

LONDON
SCHOOL of
HYGIENE
& TROPICAL
MEDICINE



Boss, JT; Li, Y; Atherton, TG; Walker, S; Williamson, SM; Rogers, J; Chaudhuri, RR; Weinert, LA; Holden, MT; Maskell, DJ; Tucker, AW; Wren, BW; Rycroft, AN; Langford, PR (2015) Characterisation of a mobilisable plasmid conferring florfenicol and chloramphenicol resistance in *Actinobacillus pleuropneumoniae*. *Veterinary microbiology*, 178 (3-4). pp. 279-82. ISSN 0378-1135 DOI: 10.1016/j.vetmic.2015.05.020

Downloaded from: <http://researchonline.lshtm.ac.uk/2210791/>

DOI: [10.1016/j.vetmic.2015.05.020](https://doi.org/10.1016/j.vetmic.2015.05.020)

Usage Guidelines

Please refer to usage guidelines at <http://researchonline.lshtm.ac.uk/policies.html> or alternatively contact researchonline@lshtm.ac.uk.

Available under license: <http://creativecommons.org/licenses/by/2.5/>



Short communication

Characterisation of a mobilisable plasmid conferring florfenicol and chloramphenicol resistance in *Actinobacillus pleuropneumoniae*



Janine T Bossé^{a,*}, Yanwen Li^{a,1}, Tom G Atherton^a, Stephanie Walker^a,
 Susanna M Williamson^b, Jon Rogers^b, Roy R Chaudhuri^{c,2}, Lucy A Weinert^c,
 Matthew TG Holden^{d,3}, Duncan J Maskell^c, Alexander W Tucker^c, Brendan W Wren^e,
 Andrew N Rycroft^f, Paul R Langford^a

^a Section of Paediatrics, Department of Medicine, Imperial College London, St. Mary's Campus, London, W2 1PG, UK

^b Animal and Plant Health Agency (APHA) Bury St Edmunds, Rougham Hill, Bury St Edmunds, Suffolk, IP33 2RX, UK

^c Department of Veterinary Medicine, University of Cambridge, Madingley Road, Cambridge, CB3 0ES, UK

^d The Wellcome Trust Sanger Institute, Wellcome Trust Genome Campus, Hinxton, Cambridge CB10 1SA, UK

^e Faculty of Infectious & Tropical Diseases, London School of Hygiene & Tropical Medicine, Keppel Street, London, WC1E 7HT, UK

^f Department of Pathology and Pathogen Biology, The Royal Veterinary College, Hawkshead Campus, Hatfield, Hertfordshire, AL9 7TA, UK

ARTICLE INFO

Article history:

Received 29 April 2015

Received in revised form 22 May 2015

Accepted 25 May 2015

ABSTRACT

The complete nucleotide sequence of a 7.7 kb mobilisable plasmid (pM3446F), isolated from a florfenicol resistant isolate of *Actinobacillus pleuropneumoniae*, showed extended similarity to plasmids found in other members of the *Pasteurellaceae* containing the *floR* gene as well as replication and mobilisation genes. Mobilisation into other *Pasteurellaceae* species confirmed that this plasmid can be transferred horizontally.

©2015 The Authors. Published by Elsevier B.V. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Actinobacillus pleuropneumoniae, one of the main pathogens contributing to swine respiratory disease throughout the world, has been used as an indicator organism in surveillance studies of antimicrobial resistance in bacteria of animal origin (de Jong et al., 2014; Hendriksen et al., 2008). Tetracyclines, beta-lactams, and trimethoprim/sulphonamides are the commonest frontline treatments for porcine pleuropneumonia in Europe (European Medicines Agency, 2012), however, levels of resistance are increasing (de Jong et al., 2014; Vanni et al., 2012). To a lesser extent, florfenicol, a fluorinated thiamphenicol derivative licensed to treat respiratory diseases of pigs and cattle in Europe since 2000 and 1995, respectively (Kehrenberg et al., 2004), has been used. Most surveys of *A. pleuropneumoniae* antimicrobial susceptibility have shown that nearly all isolates are susceptible to florfenicol, except in Korea where resistance levels of 34% were recently reported

(Gutiérrez-Martín et al., 2006; Kucerova et al., 2011; Priebe and Schwarz, 2003; Shin et al., 2005; Vanni et al., 2012; Yoo et al., 2014).

A chloramphenicol/florfenicol efflux pump, encoded by *floR* (Schwarz et al., 2004), has been detected by PCR in florfenicol-resistant *A. pleuropneumoniae* isolates (Kucerova et al., 2011; Yoo et al., 2014), and this resistance was transferrable to *Escherichia coli* (Yoo et al., 2014), however no plasmid was characterised.

Here we describe the isolation and characterisation of a 7.7 kb florfenicol resistance plasmid from a clinical isolate of *A. pleuropneumoniae*. Comparative sequence analysis of this plasmid with others reported as mediating florfenicol resistance in bovine isolates of *Pasteurella multocida* (Kehrenberg and Schwarz, 2005; Kehrenberg et al., 2008) and *Mannheimia haemolytica* (Katsuda et al., 2012) is also described.

1.1. Material and methods

1.1.1. Bacterial isolates and MIC measurements

A. pleuropneumoniae MIDG3446, a serovar 2 clinical isolate, was originally cultured from pneumonic lungs of a pig in Greece in 2010 and archived at the then Animal Health and Veterinary Laboratory Agency (now Animal and Plant Health Agency) in England. Minimum inhibitory concentrations (MIC) for florfenicol and chloramphenicol were determined by agar dilution using Chocolate Mueller-Hinton plates, with *A. pleuropneumoniae* ATCC

* Corresponding author. Tel.: +44 0 20 759 41803; fax: +44 0 20 759 4398.

E-mail addresses: j.bosse@imperial.ac.uk (J.T. Bossé), p.langford@imperial.ac.uk (P.R. Langford).

¹ These authors contributed equally to this work.

² Present address: Department of Molecular Biology and Biotechnology, University of Sheffield, Firth Court, Western Bank, Sheffield, S10 2TN, UK.

³ Present address: School of Medicine, University of St Andrews, St Andrews, UK.

27090, *Histophilus somni* ATCC 70025 and *Staphylococcus aureus* ATCC 29213 used as quality control strains (CLSI, 2008).

1.1.2. Plasmid isolation and confirmation of *floR* by PCR

Plasmid DNA was isolated using a Qiaprep Spin Miniprep kit (Qiagen) according to the manufacturer's protocol. The presence of the *floR* gene was confirmed by PCR using primers *floR*_for (CGACGCCGCTATGATCCAATC) and *floR*_rev (CCCAAAAAGCCG-GACTCGGAAG). Initially, florfenicol resistance was transferred to *E. coli* Stellar cells (Clontech) by heat shock with the MIDG3446 plasmid extract, according to the manufacturer's protocol, with selection of transformants on LB agar supplemented with 10 µg/ml florfenicol. Subsequently, mobilisation into selected *Pasteurellaceae* strains was assessed using a mating protocol previously described (Bossé et al., 2009; Bossé et al., 2015). Recipient strains included nalidixic acid resistant derivatives of *M. haemolytica* (MIDG1579Nal^R), *P. multocida* (MIDG1570 Nal^R), and *Haemophilus parasuis* (MIDG3176Nal^R), as well as the NAD-independent *A. pleuropneumoniae* isolate, MIDG2331Δ*ureC*::*nadV*, previously described (Bossé et al., 2009; Bossé et al., 2014). Transconjugants were selected on Brain Heart Infusion agar (with or without 0.01% NAD or 20 µg/ml nalidixic acid, as appropriate) supplemented with 2 µg/ml florfenicol. Transformants and transconjugants were tested by PCR for the presence of *floR*, as above, and for the *nadV* gene, as previously described (Bossé et al., 2015), where appropriate. MICs for florfenicol and chloramphenicol were determined, as above, for recipient strains +/- plasmid.

1.2. Plasmid sequence

A plasmid carrying the *floR* gene was isolated from a selected *E. coli* transformant (above) and the complete nucleotide sequence was determined using a primer walking strategy. Sequence analysis was carried out using BLASTn and BLASTx. Alignments with other *floR* plasmids were done using ClustalW. The sequence of the *A. pleuropneumoniae floR* plasmid (pM3446F) has been deposited to Genbank (accession number KP696484).

2. Results and discussion

A serovar 2 isolate of *A. pleuropneumoniae*, MIDG3446, was found to be resistant to florfenicol and chloramphenicol (Table 1). Analysis of DNA from MIDG3446 revealed multiple small plasmids, one of which when transformed into *E. coli* Stellar cells (Clontech), conferred resistance to florfenicol and chloramphenicol (Fig. 1; Table 1). Additionally, mobilisation from MIDG3446 was successful into *A. pleuropneumoniae* MIDG2331Δ*ureC*::*nadV* and *M. haemolytica* MIDG1579Nal^R, but not *P. multocida* MIDG1570Nal^R nor *H. parasuis* MIDG3176Nal^R. The location of the genes encoding the conjugation machinery was not determined, and no attempts were

made to optimise mating conditions, nor were other isolates of these species tested. PCR confirmed the presence of *floR* in transformants and transconjugants (Fig. 1), and MICs for florfenicol and chloramphenicol, determined as above, for recipient strains +/- plasmid indicated transfer of resistance with the plasmid (Table 1).

The complete 7,709-bp sequence of pM3446F, isolated from a selected *E. coli* transformant, was determined using a primer walking strategy. Sequence analysis using BLASTn and BLASTx (<http://blast.ncbi.nlm.nih.gov/Blast.cgi>) revealed extended similarity to pMh1405 (AB621552), a 7,674-bp plasmid isolated in 2009 from an *M. haemolytica* strain recovered from a calf in Japan (Katsuda et al., 2012), and to a lesser extent to pCCK381 (AJ871969), a 10,874-bp plasmid isolated in 2005 from a *P. multocida* strain recovered from a calf in Thirsk, England (Kehrenberg and Schwarz, 2005), and subsequently from bovine and porcine isolates of *P. multocida* in Germany (Kehrenberg et al., 2008). All three *Pasteurellaceae*-derived plasmids (Fig. 2) have virtually identical replication (*repA-C*) and mobilisation genes (*mobA-C*), as well as *oriV* and *oriT* sequences, as those comprising pDN1 (Fig. 2), a 5,112-bp RSF1010-like IncQ broad-host-range plasmid (Y19120) isolated from *Dichelobacter nodosus*, the causative agent of ovine footrot (Kehrenberg and Schwarz, 2005; Rawlings and Tietze, 2001; Whittle et al., 2000). However, whereas pDN1 does not encode resistance genes, these *Pasteurellaceae* plasmids contain differing fragments of what was initially described as Tn*floR*, a transposable element encoding *floR*, a *lysR* transcriptional regulator, and the *tnpA* transposase (Doublet et al., 2005), and subsequently shown to be an ISCR2-*floR* element (Toleman et al., 2006). Similar sequences have been found in various plasmids and chromosomes of different bacterial species, suggesting recombination and/or deletion events following integration have given rise to different truncated forms of the genes flanking *floR* (Doublet et al., 2005; Schwarz et al., 2004; Toleman et al., 2006).

The sequences flanking *floR* in these *Pasteurellaceae* plasmids show greatest similarity to those found in pAQU1 (AB571865), a 200 kb conjugative plasmid isolated from *Photobacterium damsela*, subsp. *damsela* in Japan (Nonaka et al., 2012). In Figure 2, only the sequence around the ISCR2-*floR* element of pAQU1 is shown (from bases 138014 to 143017 in the Genbank annotated sequence), with the Δ*tnp* gene upstream of *floR* almost identical to the last 720 bases of the *tnp* gene downstream. In pCCK381, in addition to the partial ISCR2-*floR* (99% identity with pAQU1 bases 138160–141381), the 2062 bases comprising the end of the *lysR* gene to the start of the *oriV* are 99% identical to bases 146509–148591 of pAQU1. Whereas, upstream of the Δ*tnp* gene in pCCK381, there are sequences 99–100% identity with other regions (bases 148616–148778 and bases 148992–149190) of pAQU1. In pMh1405, the 2.5 kb upstream of *oriV* shares 99% identity with

Table 1
Antimicrobial susceptibility for strains with and without plasmids.

Strain	Species	MIC (µg/ml)		Source or Reference
		Florfenicol	Chloramphenicol	
MIDG3446	Ap ^a	8	8	This study
MIDG2331Δ <i>ureC</i> :: <i>nadV</i>	Ap	2	2	(Bossé et al., 2014)
MIDG2331Δ <i>ureC</i> :: <i>nadV</i> + pM3446F	Ap	8	8	This study
MIDG1579	Mh ^b	1	1	(Bossé et al., 2009)
MIDG1579 + pM3446F	Mh	16	16	This study
Stellar	Ec ^c	2	4	Clontech
Stellar + pM3446F	Ec	16	32	This study

^a Ap = *A. pleuropneumoniae*.

^b Mh = *M. haemolytica*.

^c Ec = *E. coli*.

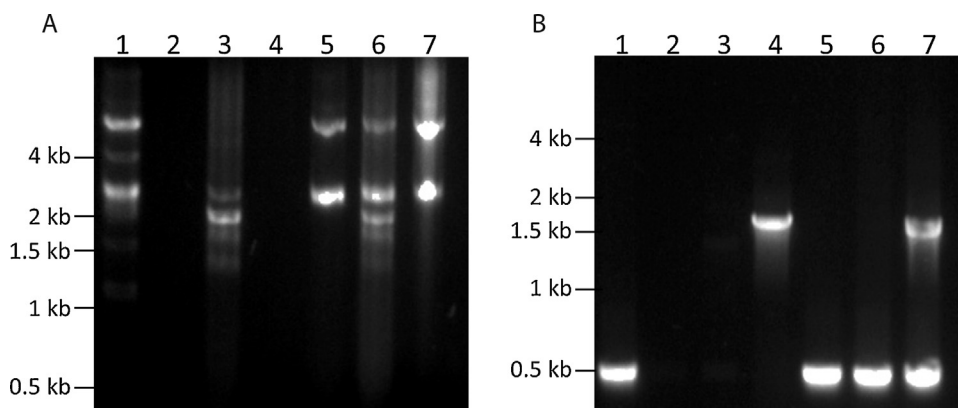


Fig. 1. Transfer of florfenicol resistance plasmid from *A. pleuropneumoniae* MIDG3446 by transformation into *E. coli* Stellar, or mating into *M. heamolytica* MIDG1579 and *A. pleuropneumoniae* MIDG2331 $\Delta ureC::nadV$. (A) Comparison of plasmid extracts from donor strain MIDG3446 (lane 1), recipient strains *E. coli* Stellar, *M. heamolytica* MIDG1579 and *A. pleuropneumoniae* MIDG2331 $\Delta ureC::nadV$ (lanes 2–4), and respective florfenicol resistant transformant/transconjugants showing transfer of plasmid pM3446F into *E. coli* Stellar, *M. heamolytica* MIDG1579, and *A. pleuropneumoniae* MIDG2331 $\Delta ureC::nadV$ (lanes 5–7) (B) Multiplex PCR amplification of *floR* (510-bp amplicon) and *nadV* (1.5 kb amplicon) from *A. pleuropneumoniae* MIDG3446 (lane 1), *E. coli* Stellar (lane 2), *M. heamolytica* MIDG1579 (lane 3), *A. pleuropneumoniae* MIDG2331 $\Delta ureC::nadV$ (lane 4), *E. coli* Stellar + pM3446F (lane 5), *M. heamolytica* MIDG1579 + pM3446F (lane 6), *A. pleuropneumoniae* MIDG2331 $\Delta ureC::nadV$ + pM3446F (lane 7).

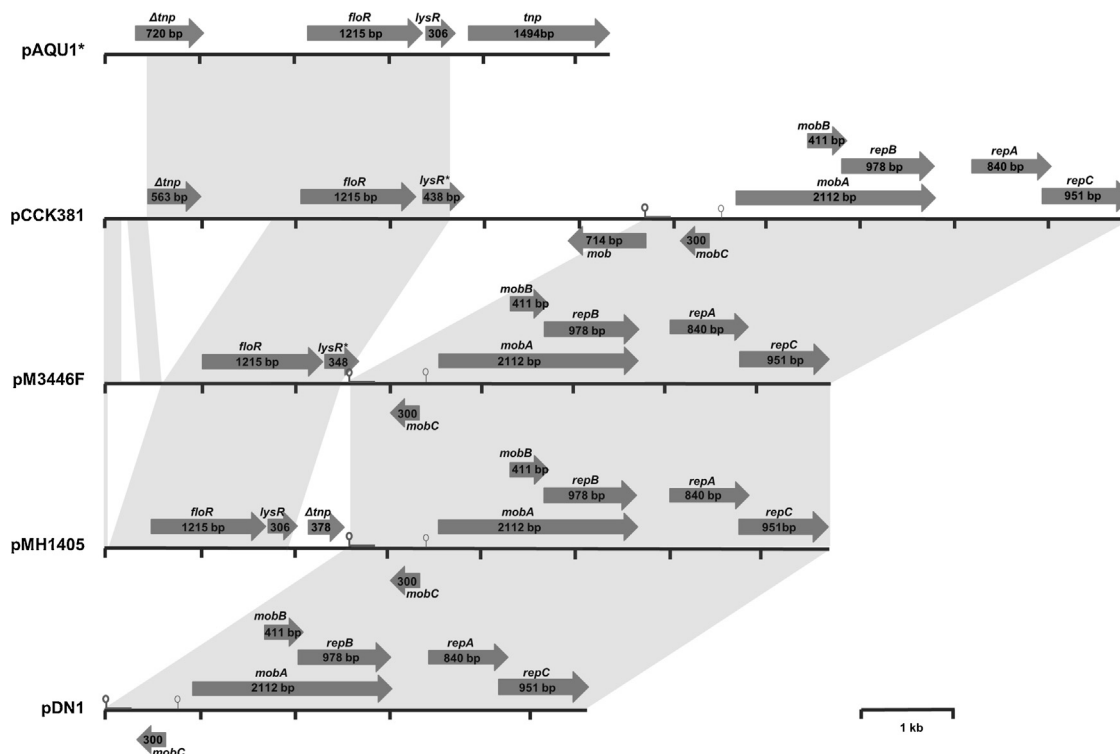


Fig. 2. Schematic comparison of florfenicol resistance plasmid pM3446F from *A. pleuropneumoniae* with plasmids pAQU1 (from *Photobacterium damsela*, subsp. *damsela*), pCCK381 and pMH1405 (from related *Pasteurellaceae*), and pDN1 (from *Dichelobacter nodosus*). Note: only 5.3 kb (from bases 138014–143017 of 204052 total) of pAQU1 is shown. Reading frames are indicated by arrows, with arrowheads showing direction of transcription (*floR*: florfenicol resistance; *lysR*: transcriptional regulator; *lysR**: partial *lysR*; *tnp*: transposase; Δtnp : truncated transposase; *mob*, *mobA*, *mobB*, *mobC*: plasmid mobilisation; *repA*, *repB*, *repC*: plasmid replication). The predicted origins of replication (*oriV*; \circ) and transfer (*oriT*; \circ) are shown. Grey blocks between sequences indicate $\geq 98\%$ nucleotide sequence identity. A distance scale in kb is shown.

bases 139426–141904 of pAQU1 (a different deletion fragment of ISCR2-*floR* than that seen in pCCK381); and in pM3446F no *tnp* sequences remain (with bases 622–2522 of pM3446F being 99% identical to bases 139426–141904 of pAQU1). An orf downstream of *floR* in pM3446F shows partial identity with *lysR*, with the first 223/348 bp being 100% identical to bases 141107–141329, and bases 214–334 of the orf having 98% identity with bases 148514–148633, of pAQU1. In addition, bases 25–597 of pM3446F are 99%

identical to bases 148616–149190 of pAQU1, parts of which are conserved in the sequence upstream of the Δtnp gene in pCCK381. These data suggest that these *Pasteurellaceae* plasmids share a common origin, with sequences from pAQU1 (or a related plasmid) having integrated into pDN1, and subsequent deletions/rearrangements of the inserted sequences.

Two further *Pasteurellaceae floR* plasmids have been identified: pCCK1900 (NC_011378), a 10,226-bp plasmid from a porcine

isolate of *P. multocida* in Germany (Kehrenberg et al., 2008) that encodes resistance to florfenicol (*floR*), sulphonamide (*sul2*), and streptomycin (*strA*, *strB*); pCCK13698 (NC_007800), a 14,969-bp plasmid from a bovine isolate of *Bibersteinia trehalosi* in France (Kehrenberg et al., 2006) that encodes resistance to florfenicol (*floR*), chloramphenicol (*catA3*) and sulphonamide (*sul2*). The sequence of pCCK1900 shows simple integration of the *floR* and *lysR* genes into an RSF1010 backbone (Kehrenberg et al., 2008). Although pCCK13698 shares some *rep* and *mob* gene sequences, as well as the *sul2* gene, with RSF1010, it shows more extensive rearrangements with sequences from other plasmids including pHS-rec from *H. parasuis* (Lancashire et al., 2005) and pMVSCS1 from *Mannheimia varigena* (Kehrenberg and Schwarz, 2002), and the *floR* and *lysR* genes from ISCR2-*floR* (Kehrenberg et al., 2006). Both pCCK1900 and pCCK13698 appear to have arisen separately from the pDN1-like plasmids described above.

3. Conclusion

In summary, to our knowledge, this is the first report of a complete sequence of a mobilisable florfenicol resistance plasmid from *A. pleuropneumoniae*. Structural analysis of pM3446F revealed extensive similarity to two florfenicol resistance plasmids found in other members of the *Pasteurellaceae*, and mating experiments confirmed the ability to mobilise between species. This highlights the importance of continued surveillance of florfenicol susceptibility in *A. pleuropneumoniae* and other Gram-negative pathogens that may co-exist within the respiratory tract of pigs.

3.1. Funding

This work was supported by a Longer and Larger (LoLa) grant from the Biotechnology and Biological Sciences Research Council (grant numbers BB/G020744/1, BB/G019177/1, BB/G019274/1 and BB/G018553/1), the UK Department for Environment, Food and Rural Affairs, and Zoetis (formerly Pfizer Animal Health) awarded to the Bacterial Respiratory Diseases of Pigs-1 Technology (BRaDP1T) consortium. MTGH was supported by Wellcome Trust grant 98051. The MIC work was funded from the former AHVLA's Research and Development Internal Investment Fund RD0030c. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Acknowledgements

The BRaDP1T Consortium comprises: Duncan J. Maskell, Alexander W. (Dan) Tucker, Sarah E. Peters, Lucy A. Weinert, Jinhong (Tracy) Wang, Shi-Lu Luan, Roy R. Chaudhuri (University of Cambridge; present address for R. Chaudhuri is: Department of Molecular Biology and Biotechnology, University of Sheffield, Firth Court, Western Bank, Sheffield, S10 2TN, UK), Andrew N. Rycroft, Gareth A. Maglennon, Dominic Matthews (Royal Veterinary College); Brendan W. Wren, Jon Cuccui, Vanessa S. Terra (London School of Hygiene and Tropical Medicine); and Paul R. Langford, Janine T. Bossé, Yanwen Li (Imperial College London). We wish to thank Susanna Williamson and Chris Teale from the APHA for their advice and input.

References

Bossé, J.T., Durham, A.L., Rycroft, A.N., Kroll, J.S., Langford, P.R., 2009. New plasmid tools for genetic analysis of *Actinobacillus pleuropneumoniae* and other *Pasteurellaceae*. *Appl. Environ. Microbiol.* 75, 6124–6131. doi:http://dx.doi.org/10.1128/AEM.00809-09.

Bossé, J.T., Soares-Bazzolli, D.M., Li, Y., Wren, B.W., Tucker, A.W., Maskell, D.J., Rycroft, A.N., Langford, P.R., BRaDP1T consortium, 2014. The generation of successive unmarked mutations and chromosomal insertion of heterologous genes in *Actinobacillus pleuropneumoniae* using natural transformation. *PLoS ONE* 9, e111252. doi:http://dx.doi.org/10.1371/journal.pone.0111252.

Bossé, J.T., Li, Y., Walker, S., Atherton, T., Fernandez Crespo, R., Williamson, S.M., Rogers, J., Chaudhuri, R.R., Weinert, L.A., Oshota, O., Holden, M.T.G., Maskell, D.J., Tucker, A.W., Wren, B.W., Rycroft, A.N., Langford, P.R., 2015. BRaDP1T consortium, 2015. Identification of *dfiA 14* in two distinct plasmids conferring trimethoprim resistance in *Actinobacillus pleuropneumoniae*. *J. Antimicrob. Chemother.* doi:http://dx.doi.org/10.1093/jac/dkv121.

de Jong, A., Thomas, V., Simjee, S., Moyaert, H., Garch, El, F., Maher, K., Morrissey, L., Butty, P., Klein, U., Marion, H., Rigaut, D., Vallé, M., 2014. Antimicrobial susceptibility monitoring of respiratory tract pathogens isolated from diseased cattle and pigs across Europe: the vetPath study. *Vet. Microbiol.* 172, 202–215.

Clinical Laboratory Standards Institute, C.L.S.I., 2008. Performance Standards for Antimicrobial Disk and Dilution Susceptibility Tests for Bacteria Isolated From Animals - Third Edition: Approved Standard M31-A3. CLSI, Wayne, PA.

Doublet, B., Schwarz, S., Kehrenberg, C., Cloeckaert, A., 2005. Florfenicol resistance gene *floR* is part of a novel transposon. *Antimicrob. Agents Chemother.* 49, 2106–2108.

European Medicines Agency, 2012. Sales of veterinary antimicrobial agents in 19 EU/EEA countries in 2010. (EMA/88728/2012).

Gutiérrez-Martín, C.B., del Blanco, N.G., Blanco, M., Navas, J., Rodríguez-Ferri, E.F., 2006. Changes in antimicrobial susceptibility of *Actinobacillus pleuropneumoniae* isolated from pigs in Spain during the last decade. *Vet. Microbiol.* 115, 218–222.

Hendriksen, R.S., Mevius, D.J., Schroeter, A., Teale, C., Jouy, E., Butaye, P., Franco, A., Utinane, A., Amado, A., Moreno, M., Greko, C., Stärk, K.D., Berghold, C., Myllyniemi, A.-L., Hoszowski, A., Sunde, M., Aarestrup, F.M., 2008. Occurrence of antimicrobial resistance among bacterial pathogens and indicator bacteria in pigs in different European countries from year 2002–2004: the ARBAO-II study. *Acta. Vet. Scand.* 50, 19.

Katsuda, K., Kohmoto, M., Mikami, O., Tamamura, Y., Uchida, I., 2012. Plasmid-mediated florfenicol resistance in *Mannheimia haemolytica* isolated from cattle. *Vet. Microbiol.* 155, 444–447.

Kehrenberg, C., Schwarz, S., 2005. Plasmid-borne florfenicol resistance in *Pasteurella multocida*. *J. Antimicrob. Chemother.* 55, 773–775.

Kehrenberg, C., Meunier, D., Targant, H., Cloeckaert, A., Schwarz, S., Madec, J.-Y., 2006. Plasmid-mediated florfenicol resistance in *Pasteurella trehalosi*. *J. Antimicrob. Chemother.* 58, 13–17.

Kehrenberg, C., Mumme, J., Wallmann, J., Verspohl, J., Tegeler, R., Kühn, T., Schwarz, S., 2004. Monitoring of florfenicol susceptibility among bovine and porcine respiratory tract pathogens collected in Germany during the years 2002 and 2003. *J. Antimicrob. Chemother.* 54, 572–574.

Kehrenberg, C., Schwarz, S., 2002. Nucleotide sequence and organization of plasmid pMVSCS1 from *Mannheimia varigena*: identification of a multiresistance gene cluster. *J. Antimicrob. Chemother.* 49, 383–386.

Kehrenberg, C., Wallmann, J., Schwarz, S., 2008. Molecular analysis of florfenicol-resistant *Pasteurella multocida* isolates in Germany. *J. Antimicrob. Chemother.* 62, 951–955. doi:http://dx.doi.org/10.1093/jac/dkn359.

Kucerova, Z., Hradecka, H., Nechvatalova, K., Nedbalcova, K., 2011. Antimicrobial susceptibility of *Actinobacillus pleuropneumoniae* isolates from clinical outbreaks of porcine respiratory diseases. *Vet. Microbiol.* 150, 203–206.

Lancashire, J.F., Terry, T.D., Blackall, P.J., Jennings, M.P., 2005. Plasmid-encoded Tet B tetracycline resistance in *Haemophilus parasuis*. *Antimicrob. Agents Chemother.* 49, 1927–1931.

Nonaka, L., Maruyama, F., Miyamoto, M., Miyakoshi, M., Kurokawa, K., Masuda, M., 2012. Novel conjugative transferable multiple drug resistance plasmid pAQU1 from *Photobacterium damsela* subsp. *damsela* isolated from marine aquaculture environment. *Microbes Environ.* 27, 263–272.

Priebe, S., Schwarz, S., 2003. *In vitro* activities of florfenicol against bovine and porcine respiratory tract pathogens. *Antimicrob. Agents Chemother.* 47, 2703–2705.

Rawlings, D.E., Tietze, E., 2001. Comparative biology of IncQ and IncQ-like plasmids. *Microbiol. Mol. Biol. Rev.* 65, 481–496.

Schwarz, S., Kehrenberg, C., Doublet, B., Cloeckaert, A., 2004. Molecular basis of bacterial resistance to chloramphenicol and florfenicol. *FEMS Microbiol. Rev.* 28, 519–542.

Shin, S.J., Kang, S.G., Nabin, Kang, R., Yoo, M.L., HS, 2005. Evaluation of the antimicrobial activity of florfenicol against bacteria isolated from bovine and porcine respiratory disease. *Vet. Microbiol.* 106, 73–77.

Toleman, M.A., Bennet, P.M., Walsh, T.R., 2006. ISCR Elements: novel gene-capturing systems of the 21st century. *Microbiol. Mol. Biol. Rev.* 70, 296–316.

Vanni, M., Merenda, M., Barigazzi, G., Garbarino, C., Luppi, A., Tognetti, R., Intorre, L., 2012. Antimicrobial resistance of *Actinobacillus pleuropneumoniae* isolated from swine. *Vet. Microbiol.* 156, 172–177.

Whittle, G., Katz, M.E., Clayton, E.H., Cheetham, B.F., 2000. Identification and characterization of a native *Dichelobacter nodosus* plasmid, pDN1. *Plasmid* 43, 230–234.

Yoo, A.N., Cha, S.B., Shin, M.K., Won, H.K., Kim, E.H., Choi, H.W., Yoo, H.S., 2014. Serotypes and antimicrobial resistance patterns of the recent Korean *Actinobacillus pleuropneumoniae* isolates. *Vet. Rec.* 174, 223.