survival and transmission
 of malaria parasites. Cell Host Microbe 16, 165-176.

Brown, H.C., Chau, T.T., Mai, N.T., Day, N.P., Sinh, D.X., White, N.J., Hien, T.T., Farrar, J., Turner, G.D.,
 2000. Blood-brain barrier function in cerebral malaria and CNS infections in Vietnam. Neurology 55, 104-
 111.

Carlson, J., Helmby, H., Hill, A.V., Brewster, D., Greenwood, B.M., Wahlgren, M., 1990. Human cerebral
 malaria: association with erythrocyte rosetting and lack of anti-rosetting antibodies. Lancet 336, 1457-
 1460.

Coleman, B.I., Skillman, K.M., Jiang, R.H., Childs, L.M., Altenhofen, L.M., Ganter, M., Leung, Y., Goldowitz, I.,
 Kafsack, B.F., Marti, M., Llinas, M., Buckee, C.O., Duraisingh, M.T., 2014. A *Plasmodium falciparum*
 histone deacetylase regulates antigenic variation and gametocyte conversion. Cell Host Microbe 16, 177-
 186.

492 Daniyan, M.O., Boshoff, A., Prinsloo, E., Pesce, E.R., Blatch, G.L., 2016. The Malarial Exported PFA0660w Is
 493 an Hsp40 Co-Chaperone of PfHsp70-x. PloS One 11, e0148517.

494 Desai, M., ter Kuile, F.O., Nosten, F., McGready, R., Asamo, K., Brabin, B., Newman, R.D., 2007.
 495 Epidemiology and burden of malaria in pregnancy. Lancet. Infect Dis 7, 93-104.

496 Doritchamou, J.Y., Herrera, R., Aebig, J.A., Morrison, R., Nguyen, V., Reiter, K., Shimp, R., MacDonald, N.J.,
 497 Narum, D.L., Fried, M., Duffy, P.E., 2016. VAR2CSA domain-specific analysis of naturally acquired
 498 functional antibodies to *P. falciparum* placental malaria. J Infect Dis, 214. 577-586.

499 Dorovini-Zis, K., Schmidt, K., Huynh, H., Fu, W., Whitten, R.O., Milner, D., Kamiza, S., Molyneux, M., Taylor,
 500 T.E., 2011. The neuropathology of fatal cerebral malaria in malawian children. Am J Pathol 178, 2146-
 501 2158.

502 Fried, M., Duffy, P.E., 2015. Designing a VAR2CSA-based vaccine to prevent placental malaria. Vaccine 33,
 503 7483-7488.

504 Gillrie, M.R., Avril, M., Brazier, A.J., Davis, S.P., Stins, M.F., Smith, J.D., Ho, M., 2015. Diverse functional
 505 outcomes of *Plasmodium falciparum* ligation of EPCR: potential implications for malarial pathogenesis.
 506 Cell Micro 17, 1883-1899.

507 Gilson, P.R., Chisholm, S.A., Crabb, B.S., de Koning-Ward, T.F., 2016. Host cell remodelling in malaria
 508 parasites: a new pool of potential drug targets. Int J Parasitol. [This issue].

509 Goel, S., Palmkvist, M., Moll, K., Joannin, N., Lara, P., Akhouri, R.R., Moradi, N., Ojemalm, K., Westman, M.,
 510 Angeletti, D., Kjellin, H., Lehtio, J., Blixt, O., Idestrom, L., Gahmberg, C.G., Storry, J.R., Hult, A.K., Olsson,
 511 M.L., von Heijne, G., Nilsson, I., Wahlgren, M., 2015. RIFINs are adhesins implicated in severe
 512 *Plasmodium falciparum* malaria. Nat Med 21, 314-317.

513 Greiner, J., Dorovini-Zis, K., Taylor, T.E., Molyneux, M.E., Beare, N.A., Kamiza, S., White, V.A., 2015.
 514 Correlation of hemorrhage, axonal damage, and blood-tissue barrier disruption in brain and retina of
 515 Malawian children with fatal cerebral malaria. Front Cell Infect Microbiol 5, 18.

516 Hackett, M.J., Aitken, J.B., El-Assaad, F., McQuillan, J.A., Carter, E.A., Ball, H.J., Tobin, M.J., Paterson, D., de
 517 Jonge, M.D., Siegele, R., Cohen, D.D., Vogt, S., Grau, G.E., Hunt, N.H., Lay, P.A., 2015. Mechanisms of
 518 murine cerebral malaria: Multimodal imaging of altered cerebral metabolism and protein oxidation at
 519 hemorrhage sites. Sci Adv 1, e1500911.

520 Handunnetti, S.M., David, P.H., Perera, K.L., Mendis, K.N., 1989. Uninfected erythrocytes form "rosettes"
 521 around *Plasmodium falciparum* infected erythrocytes. Am J Trop Med Hyg 40, 115-118.

522 Hanson, J., Anstey, N.M., Bihari, D., White, N.J., Day, N.P., Dondorp, A.M., 2014. The fluid management of
 523 adults with severe malaria. Crit Care 18, 642.

524 Harris, L.M., Merrick, C.J., 2015. G-quadruplexes in pathogens: a common route to virulence control? PLoS
 525 Pathog 11, e1004562.

526 Hawkes, M., Opoka, R.O., Namasopo, S., Miller, C., Conroy, A.L., Serghides, L., Kim, H., Thampi, N., Liles,
 527 W.C., John, C.C., Kain, K.C., 2011. Nitric oxide for the adjunctive treatment of severe malaria: hypothesis
 528 and rationale. Med Hypotheses 77, 437-444.

529 Hendriksen, I.C., Mwanga-Amumpaire, J., von Seidlein, L., Mtove, G., White, L.J., Olaosebikan, R., Lee, S.J.,
 530 Tshefu, A.K., Woodrow, C., Amos, B., Karema, C., Saiwaew, S., Maitland, K., Gomes, E., Pan-Ngum, W.,
 531 Gesase, S., Silamut, K., Reyburn, H., Joseph, S., Chotivanich, K., Fanello, C.I., Day, N.P., White, N.J.,
 532 Dondorp, A.M., 2012. Diagnosing severe falciparum malaria in parasitaemic African children: a
 533 prospective evaluation of plasma PfHRP2 measurement. PLoS Med 9, e1001297.

534 Hoffmann, A., Pfeil, J., Alfonso, J., Kurz, F.T., Sahm, F., Heiland, S., Monyer, H., Bendszus, M., Mueller, A.K.,
 535 Helluy, X., Pham, M., 2016. Experimental Cerebral Malaria Spreads along the Rostral Migratory Stream.
 536 PLoS Pathog 12, e1005470.

537 Hommel, M., Elliott, S.R., Soma, V., Kelly, G., Fowkes, F.J., Chesson, J.M., Duffy, M.F., Bockhorst, J., Avril,
 538 M., Mueller, I., Raiko, A., Stanisic, D.I., Rogerson, S.J., Smith, J.D., Beeson, J.G., 2010. Evaluation of the
 539 antigenic diversity of placenta-binding *Plasmodium falciparum* variants and the antibody repertoire among
 540 pregnant women. Infect Immun 78, 1963-1978.

541 Hviid, L., Jensen, A.T., 2015. PfEMP1 - A Parasite Protein Family of Key Importance in *Plasmodium falciparum*
 542 Malaria Immunity and Pathogenesis. Adv Parasitol 88, 51-84.

543 Jespersen, J.S., Wang, C.W., Mkumbaye, S.I., Minja, D.T., Petersen, B., Turner, L., Petersen, J.E., Lusingu,
 544 J.P., Theander, T.G., Lavstsen, T., 2016. *Plasmodium falciparum* var genes expressed in children with
 545 severe malaria encode CIDRalpha1 domains. EMBO Mol Med 8, 839-850.

546 Josling, G.A., Petter, M., Oehring, S.C., Gupta, A.P., Dietz, O., Wilson, D.W., Schubert, T., Langst, G., Gilson,
 547 P.R., Crabb, B.S., Moes, S., Jenoe, P., Lim, S.W., Brown, G.V., Bozdech, Z., Voss, T.S., Duffy, M.F.,

2015. A *Plasmodium falciparum* Bromodomain Protein Regulates Invasion Gene Expression. Cell Host Microbe 17, 741-751.

Joyner, C., Barnwell, J.W., Galinski, M.R., 2015. No more monkeying around: primate malaria model systems are key to understanding *Plasmodium vivax* liver-stage biology, hypnozoites, and relapses. Front Microbiol 6, 145.

Kalanon, M., Bargieri, D., Sturm, A., Matthews, K., Ghosh, S., Goodman, C.D., Thiberge, S., Mollard, V., McFadden, G.I., Menard, R., de Koning-Ward, T.F., 2016. The *Plasmodium* translocon of exported proteins component EXP2 is critical for establishing a patent malaria infection in mice. Cell Microbiol 18, 399-412.

Karmodiya, K., Pradhan, S.J., Joshi, B., Jangid, R., Reddy, P.C., Galande, S., 2015. A comprehensive epigenome map of *Plasmodium falciparum* reveals unique mechanisms of transcriptional regulation and identifies H3K36me2 as a global mark of gene suppression. Epigenetics Chromatin 8, 32.

Lapp, S.A., Mok, S., Zhu, L., Wu, H., Preiser, P.R., Bozdech, Z., Galinski, M.R., 2015. *Plasmodium knowlesi* gene expression differs in ex vivo compared to in vitro blood-stage cultures. Malaria J 14, 110.

Maier, A.G., Cooke, B.M., Cowman, A.F., Tilley, L., 2009. Malaria parasite proteins that remodel the host erythrocyte. Nat Rev Microbiol 7, 341-354.

Manning, L., Laman, M., Davis, W.A., Davis, T.M., 2014. Clinical features and outcome in children with severe *Plasmodium falciparum* malaria: a meta-analysis. PloS One 9, e86737.

Maude, R.J., Barkhof, F., Hassan, M.U., Ghose, A., Hossain, A., Abul Faiz, M., Choudhury, E., Rashid, R., Abu Sayeed, A., Charunwatthana, P., Plewes, K., Kingston, H., Maude, R.R., Silamut, K., Day, N.P., White, N.J., Dondorp, A.M., 2014. Magnetic resonance imaging of the brain in adults with severe falciparum malaria. Malaria J 13, 177.

Mayor, A., Bardaji, A., Macete, E., Nhampossa, T., Fonseca, A.M., Gonzalez, R., Maculuve, S., Cistero, P., Ruperez, M., Campo, J., Vala, A., Sigauque, B., Jimenez, A., Machevo, S., de la Fuente, L., Nhama, A., Luis, L., Aponte, J.J., Acacio, S., Nhacolo, A., Chitnis, C., Dobano, C., Sevene, E., Alonso, P.L., Menendez, C., 2015. Changing Trends in *P. falciparum* Burden, Immunity, and Disease in Pregnancy. New Engl J Med 373, 1607-1617.

575 Mayor, A., Hafiz, A., Bassat, Q., Rovira-Vallbona, E., Sanz, S., Machevo, S., Aguilar, R., Cistero, P., Sigauque,
 576 B., Menendez, C., Alonso, P.L., Chitnis, C.E., 2011. Association of severe malaria outcomes with platelet-
 577 mediated clumping and adhesion to a novel host receptor. PloS One 6, e19422.

578 McDonald, C.R., Darling, A.M., Conroy, A.L., Tran, V., Cabrera, A., Liles, W.C., Wang, M., Aboud, S., Urassa,
 579 W., Fawzi, W.W., Kain, K.C., 2015a. Inflammatory and Angiogenic Factors at Mid-Pregnancy Are
 580 Associated with Spontaneous Preterm Birth in a Cohort of Tanzanian Women. PloS One 10, e0134619.

581 McDonald, C.R., Tran, V., Kain, K.C., 2015b. Complement Activation in Placental Malaria. Front Microbiol 6,
 582 1460.

583 Medana, I.M., Day, N.P., Sachanonta, N., Mai, N.T., Dondorp, A.M., Pongponratn, E., Hien, T.T., White, N.J.,
 584 Turner, G.D., 2011. Coma in fatal adult human malaria is not caused by cerebral oedema. Malaria J, 10,
 585 267.

586 Mohanty, S., Mishra, S.K., Patnaik, R., Dutt, A.K., Pradhan, S., Das, B., Patnaik, J., Mohanty, A.K., Lee, S.J.,
 587 Dondorp, A.M., 2011. Brain swelling and mannitol therapy in adult cerebral malaria: a randomized trial.
 588 Clin Infect Dis 53, 349-355.

589 Moxon, C.A., Chisala, N.V., Wassmer, S.C., Taylor, T.E., Seydel, K.B., Molyneux, M.E., Faragher, B.,
 590 Kennedy, N., Toh, C.H., Craig, A.G., Heyderman, R.S., 2014. Persistent endothelial activation and
 591 inflammation after *Plasmodium falciparum* Infection in Malawian children. J Infect Dis 209, 610-615.

592 Moxon, C.A., Wassmer, S.C., Milner, D.A., Jr., Chisala, N.V., Taylor, T.E., Seydel, K.B., Molyneux, M.E.,
 593 Faragher, B., Esmon, C.T., Downey, C., Toh, C.H., Craig, A.G., Heyderman, R.S., 2013. Loss of
 594 endothelial protein C receptors links coagulation and inflammation to parasite sequestration in cerebral
 595 malaria in African children. Blood 122, 842-851.

596 Nacer, A., Movila, A., Sohet, F., Girgis, N.M., Gundra, U.M., Loke, P., Daneman, R., Frevert, U., 2014.
 597 Experimental cerebral malaria pathogenesis--hemodynamics at the blood brain barrier. PLoS Pathog 10,
 598 e1004528.

599 O'Sullivan, J.M., Preston, R.J., O'Regan, N., O'Donnell, J.S., 2016. Emerging roles for hemostatic dysfunction
 600 in malaria pathogenesis. Blood 127, 2281-2288.

601 Padmanabhan, B., Mathur, S., Manjula, R., Tripathi, S., 2016. Bromodomain and extra-terminal (BET) family
 602 proteins: New therapeutic targets in major diseases. J Biosci 41, 295-311.

Pai, S., Qin, J., Cavanagh, L., Mitchell, A., El-Assaad, F., Jain, R., Combes, V., Hunt, N.H., Grau, G.E., Weninger, W., 2014. Real-time imaging reveals the dynamics of leukocyte behaviour during experimental cerebral malaria pathogenesis. *PLoS Pathog* 10, e1004236.

Pain, A., Ferguson, D.J., Kai, O., Urban, B.C., Lowe, B., Marsh, K., Roberts, D.J., 2001. Platelet-mediated clumping of *Plasmodium falciparum*-infected erythrocytes is a common adhesive phenotype and is associated with severe malaria. *Proc Natl Acad Sci (USA)* 98, 1805-1810.

Penet, M.F., Viola, A., Confort-Gouny, S., Le Fur, Y., Duhamel, G., Kober, F., Ibarrola, D., Izquierdo, M., Coltel, N., Gharib, B., Grau, G.E., Cozzone, P.J., 2005. Imaging experimental cerebral malaria in vivo: significant role of ischemic brain edema. *J Neurosci* 25, 7352-7358.

Petersen, J.E., Bouwens, E.A., Tamayo, I., Turner, L., Wang, C.W., Stins, M., Theander, T.G., Hermida, J., Mosnier, L.O., Lavstsen, T., 2015. Protein C system defects inflicted by the malaria parasite protein PfEMP1 can be overcome by a soluble EPCR variant. *Thromb Haemost* 114, 1038-1048.

Potchen, M.J., Kampondeni, S.D., Seydel, K.B., Birbeck, G.L., Hammond, C.A., Bradley, W.G., DeMarco, J.K., Glover, S.J., Ugorji, J.O., Latourette, M.T., Siebert, J.E., Molyneux, M.E., Taylor, T.E., 2012. Acute brain MRI findings in 120 Malawian children with cerebral malaria: new insights into an ancient disease. *AJNR Am J Neuroradiol* 33, 1740-1746.

Rogerson, S.J., Pollina, E., Getachew, A., Tadesse, E., Lema, V.M., Molyneux, M.E., 2003. Placental monocyte infiltrates in response to *Plasmodium falciparum* malaria infection and their association with adverse pregnancy outcomes. *American J Trop Med Hyg* 68, 115-119.

Rowe, A., Obeiro, J., Newbold, C.I., Marsh, K., 1995. *Plasmodium falciparum* rosetting is associated with malaria severity in Kenya. *Infect Immun* 63, 2323-2326.

Rowe, J.A., Claessens, A., Corrigan, R.A., Arman, M., 2009. Adhesion of *Plasmodium falciparum*-infected erythrocytes to human cells: molecular mechanisms and therapeutic implications. *Expert Rev Mol Med* 11, e16.

Rowe, J.A., Handel, I.G., Thera, M.A., Deans, A.M., Lyke, K.E., Kone, A., Diallo, D.A., Raza, A., Kai, O., Marsh, K., Plowe, C.V., Doumbo, O.K., Moulds, J.M., 2007. Blood group O protects against severe *Plasmodium falciparum* malaria through the mechanism of reduced rosetting. *Proc Natl Acad Sci (USA)* 104, 17471-17476.

631 Rowe, J.A., Shafi, J., Kai, O.K., Marsh, K., Raza, A., 2002. Nonimmune IgM, but not IgG binds to the surface of
 632 *Plasmodium falciparum*-infected erythrocytes and correlates with rosetting and severe malaria. Am J Trop
 633 Med Hyg 66, 692-699.

634 Sahu, P.K., Satpathi, S., Behera, P.K., Mishra, S.K., Mohanty, S., Wassmer, S.C., 2015. Pathogenesis of
 635 cerebral malaria: new diagnostic tools, biomarkers, and therapeutic approaches. Front Cell Infect Microbiol
 636 5, 75.

637 Salinas, J.L., Kissinger, J.C., Jones, D.P., Galinski, M.R., 2014. Metabolomics in the fight against malaria.
 638 Memorias do Instituto Oswaldo Cruz 109, 589-597.

639 Serghides, L., Kim, H., Lu, Z., Kain, D.C., Miller, C., Francis, R.C., Liles, W.C., Zapol, W.M., Kain, K.C., 2011.
 640 Inhaled nitric oxide reduces endothelial activation and parasite accumulation in the brain, and enhances
 641 survival in experimental cerebral malaria. PLoS One 6, e27714.

642 Seydel, K.B., Kampondeni, S.D., Valim, C., Potchen, M.J., Milner, D.A., Muwalo, F.W., Birbeck, G.L., Bradley,
 643 W.G., Fox, L.L., Glover, S.J., Hammond, C.A., Heyderman, R.S., Chilingulo, C.A., Molyneux, M.E., Taylor,
 644 T.E., 2015. Brain swelling and death in children with cerebral malaria. New Engl J Med 372, 1126-1137.

645 Souza, R.M., Ataide, R., Dombrowski, J.G., Ippolito, V., Aitken, E.H., Valle, S.N., Alvarez, J.M., Epiphanyo, S.,
 646 Marinho, C.R., 2013. Placental histopathological changes associated with *Plasmodium vivax* infection
 647 during pregnancy. PLoS Negl Trop Dis 7, e2071.

648 Stevenson, L., Laursen, E., Cowan, G.J., Bandoh, B., Barfod, L., Cavanagh, D.R., Andersen, G.R., Hviid, L.,
 649 2015. alpha2-Macroglobulin Can Crosslink Multiple *Plasmodium falciparum* Erythrocyte Membrane Protein
 650 1 (PfEMP1) Molecules and May Facilitate Adhesion of Parasitized Erythrocytes. PLoS Pathog 11,
 651 e1005022.

652 Storm, J., Craig, A.G., 2014. Pathogenesis of cerebral malaria-inflammation and cytoadherence. Front Cell
 653 Infect Microbiol 4, 100.

654 Taylor, T.E., Fu, W.J., Carr, R.A., Whitten, R.O., Mueller, J.S., Fosiko, N.G., Lewallen, S., Liomba, N.G.,
 655 Molyneux, M.E., 2004. Differentiating the pathologies of cerebral malaria by postmortem parasite counts.
 656 Nat Med, 10, 143-145.

657 Turner, L., Lavstsen, T., Berger, S.S., Wang, C.W., Petersen, J.E., Avril, M., Brazier, A.J., Freeth, J.,
 658 Jespersen, J.S., Nielsen, M.A., Magistrado, P., Lusingu, J., Smith, J.D., Higgins, M.K., Theander, T.G.,

2013. Severe malaria is associated with parasite binding to endothelial protein C receptor. *Nature* 498, 502-505.

Warrell, D.A., Looareesuwan, S., Phillips, R.E., White, N.J., Warrell, M.J., Chapel, H.M., Areekul, S., Tharavanij, S., 1986. Function of the blood-cerebrospinal fluid barrier in human cerebral malaria: rejection of the permeability hypothesis. *Am J Trop Med Hyg* 35, 882-889.

Wassmer, S.C., Combes, V., Grau, G.E., 2011a. Platelets and microparticles in cerebral malaria: the unusual suspects. *Drug Discov Today Dis Mech* 8, e15-e23.

Wassmer, S.C., Moxon, C.A., Taylor, T., Grau, G.E., Molyneux, M.E., Craig, A.G., 2011b. Vascular endothelial cells cultured from patients with cerebral or uncomplicated malaria exhibit differential reactivity to TNF. *Cell Microbiol* 13, 198-209.

Wassmer, S.C., Taylor, T., MacLennan, C.A., Kanjala, M., Mukaka, M., Molyneux, M.E., Grau, G.E., 2008. Platelet-induced clumping of *Plasmodium falciparum*-infected erythrocytes from Malawian patients with cerebral malaria-possible modulation in vivo by thrombocytopenia. *J Infect Dis* 197, 72-78.

Wassmer, S.C., Taylor, T.E., Rathod, P.K., Mishra, S.K., Mohanty, S., Arevalo-Herrera, M., Duraisingh, M.T., Smith, J.D., 2015. Investigating the Pathogenesis of Severe Malaria: A Multidisciplinary and Cross-Geographical Approach. *Am J Trop Med Hyg* 93, 42-56.

White, N.J., Turner, G.D., Day, N.P., Dondorp, A.M., 2013. Lethal malaria: Marchiafava and Bignami were right. *J Infect Dis* 208, 192-198.

Yeo, T.W., Lampah, D.A., Gitawati, R., Tjitra, E., Kenangalem, E., McNeil, Y.R., Darcy, C.J., Granger, D.L., Weinberg, J.B., Lopansri, B.K., Price, R.N., Duffull, S.B., Celermajer, D.S., Anstey, N.M., 2007. Impaired nitric oxide bioavailability and L-arginine reversible endothelial dysfunction in adults with falciparum malaria. *J Exp Med* 204, 2693-2704.

Yeo, T.W., Lampah, D.A., Kenangalem, E., Tjitra, E., Weinberg, J.B., Granger, D.L., Price, R.N., Anstey, N.M., 2014. Decreased endothelial nitric oxide bioavailability, impaired microvascular function, and increased tissue oxygen consumption in children with falciparum malaria. *J Infect Dis* 210, 1627-1632.

Yeo, T.W., Lampah, D.A., Rooslamati, I., Gitawati, R., Tjitra, E., Kenangalem, E., Price, R.N., Duffull, S.B., Anstey, N.M., 2013. A randomized pilot study of L-arginine infusion in severe falciparum malaria: preliminary safety, efficacy and pharmacokinetics. *PLoS One* 8, e69587.