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Impact of Mass Azithromycin Distribution on Malaria Parasitemia during the Low-Transmission Season in Niger: A Cluster-Randomized Trial


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Abstract. We assessed the effect of mass azithromycin treatment on malaria parasitemia in a trachoma trial in Niger. Twenty-four study communities received treatment during the wet, high-transmission season. Twelve of the 24 communities were randomized to receive an additional treatment during the dry, low-transmission season. Outcome measurements were conducted at the community-level in children <1–72 months of age in May–June 2011. Parasitemia was higher in the 12 once-treated communities (29.8%, 95% confidence interval [CI] = 21.5–40.0%) than in the 12 twice-treated communities (19.5%, 95% CI = 13.0–26.5%, P = 0.03). Parasite density was higher in once-treated communities (354 parasites/μL, 95% CI = 117–528 parasites/μL) than in twice-treated communities (74 parasites/μL, 95% CI = 41–202 parasites/μL, P = 0.03). Mass distribution of azithromycin reduced malaria parasitemia 4–5 months after the intervention. The results suggest that drugs with antimalaria activity can have long-lasting impacts on malaria during periods of low transmission.

INTRODUCTION

The World Health Organization (WHO) currently recommends repeated mass oral azithromycin distributions in trachoma-endemic areas to control the ocular strains of Chlamydia that cause the disease. A cohort study and a cluster-randomized trial have showed that these distributions could reduce childhood mortality.1,2 Cohort studies have suggested that azithromycin may reduce leading causes of childhood mortality, including respiratory disease, diarrhea, and malaria.3–7 In areas with seasonal malaria transmission, repeated administration of antimalarial drugs to children during the transmission season (intermittent preventive therapy of children [IPTc])8 has been shown to reduce malaria transmission, although preventive therapy outside of the transmission season is not typically recommended.3 However, transmission models suggest treatment during the low-transmission season of an infectious disease may provide significant or even maximum impact.10–12 Azithromycin has antimalaria activity, although lower than first-line agents.13–15 It is not clear whether mass administration of azithromycin during the low malaria transmission season could have a long-lasting antimalaria effect.

In a large, cluster-randomized clinical trial for trachoma, we assessed whether mass oral azithromycin distributions reduced malaria parasitemia and increased hemoglobin concentration.16 We compared community-level malaria asexual parasitemia, parasite density and gametocytemia, and the hemoglobin concentration in communities randomized to a single mass treatment with azithromycin at the beginning of the rainy season to that in communities randomized to this treatment plus a second azithromycin distribution during the dry season.

METHODS

Study design. The Program for the Rapid Elimination of Trachoma (PRET) is a set of three cluster-randomized trials in Niger, Tanzania, and the Gambia.16,17 In the Niger trials, 48 grappes (government health units, termed communities in this report) were randomized into 4 arms of 12 grappes each to evaluate different treatment frequencies and treatment coverage levels (Figure 1). Inclusion criteria for the communities have been described and included total population on the last census between 250 and 600 persons and prevalence of active trachoma (trachomatous inflammation follicular, by using the WHO system)18 ≥10% in children <1–72 months of age.16 We conducted a malaria assessment in 24 communities from 2 of the 4 PRET study arms during May–June 2011.

Intervention. Twelve communities were randomized to receive a single mass distribution of oral azithromycin to all persons ≥6 months of age, and 12 communities were randomized to receive a mass distribution of oral azithromycin in children 6 months to 12 years of age plus a second distribution. The treatment coverage goal was 80% of the targeted group.

The initial distribution of directly observed oral azithromycin (height-based dosing approximating 20 mg/kg19,20) was given to all eligible persons during July 3–August 9, 2010 in the 24 communities. The second mass treatment was provided only to the 12 communities randomized to receive this intervention during January 7–10, 2011. Thus, communities in the twice-treated arm were randomized to an extra mass distribution in the dry, low transmission season. During May 24–June 7, 2011, a cross-sectional survey of 50 randomly chosen children <1–72 months of age was performed in all 24 communities. Because the selection of these 50 children was based on the 12-month census, some of these children may not have been present at the time of azithromycin distributions. If a community contained <50 children, all eligible children were included, but note this is a community-randomized, intent-to-treat analysis of direct and indirect effects of the antibiotic.

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Clinical and laboratory assessments. Thick blood smears and blood spots of approximately 20 μL were prepared on Whatman FTA Elute cards (GE Healthcare, Piscataway, NJ) from a finger prick, after obtaining consent from a parent or guardian. Blood spots were air-dried and stored at room temperature in sealed plastic bags containing desiccant. The thick blood smears were stained with 3% Giemsa and read with a light microscope by two experienced microscopists at the Zinder Hospital in Niger. The microscopists were masked to community treatment assignment and to the readings of the other microscopist. The malaria case definition was based on the presence or absence of *Plasmodium* parasites on microscopy slides. Parasite densities were determined from thick blood smears by counting the number of asexual parasites per 200 leukocytes and assuming a leukocyte count of 8,000/μL. Parasites were considered present if both microscopists observed them. Because prevalence was low, gametocytes were considered present if either microscopist observed them.

To determine *Plasmodium* species, one thick blood smear–positive case was randomly chosen from each of the 24 study communities and tested for *Plasmodium* DNA by polymerase
chain reaction (PCR). A 3-mm punch was prepared from a blood spot (Harris Micro-Punch; Ted Pella, Inc., Redding, CA), and DNA was extracted and eluted in 30 µL of sterile water, according to the FTA card manufacturer’s instructions. Between each sample, the micro-punch was cleaned with bleach, rinsed with ethanol, and dried. A blank punch from a clean FTA card was taken between each sample as a negative control to test for cross-contamination.

For species determination, a portion of the cytochrome b region was amplified by using nested PCR.22 The PCRs were performed in 50-µL reactions using the TopTaq DNA polymerase kit (QIAGEN, Valencia, CA) and 5 µL of the extracted DNA (first round) or the amplicon (nested reaction). The PCR products were confirmed after electrophoresis on a 2% agarose gel. Products from samples positive 

Participants and treatment coverage. Forty-eight communities were enrolled in PRET-Niger, 24 of which were by design included in this trial.16 Twelve communities were randomly assigned to receive a single mass treatment, and 12 were randomized to receive 2 mass treatments, with a goal of 80% antibiotic coverage to the targeted group (Figure 1).

A total of 2,133 children <1–72 months of age were treated at baseline (1,117 in once-treated communities and 1,016 in twice-treated communities), and 1,030 were analyzed at one year (485 in once-treated communities and 545 in twice-treated communities). No communities were missing or lost to follow-up throughout the study. The mean azithromycin coverage of children <1–72 months of age based on the 12-month census was 80.4% (95% CI = 76.8–84.1%) for the single treatment of the once-treated communities, and 74.4% (95% CI = 69.4–79.3%) and 78.7% (95% CI = 74.5–82.7%) for the baseline and six-month treatments, respectively, in the twice-treated communities. The total number of children 6–72 months of age who received treatment based on the 12-month census was 808 at baseline (394 in once-treated communities and 414 in twice-treated communities) and 409 at six months (in twice-treated communities). Baseline and demographic characteristics were comparable in communities randomized to receive one or two treatments (Table 1).

Malarialometric and blood indices. Malaria parasitemia was measured by thick blood smear in children <1–72 months of age, mean months 30.8 (29.8–31.9) 118 (88–147) 72-month-old children per community 124 (42–207) 118 (88–147) 72 months of age based on the

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Once-treated, n = 12 communities</th>
<th>Twice-treated, n = 12 communities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Children per community</td>
<td>124 (42–207)</td>
<td>118 (88–147)</td>
</tr>
<tr>
<td>Age, mean months</td>
<td>30.8 (29.8–31.9)</td>
<td>31.9 (30.7–33.0)</td>
</tr>
<tr>
<td>Proportion female</td>
<td>51.3% (48.2–54.4%)</td>
<td>50.1% (47.1–53.1%)</td>
</tr>
<tr>
<td>Prevalence of clinical trachoma TF†</td>
<td>26.5% (14.9–38.0%)</td>
<td>24.1% (15.9–32.3%)</td>
</tr>
<tr>
<td>Prevalence of clinical trachoma TI†</td>
<td>8.6% (4.4–12.8%)</td>
<td>9.4% (4.9–14.0%)</td>
</tr>
</tbody>
</table>

*Values in parentheses are 95% confidence intervals. †Trachomatous inflammation follicular (TF) and trachomatous inflammation intense (TI) according to a World Health Organization simplified grading system.15
TABLE 2
Malarialometric and blood indices in 24 communities in Niger randomized to one mass azithromycin treatment or one mass azithromycin treatment plus a second mass treatment in the dry, low-transmission season in children ≥ 6 months of age*

<table>
<thead>
<tr>
<th>Measurement</th>
<th>Once-treated, n = 12 communities</th>
<th>Twice-treated, n = 12 communities</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>Malaria parasitemia</td>
<td>29.8% (21.5–40.0%)</td>
<td>19.5% (13.0–26.5%)</td>
<td>0.03</td>
</tr>
<tr>
<td>Parasite density, parasites/µL</td>
<td>354 (117–528)</td>
<td>74 (41–202)</td>
<td>0.03</td>
</tr>
<tr>
<td>Gametocytemia</td>
<td>1.5% (0–6.0%)</td>
<td>0% (0–1.5%)</td>
<td>0.29</td>
</tr>
<tr>
<td>Hemoglobin, g/dL</td>
<td>10.0 (9.8–10.2)</td>
<td>10.2 (10.0–10.4)</td>
<td>0.20</td>
</tr>
</tbody>
</table>

*All values were determined by using the Hodges-Lehmann estimator (pseudomedian). Values in parentheses are 95% confidence intervals.

The community-level Hodges-Lehmann estimators and 95% CIs for all measurement in the two arms — Hemoglobin, g/dL 10.0 (9.8–10.2) — were significantly less likely to have malaria parasitemia observed on thick blood smear (odds ratio 0.95, CI = 0.97–1.00).

DISCUSSION

In this cluster-randomized trial, communities randomized to receive an extra mass distribution of oral azithromycin during the low malaria transmission season had reduced malaria parasitemia and parasite density 4–5 months later than communities randomized to receive only a single distribution. We were unable to demonstrate a significant difference in gametocytemia or hemoglobin concentration associated with the extra mass treatment, although gametocytemia was lower and hemoglobin levels were higher in the twice-treated communities.

Although azithromycin has antimalaria activity, it is not as potent as many standard antimalaria agents and, as with other antibiotics for malaria (e.g., tetracyclines and clindamycin), has a slow action. For treatment of malaria, azithromycin has shown excellent efficacy against P. vivax and modest efficacy against P. falciparum in areas with limited immunity. In two individual-randomized trials, azithromycin demonstrated a range of preventive efficacy of 64–83% against falciparum malaria, but this was less than that of first-line agents. Azithromycin may nonetheless play some role in malaria control because it has a long half-life, is well tolerated, and is approved for use in children and pregnant women.

Furthermore, azithromycin is distributed on such a large scale for trachoma that even a modest effect could be of interest.

The IPTc has been widely advocated for the control of malaria in Africa, and it has shown efficacy when administered during the wet, high-transmission season in areas with highly seasonal malaria transmission. Combination therapies for IPTc such as amodiaquine plus sulfadoxine-pyrimethamine or artesunate plus sulfadoxine-pyrimethamine have been effective in areas of seasonal malaria transmission.

Emerging antimicrobial drug resistance has spread through malaria-endemic areas and has made treatment difficult. We did not measure resistance to azithromycin in Plasmodium because we were unable to perform cultures at our field site and biochemical assays for this have not been adequately described. To the best of our knowledge, resistance to antibiotics has not been demonstrated in malaria except to antifolates.

Nevertheless, surveillance for resistance in malaria is important to track whenever any malaria therapy is used.

Transmission models suggest treatment during the low transmission season of an infectious disease may provide significant or even maximum impact. For malaria, this may be because circulating malaria parasites that are eliminated with mass treatment are not replaced in the absence of transmission. Mass drug administration before the malaria transmission season prevents parasite prevalence levels from recovering to their pretreatment levels, and this raises the probability of parasite elimination in these low transmission settings. Mathematical models of mass treatment of another seasonal disease (trachoma) imply that the optimal time for treatment to achieve elimination may be during the season of lowest transmission. Seasonal variations in transmission might be able to be exploited to maximize the impact of treatment.

This cluster-randomized trial, designed to monitor trachoma, can be applied to secondary questions such as the effect of azithromycin distributions on malaria. Malaria indices were monitored at 12 months into the trial and provided a convenient means of comparing the impact of a single, low-transmission season mass treatment with azithromycin. To account for clustering, we used the prevalence of parasitemia in a community rather than an individual-based analysis. This design necessarily has some important limitations. It did not enable detection of clinical malaria cases or the evaluation of malaria during the subsequent rainy season. No assessment of the prevalence of malaria parasitemia at baseline was conducted. Inclusion of baseline prevalence as a covariate may have offered a more powerful study design or provided evidence that baseline measurements did not explain the
observed association. However, the randomized post-test design does permit valid inference to the treatment assignments are stochastically independent of any other explanatory covariate (including baseline malaria parasitemia). Lack of baseline prevalence also makes it impossible to know whether malaria parasitemia decreased more in the twice-treated arm or simply increased less than in the once-treated arm. The parasitemia outcome was validated by two independent masked graders. However, external monitors were not used and molecular techniques were not used beyond the speciation analysis. In the twice-treated arm, only children received antibiotics, whereas in the once-treated arm, all ages were eligible. It is possible that treating adults had an indirect effect on children, and that this decreased the effect we observed with the extra dose in the twice-treated arm. When the readings of the two microscopists were discordant, we did not have a third adjudicate. For the primary analysis, we assumed positivity if both agreed. It should be noted that concordance was extremely high, and that we obtained similar results in sensitivity analyses using only one grader or assuming positivity if either grader observed parasites.

In summary, mass distribution of azithromycin provided during the dry, low-transmission season in Niger was associated with a reduced community prevalence of malaria parasitemia and parasite density 4–5 months later. Gametocytemia and hemoglobin concentration were not significantly different after an additional dry season administration of azithromycin. Mass administration of azithromycin was associated with reduced mortality in Ethiopia, although it is unclear if this finding was related to protection against malaria. Further studies could address the impact of mass administration of azithromycin on the incidence of clinical episodes of malaria, severe malaria, and other health outcomes.

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